





REVIEW

Biologics and airway remodeling in asthma: early, late, and potential preventive effects

G. Varricchi^{1,2,3,4}  | R. Poto^{1,2,3}  | M. Lommatzsch⁵  | G. Brusselle⁶ | F. Braido⁷ | J. C. Virchow⁵  | G. W. Canonica^{7,8,9}

¹Department of Translational Medical Sciences, University of Naples Federico II, Naples, Italy

²Center for Basic and Clinical Immunology Research (CISI), University of Naples Federico II, Naples, Italy

³World Allergy Organization (WAO) Center of Excellence, Naples, Italy

⁴Institute of Experimental Endocrinology and Oncology (IEOS), National Research Council, Naples, Italy

⁵Department of Pneumology and Critical Care Medicine, University of Rostock, Rostock, Germany

⁶Department of Respiratory Medicine, Ghent University Hospital, Ghent, Belgium

⁷Respiratory Diseases and Allergy Department, IRCCS Polyclinic Hospital San Martino, Genoa, Italy

⁸Department of Biomedical Sciences, Humanitas University, Milan, Italy

⁹Asthma & Allergy Unit-IRCCS Humanitas Research Hospital, Milan, Italy

Correspondence

J. C. Virchow, Departments of Pneumology/Intensive Care Medicine, Universitätsmedizin Rostock, Ernst-Heydemannstrasse 6, 18055 Rostock, Germany.

Email: j.c.virchow@med.uni-rostock.de and jc.h.virchow@sunrise.ch

Abstract

Although airway remodeling in severe and/or fatal asthma is still considered irreversible, its individual components as a cause of clinical symptoms and/or lung function changes remain largely unknown. While inhaled glucocorticoids have not consistently been shown to affect airway remodeling, biologics targeting specific pathways of airway inflammation have been shown to improve lung function, mucus plugging, and airway structural changes that can exceed those seen with glucocorticoids. This superiority of biologic treatment, which cannot be solely explained by insufficient doses or limited durations of glucocorticoid therapies, needs to be further explored. For this field of research, we propose a novel classification of the potential effects of biologics on airway remodeling into three temporal effects: *early effects* (days to weeks, primarily modulating inflammatory processes), *late effects* (months to years, predominantly affecting structural changes), and potential *preventive effects* (outcomes of early treatment with biologics). For the identification of potential *preventive effects* of biologics, we call for studies exploring the impact of early biological treatment on airway remodeling in patients with moderate-to-severe asthma, which should be accompanied by a long-term evaluation of clinical parameters, biomarkers, treatment burden, and socioeconomic implications.

KEYWORDS

asthma treatment, biologics, remodeling

1 | PREMISE

In the context of a recent classification which includes biologics in the treatment of asthma among the disease-modifying anti-asthmatic drugs (DMAADs),¹ this viewpoint examines the

disease-modifying potential of biologics beyond that of achieving remission by controlling symptoms, avoiding exacerbations, and the need for systemic glucocorticoids. Prior to the introduction of biologics, persistent fixed airflow obstruction (FAO) in severe asthma, unresponsive to oral glucocorticoids (OCS), was attributed

Virchow J.C. and Canonica G.W. contributed equally.

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to permanent structural changes in the airways. Although ill-defined in terms of structural and/or pathophysiological alterations, these changes were frequently referred to as “airway remodeling” (AiRem). Its pathobiological relevance in asthma was first described in 1992² and later referred to as “structural consequences of airway inflammation in asthma”,³ underscoring its presumed irreversible nature. Autopsy studies of fatal asthma cases have documented various histological alterations, including epithelial damage, reticular basement membrane (RBM) thickening, goblet cell hyperplasia/metaplasia, mucus plugging, airway smooth muscle (ASM) hyperplasia, altered extracellular matrix composition, and increased inflammatory angiogenesis in both large and small airways.^{4–6} Moreover, airway inflammation is characterized by prominent infiltration of eosinophils, mast cells, and neutrophils.^{7–9} Based on the multifactorial pathogenetic heterogeneity of AiRem, the underlying mechanisms, as well as personalized approaches to target different components of AiRem, remain challenging to address.¹⁰

Clinical observations suggest that AiRem can be differentiated into two major components:

1. An inflammatory component characterized by persistent airway infiltration/activation of immune cells, including eosinophils, mast cells, monocytes/macrophages, basophils, and neutrophils, associated with mucosal oedema and mucus plugs.^{11,12} This component contributes to airway alterations not only due to their proinflammatory effects but also due to steric changes caused by their mere accumulation.¹³
2. A structural component characterized by epithelial and connective tissue changes involving fibroblast/myofibroblast transition, ASM hypertrophy/hyperplasia, RBM thickening, goblet cell hyperplasia/metaplasia, angiogenesis, increased sensory nerve endings, and altered neuronal function.^{12,14–17} The combination of inflammatory and structural changes in AiRem provides the basis of what manifests as FAO in clinical settings.¹²

Currently available biologics which target IgE, interleukin (IL)-5/IL-5R α , IL-4R α , and the alarmin thymic stromal lymphopoietin (TSLP) as well as IL-25 and IL-33 interfere with specific mechanisms relevant to the pathogenesis of both components of AiRem mentioned above. For instance, IgE contributed to AiRem by inducing human ASM cell proliferation¹⁸ and by stimulating the production of several cytokines (e.g., IL-13 and IL-4) and chemokines (e.g., CXCL8) from mast cells.^{19,20} IL-5 induced subepithelial and peribronchial fibrosis by recruiting and activating eosinophils, which are a major source of TGF- β .^{21,22} In a preclinical study, treatment of wild-type mice with neutralizing anti-IL-5 antibody almost completely prevented allergen-induced subepithelial and peribronchial fibrosis.²² Similarly, IL-4 stimulated ASM cells with increased actin and collagen synthesis and TGF- β release by the airway epithelium,^{23,24} IL-13 disrupted epithelial integrity,²⁵ promoted TGF- β release from airway epithelial cells, stimulated airway fibrosis²³ through the action of matrix metalloproteinases (MMPs),²⁶ and enhanced goblet cell metaplasia.^{27–29} In a mouse model of asthma, anti-IL-13 antibody suppressed

airway inflammation and remodeling.³⁰ TSLP promoted AiRem by activating dendritic cells, eosinophils,³¹ mast cells,³² and human lung macrophages.^{33,34} TSLP also activated lung fibroblasts, which produce extracellular matrix (ECM) molecules such as collagen type I and MMP-1.³⁵ TSLP induced the release of vascular endothelial growth factor (VEGF)-A, the most potent angiogenic factor, from human lung macrophages.³⁴ This alarmin also induced epithelial-mesenchymal transition (EMT) in airway epithelial cells.³⁶ In a mouse model of allergic asthma induced by chronic exposure to house dust mites (HDM), neutralization of TSLP inhibited AiRem.³⁷ Additionally, microRNA-19b reduced AiRem by inhibiting Stat3 signaling through TSLP downregulation in a mouse model of asthma.³⁸ Administration of an anti-TSLP mAb decreased airway structural changes by reducing MMP, TGF- β , and connective tissue growth factor in AiRem in ovalbumin-challenged (OVA) mice.³⁹ IL-33 increased the production of collagen and fibronectin-1 in lung fibroblasts contributing to AiRem.^{40,41} In a mouse asthma model, neutralization of the IL-33 pathway improved several aspects of AiRem and lung function.⁴² IL-25 stimulated lung fibroblast proliferation⁴³ and collagen production.⁴⁴ IL-25 contributed to AiRem by inducing airway angiogenesis in mouse asthma models.^{44,45} Finally, the combined blockade of IL-25, IL-33, and TSLP more effectively inhibited AiRem in a mouse model of asthma compared to blocking each cytokine individually.⁴²

2 | AIRWAY INFLAMMATION AND REMODELING

Type (T) 2 inflammation has been postulated to be involved in AiRem development in patients with severe asthma.⁴⁶ The contribution of eosinophilic inflammation to AiRem varies between allergic and eosinophilic or intrinsic asthma, with the latter expressing higher levels of T2-markers¹ and a more difficult-to-treat phenotype.⁴⁷ Activated eosinophils and mast cells induce AiRem by stimulating the proliferation of ASM cells and the release of TGF- β , cationic proteins, and cytokines.^{48,49} Blood eosinophils are associated with airflow obstruction and enhanced lung function decline, independently of asthma or smoking.⁵⁰ Moreover, in a large longitudinal study, blood eosinophils were associated with the risk of developing obstructive lung disease.⁵¹ A recent study identified IL-5R α expression in lower airway fibroblasts from healthy individuals and patients with asthma, suggesting a potential pathogenic role of IL-5 in fibrosis and AiRem. This observation expands the scope of IL-5 biology beyond its effects on eosinophils, suggesting that IL-5 inhibition could offer a therapeutic pathway to prevent, mitigate, or even reverse AiRem.⁵² Accordingly, it could be postulated that any sustained reduction in peripheral blood eosinophil levels, and consequently in the airways, might lead to a reduction in the progressive decline of lung function. Similarly, elevated FeNO levels have been associated with a progressive decline in lung function⁵³; however, it remains unclear to what extent elevated FeNO levels contribute to this decline or whether they are merely a consequence of underlying airway inflammation. Thus, as yet there is only circumstantial evidence that

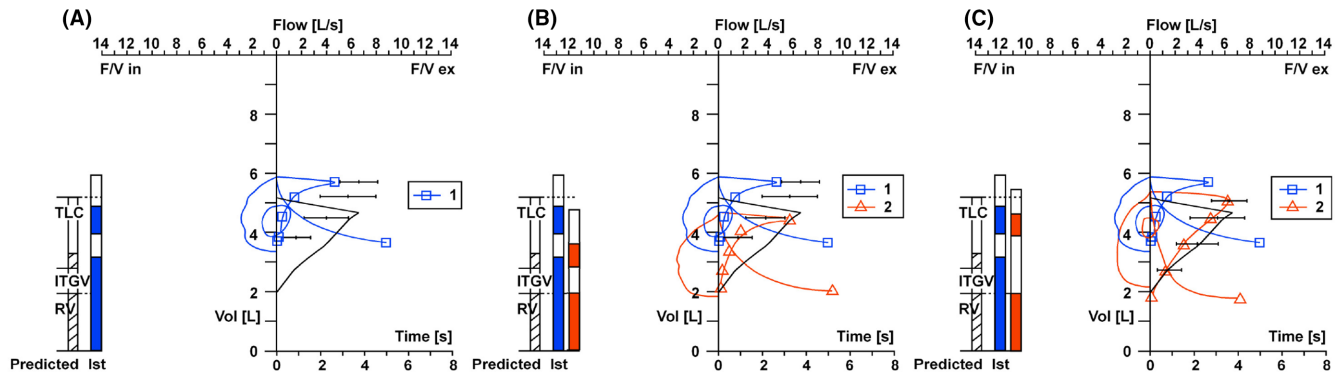


FIGURE 1 (A) FEV₁ (1.3L [49% predicted]) in a patient with severe asthma and repeated severe OCS-dependent exacerbations despite maintenance treatment with oral prednisolone (5 mg/day per os) prior to biologics. (B) The red lines show the FEV₁ improvement (1.6L [61% predicted]) after seven courses of mepolizumab (100 mg s.c./4 weeks) without prednisolone. (C) The red lines show FEV₁ improvement [FEV₁ 2.8L [105% predicted]] after five courses of reslizumab (3 mg/kg IV/4 weeks) without prednisolone. F/V ex, flow/volume during expiration; F/V in, flow/volume during inspiration; ITGV, intrathoracic gas volume; RV, residual volume; TLC, total lung capacity; Vol, volume.

as therapeutic interventions biologics can achieve these outcomes. Therefore, studies such as the ongoing ATLAS trial ([clinicaltrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT05097287) identifier NCT05097287) will be instrumental in evaluating the long-term effects of biologics on preventing or slowing lung function decline. The ATLAS trial will assess whether dupilumab can prevent or slow lung function decline over a 3-year period, focusing on patients with elevated FeNO levels.⁵⁴

The contribution of non-T2 inflammation to AiRem remains controversial.^{55–58} Dendritic cells activated by TSLP drive the differentiation of naïve T-cells into a Th17 phenotype,⁵⁹ which enhances neutrophilic inflammation and fibrotic matrix accumulation that correlate with TGF- β expression.^{60,61} However, in the absence of T2 inflammation, the diagnosis of asthma needs to be questioned.⁶² Accordingly, the pathobiology of what is defined as “T2-low” asthma remains unclear and might include patients in whom asthma is unlikely but also those with spontaneously (i.e., infections) or pharmacologically (i.e., glucocorticoids) suppressed T2-markers.

3 | GLUCOCORTICOIDS AND AIRWAY REMODELING

Although inhaled glucocorticoids (ICS) have been shown to down-regulate some mediators (i.e., tenascin) involved in airway structural changes, other aspects such as RBM thickening, remain unchanged.^{63–66} Intramuscular glucocorticoids do not improve FEV₁ in approximately 80% of patients with severe asthma,⁶⁷ and rescue OCS in patients already treated with high dose ICS/LABA only partly improve lung function.⁶⁸ Finally, pharmacologic treatment might paradoxically contribute to AiRem: β_2 -agonists can increase neurotrophin levels in patients with asthma,⁶⁹ potentially augmenting the autonomous nervous system’s hyperresponsiveness.⁷⁰

In contrast, biologics have been shown to improve airflow obstruction unresponsive to OCS in a substantial percentage of patients with severe asthma.^{71–76} Benralizumab,⁷⁷ mepolizumab,^{78,79} and dupilumab^{71,80} improve lung function and reduce the frequency

of exacerbations despite an average reduction of their OCS dose of approximately 75%.

Accordingly, the definition of FAO should no longer be defined by an unresponsiveness to OCS, but rather on pathobiological assessments. The dichotomy between inflammatory and functional changes refractory to OCS and the seemingly irreversible sequelae of airway structural changes needs to be reevaluated in the biological era. Unfortunately, but understandably from an ethical point of view, there are no direct comparative studies between biologics and OCS. Moreover, the clinical impression that remission achieved by biologics exceeds that achievable with OCS may be influenced by the tapering of OCS dosage during the observation period. This practice might lead to inadequate dosing of OCS, as it represents a compromise between suppressing inflammation and patients’ willingness to adhere to the treatment.

4 | EVIDENCE FOR THE EFFECTS OF BIOLOGICS ON AIRWAY REMODELING

The current hypothesis holds that FAO in asthma, particularly in nonsmokers, is at least partially linked to AiRem driven by persistent airway inflammation.^{81,82} AiRem refers to pathologic alterations of the airway wall that have been documented in patients with different asthma severity and age and involve both large and small airways.⁸² Parenchymal and vascular remodeling may also contribute to FAO and longitudinal decline in FEV₁.⁸¹ Thus, AiRem affects the “whole lung”, with functional and structural changes in the airways and lung parenchyma (e.g., loss of lung tissue elasticity and alveolar damage) leading to FAO.⁸¹

A reduction in the rate of asthma exacerbations has been the primary endpoint in all clinical studies with biologics in severe asthma since the INNOVATE trial for omalizumab.⁸³ Clinical experience as well as real-life studies suggest that biologics can normalize airway patency in a substantial percentage of patients, even if FEV₁ impairment was poorly or unresponsive to inhaled bronchodilators, ICS,

and OCS. In the past, these patients' airflow obstruction had been labeled as "fixed" or "nonresponsive" even to OCS and was, therefore, allegedly due to irreversible structural changes. Figure 1 exemplifies the normalization of lung function after biologics treatment in a patient with severe eosinophilic asthma, challenging the notion of irreversibility and highlighting the disease-modifying potential of biologics.

These clinical observations have never been formally analyzed in randomized clinical trials (RCT). However, the term "super-responder" has been introduced⁸⁴ to define patients treated with biologics who achieve no further exacerbations. This definition, which varies across the studies discussed below, does not address the newer and overarching concept of remission, which refers to few or no symptoms, no requirement for OCS, absence of asthma exacerbations, and stable lung function for a period of 12 months.¹

Indirect evidence suggests that lung function impairment unresponsive to glucocorticoids, and therefore considered "irreversible", can improve with biologics.⁸⁵ The latter study defined "super-responders" as patients without exacerbations, OCS withdrawal, and few or no asthma symptoms (ACT score ≥ 20 and ACQ score < 1.5). Accordingly, 47% of patients on benralizumab, 58% on mepolizumab, and 61% on omalizumab achieved such improvements in FEV₁.⁸⁵

Stopping omalizumab after long-term treatment led to a recurrence of asthma exacerbation in 52.3% of patients within a year, while asthma control was maintained in 47.7% of patients.⁸⁶ Obviously, this circumstantial evidence is only suggestive of possible anti-remodeling effects but needs further investigation. Similarly, in patients with severe asthma whose treatment with mepolizumab was discontinued after 3 years, worsening of lung function and exacerbations occurred earlier and more frequently compared to those who had not discontinued mepolizumab. However, a substantial proportion of the latter remained in remission, suggesting long-term benefits which could be attributed not only to anti-inflammatory but also to anti-remodeling effects.⁸⁷ Characteristics and biomarkers of "super-responders" to biologics have not been identified but should become a priority of future prospective studies.

In a real-life study, patients treated with mepolizumab, who discontinued OCS without exacerbations, did not show any FEV₁ improvements,⁸⁸ while in a similar study, patients on benralizumab showed a significant increase in FEV₁ despite a reduction in OCS.⁸⁹ Notably, "super-responders" to benralizumab had lower baseline FEV₁ values but higher doses of OCS compared to responders, supporting the assumption that airflow obstruction responded to benralizumab better than to OCS. In another observational study conducted in patients with frequent exacerbations (median 3 OCS courses/year) and a mean FEV₁ of 57% predicted, mepolizumab treatment increased FEV₁ despite a reduction in OCS.⁹⁰

"Super-responders", defined as patients with no residual disease manifestation after a 2-year treatment with anti-IL-5/IL-5R α antibodies (mepolizumab, reslizumab, and benralizumab), had a greater improvement in FEV₁ than those with partial or no response, despite a complete withdrawal of OCS.⁹¹ In a previous RCT conducted in

patients on maintenance OCS, mepolizumab improved FEV₁ despite a mean reduction in OCS of 50%.⁷⁹ Similarly, benralizumab showed an increase in FEV₁ after 20 weeks of treatment despite a 75% reduction in maintenance OCS dose.⁸⁹ Moreover, it has been shown that lung function impairment poorly responsive to β_2 -agonists in severe asthma might predict a better response to reslizumab.⁹²

The addition of the IL-4R α blocker dupilumab to maintenance OCS led to a sustained FEV₁ improvement despite an average 70% reduction in the maintenance dose of OCS.⁹³ These findings suggest that this biologic is superior to conventional asthma treatment, including OCS, to maintain airway patency. Rapid lung function improvement has also been described in patients with uncontrolled, moderate-to-severe asthma treated with dupilumab.⁹⁴

Tezepelumab, which blocks the activity of TSLP,⁹⁵ rapidly improved lung function, reduced airway submucosal eosinophils, and biomarkers of AiRem (i.e., matrix metalloproteinase (MMP)-10 and MMP-3) in a broad population of patients with severe, uncontrolled asthma.^{96,97} Lung function parameters improved at week two and were sustained over 52 weeks.⁹⁶

Mucus plugging which has been described as localized and persistent⁹⁸ significantly contributes to symptom severity and exacerbations in asthma, correlating with airway obstruction and T2 inflammation. Biologics have shown effectiveness in reducing mucus plugs and improving airway obstruction in severe asthma.^{99,100} In particular, benralizumab, dupilumab, mepolizumab, and most recently tezepelumab have been associated with a reduction in mucus plugs in the airways of patients with severe asthma.¹⁰¹⁻¹⁰⁵ Benralizumab significantly reduced the number of mucus plugs, which were associated with sputum eosinophils.¹⁰³ Dupilumab reversed airway mucus hypersecretion, as evidenced by reduced mucus score, and imaging aspects of AiRem (e.g., airway wall area) in patients with moderate-to-severe asthma.¹⁰⁶ Tezepelumab has been shown to reduce mucus plug scores and improve lung function in adults with moderate-to-severe asthma.¹⁰⁴

In recent years, high-resolution computed tomography (HRCT) and magnetic resonance imaging (MRI) have emerged as noninvasive techniques to examine various aspects of AiRem.¹⁰⁷ Two studies reported that omalizumab reduced airway wall thickness¹⁰⁸ and airway wall area corrected for body surface¹⁰⁹ assessed by HRCT. In two studies conducted^{110,111} on AiRem using HRCT, it was found that mepolizumab treatment resulted in a greater change in pre-/post-treatment luminal area compared to the placebo group. In addition, mepolizumab reduced bronchial wall thickness, as evaluated by endobronchial ultrasound (EBUS).¹¹² In a phase 2 study, tezepelumab improved the CT scan-determined lumen area across airway generations.⁹⁷ In uncontrolled eosinophilic asthma, early ventilation defect responses at 4 weeks after benralizumab persisted at 2.5 years, along with marked improvements in mucus scores.¹¹³ Future studies, such as the BURAN study (NCT05552508) for benralizumab and FUNLUM study (NCT04512521) for mepolizumab, are needed to evaluate the effects of these biologics on airway structure and dynamics through functional respiratory imaging (FRI), a new technique for detailed 3D lung analysis.¹¹⁴ Overall, studies using CT and

MRI in asthma have the potential to noninvasively address some structural and functional changes in airways and pulmonary vessels associated with disease severity. These techniques will be useful in assessing the effects of biologics on AiRem, and possibly differentiating between reversible and potentially irreversible changes.

5 | PROPOSAL FOR A CLASSIFICATION OF THE POTENTIAL EFFECTS OF BIOLOGICS ON AIRWAY REMODELING

There is increasing evidence that biologics can improve lung function, inflammatory, and/or structural alterations, “traditionally” referred to as AiRem.^{12,46} The underlying mechanisms and specific predictors of the biologics' effects on various aspects of AiRem are still largely unknown. Future investigations, including studies using novel technologies (e.g., single-cell mRNA sequencing of airway biopsies), should investigate the effects of different biologics on distinct components (inflammatory/immune and structural) of AiRem in patients with asthma. In proposing a classification of the potential effects of biologics on AiRem, we categorize these effects into early, late, and preventive. It's imperative to clarify that while early and late effects are supported by emerging evidence showcasing biologics' impact on lung function and on inflammatory and structural changes, the “preventive effects” represent a forward-looking hypothesis. This distinction aims to underscore the nascent yet promising area of preventive potential, which, despite its current lack of direct evidence, encourages a pivotal direction for future research.

Figure 2 schematically illustrates the mechanisms of action of different biologics and their immunological and cellular targets in the context of airway remodeling. The red lines represent the potential mechanisms responsible for early effects, while the blue lines for late effects.

Figure 3 Proposed classification of the early, late, and potential preventive effects of biologics on airway remodeling in asthma.

5.1 | Early effects of biologics

Early effects of biologics on inflammatory events of AiRem are observed within days or weeks of starting treatment. Dupilumab has a fast onset of action on different T2 inflammatory disorders, including severe asthma, leading to FEV₁ improvements within 2 weeks after the first administration, maintained throughout the treatment.^{71,121,124,144} Similar rapid effects occur in patients with chronic rhinosinusitis with nasal polyps (CRSwNP), where dupilumab reduced polyps' size (NPS-Nasal Polyp Score) within 2 weeks.¹⁴⁵ This was also confirmed for benralizumab in a real-life study.¹⁴⁶

FEV₁ improvements after a single administration of benralizumab have been observed within 28 days,^{125,147,148} possibly also due in part to a reduction in peripheral airway mucus plugs. A significant change in mean predicted FEV₁ and FEV₁% was observed after 1 month of mepolizumab treatment.¹²² Moreover, mepolizumab improved

small airway function after the first administration.¹⁴⁹ A post hoc analysis demonstrated lung function improvement in patients with moderate-to-severe allergic asthma treated with omalizumab as early as 16 weeks.¹²³ Tezepelumab demonstrated significant FEV₁ improvements as early as week 2 and was sustained for the duration of the trial.¹⁵⁰ Itepekimab, which blocks IL-33 activity, has been shown to improve pre-bronchodilator FEV₁ as early as 4 weeks of treatment.¹²⁶ Additionally, tozorakimab is a novel mAb that inhibits IL-33 activity via both the ST2 and the receptor for advanced glycation end products–epidermal growth factor receptor complex signaling pathways. Although its effects on AiRem in asthma have not been studied, the broader context of IL-33 inhibition suggests potential improvements in lung function and certain features of AiRem. Figure 3 schematically illustrates a proposed classification of the early, late, and potential preventive effects of biologics on airway remodeling in asthma.

5.2 | Late effects of biologics

The evidence for a therapeutic effect of biologics on structural changes in asthma that revert after prolonged treatment is still circumstantial and requires further investigation. In analogy, atopic dermatitis, which causes hyperplastic epidermis, prominent hyperkeratosis, minimal spongiosis, and infiltration of immune cells (i.e., dendritic, Th1, Th2, and pro-B cells) can respond to dupilumab with almost complete restoration of previously damaged and scarred skin lesions.^{136,137} This reversal of skin structural changes induced by a biologic in severe atopic dermatitis was neither anticipated nor observed with any previous treatment, especially not with systemic or topical glucocorticoids. Accordingly, these effects have been attributed to direct or indirect effects of dupilumab on skin remodeling.^{136,137} Similarly, in eosinophilic esophagitis (EoE), dupilumab has been shown to improve aspects related to structural remodeling, as evidenced by the reduction in peak oesophageal intraepithelial eosinophil count, Endoscopic Reference Score (EREFs), as well as Histologic Scoring System (HSS) versus placebo.¹³⁸ There is increasing evidence that similar effects might occur in asthma. Late effects of biologics might modify some of the structural changes of AiRem which occur in severe asthma, with these changes becoming evident over the course of several months or even years of therapy.¹² Initial studies of the biologics' effects on AiRem were performed in vivo on bronchial biopsies.⁶³ Although such invasive investigations are not part of any routine assessment of the effects of anti-asthmatic treatments, noninvasive imaging techniques (HRCT and nuclear magnetic resonance [NMR]) might in the future be useful to indirectly document the effects of biologics on AiRem in asthma.^{81,93,151} Accordingly, it has been reported that omalizumab reduced airway wall thickening and sputum eosinophil numbers in patients with asthma.¹⁰⁸ Similar effects on RBM thickening were reported in biopsies of patients with severe asthma.¹⁵² Moreover, omalizumab reduced basal lamina thickness and fibronectin deposits in the bronchial mucosa of severe allergic asthmatics.¹²⁰ In vitro

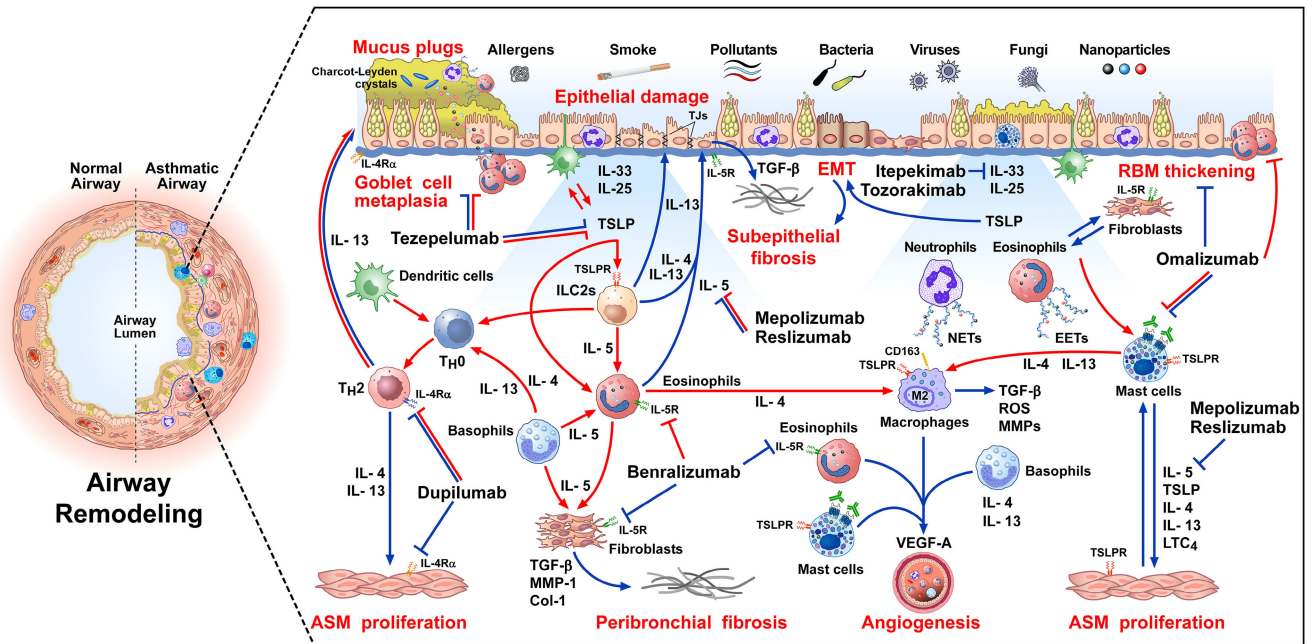


FIGURE 2 The figure schematically illustrates the complex mechanisms involved in airway remodeling (AiRem) in asthma. The figure also highlights the effects of various biological therapies in modulating these processes. The left side of the figure shows a normal airway compared to an asthmatic airway, which is characterized by inflammatory and structural changes such as epithelial damage, goblet cell metaplasia, mucus plugging, reticular basement thickening (RBM), airway smooth muscle (ASM) proliferation, subepithelial and peribronchial fibrosis, and increased angiogenesis. Several environmental factors (e.g., allergens, smoke, pollutants, microbial compounds, and nanoparticles) cause epithelial damage, leading to the release of alarmins (i.e., TSLP, IL-33, and IL-25). These cytokines activate a broad spectrum of innate (e.g., ILC2s) and adaptive (e.g., Th2 cells) immune cells, which promote airway inflammation and remodeling through the production of IL-4, IL-5, and IL-13.¹¹⁵ TSLP plays a central role in AiRem by activating dendritic cells, eosinophils,³¹ mast cells,³² and human lung macrophages.^{33,34} TSLP also stimulates lung fibroblasts to produce extracellular matrix (ECM) components, promoting peribronchial fibrosis.³⁵ In addition, TSLP induces the release of vascular endothelial growth factor (VEGF)-A, the most potent angiogenic factor, from lung macrophages,³⁴ and triggers epithelial-mesenchymal transition (EMT).³⁶ Tezepelumab, a human monoclonal antibody (mAb) that blocks TSLP, interferes with the activation of upstream and downstream inflammatory pathways, which contribute to goblet cell hyperplasia, mucus plugging, EMT, and angiogenesis. Dupilumab, an IL-4R α mAb antagonist, blocks both IL-4 and IL-13 signaling. IL-4 promotes ASM proliferation by increasing Actin and collagen synthesis and enhancing TGF- β release from the airway epithelium.^{23,24} IL-13 disrupts epithelial integrity²⁵ and promotes TGF- β release from airway epithelial cells, stimulating airway fibrosis²⁶ through the action of matrix metalloproteinases (MMPs). IL-13 also enhances goblet cell metaplasia, contributing to mucus production and airway obstruction.^{23,27,29} Dupilumab reduces mucus hypersecretion and goblet cell metaplasia¹⁰⁶ and modulates ASM responses.¹¹⁶ Mepolizumab and reslizumab block IL-5, which plays a critical role in subepithelial and peribronchial fibrosis by recruiting and activating eosinophils, which are major sources of TGF- β .^{21,22} Mepolizumab reduces ASM proliferation and prevents IL-5-mediated epithelial damage and fibrosis.^{117,118} Benralizumab, by targeting IL-5R α , induces eosinophil depletion, thereby mitigating eosinophil-driven AiRem. Benralizumab also reduces mucus plugging¹⁰³ and ASM proliferation.¹¹⁹ Omalizumab, which targets IgE, reduces RBM thickening and fibronectin deposits in the bronchial mucosa.¹²⁰ Itepekimab and Tozorakimab, interfering with IL-33 signaling, may reduce AiRem by inhibiting TGF- β pathways, although their role in clinical practice is still being evaluated. IL-33 increases the production of collagen and fibronectin-1 in lung fibroblasts, contributing to subepithelial and peribronchial fibrosis.⁴⁰ Red lines indicate inflammatory mechanisms targeted by biologics in early intervention, while blue lines represent the modulation of structural changes in the late phase of treatment. ASM, airway smooth muscle; Col-1, collagen-1; EMT, epithelial-mesenchymal transition; IL, interleukin; IL-5R, interleukin-5 receptor; ILC2, innate lymphoid cell type 2; LTC₄, leukotriene C₄; MMP, matrix metalloproteinase; RBM, reticular basement thickness; ROS, reactive oxygen species; TGF, transforming growth factor; T_H, T helper lymphocyte; TSLP, thymic stromal lymphopoietin; TSLPR, thymic stromal lymphopoietin receptor.

incubation of ASM cells with omalizumab prevented mesenchymal cell proliferation and accumulation of collagen and fibronectin induced by IgE-containing serum from asthmatic patients.¹³⁹ The latter findings might explain some of the *in vivo* effects of omalizumab on AiRem.

Examination of bronchial biopsies of patients with mild asthma before and after mepolizumab¹¹⁷ showed a reduction in bronchial eosinophil numbers, airway TGF- β 1⁺ eosinophils, RBM thickness,

and tenascin immunoreactivity as well as TGF- β 1 concentration in bronchoalveolar lavage fluid. In the MESILICO study, mepolizumab treatment was associated with a significant reduction (mean 27%) in ASM area and improved FEV₁. These findings suggest that changes associated with structural remodeling might indeed be reversible.^{12,117,118}

Benralizumab reduces eosinophil infiltration in the airway mucosa/submucosa and sputum.¹⁵³ Moreover, benralizumab leads to

Effects of Biologics on Airway Remodeling

Early effects

On inflammatory changes unresponsive to glucocorticoids

FEV₁^a
 Airway eosinophil infiltration^b
 Eosinophil cationic protein deposition^c
 Small airway obstruction^d
 Mucus plugging^e
 T-cell infiltration^f
 Neurotrophin release^g
 IL-13 / IL-4 dependent T-cell activation^h
 Cytokine release / action (IL-5, IL-4 / 13)ⁱ
 IgE-mediated inhibition of IFN- γ release by pDCs^j

Late effects

On anatomical / histological / functional changes

FEV₁^k
 Airway eosinophil infiltration^l
 Myofibroblast activation^m
 RBM thickeningⁿ
 ASM hypertrophy / hyperplasia^o
 ECM deposition^p
 Tenascin expression^q
 Neuronal remodeling^r

Preventive effects

(To be demonstrated)

Progression to chronic asthma^s
 Development of secondary structural changes^t
 Progressive loss of pulmonary function^u
 Requirement for chronic long-term treatment^v

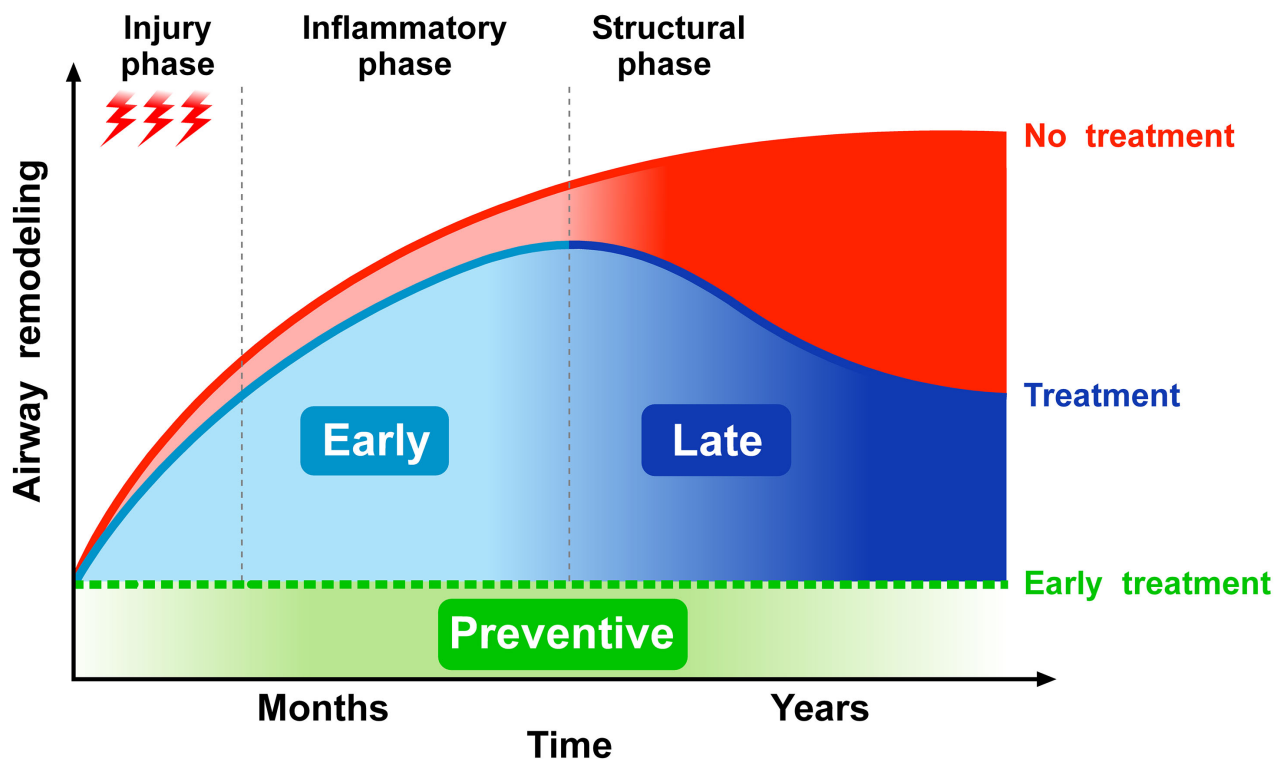


FIGURE 3 Early effects: Evidence of improvements in lung function and reduction in inflammation shortly after biologics' initiation illustrate the rapid benefits of targeting specific inflammatory pathways (a^{94,95,113,121-126}, b¹²⁷, c¹²⁸, d¹²⁹, e¹³⁰, f¹²⁷, g¹³¹, h^{132,133}, i¹³⁴, j¹³⁵). Late effects: Analogous to observed outcomes in conditions like atopic dermatitis^{136,137} and eosinophilic oesophagitis,¹³⁸ biologics may also impact long-term structural changes within the airways, suggesting a potential for reversing or ameliorating established AiRem (k^{79,85,89-91,93,96}; l¹²⁷; m⁵²; n^{117,120}; o^{118,119,139}; p¹¹⁷; q¹¹⁷; r¹⁴⁰). Potential preventive effects: This represents an emerging research domain, emphasizing the potential of biologics to prevent the onset or progression of AiRem. Although empirical evidence is presently limited in this area, it is identified as a vital avenue for future research aimed at elucidating the comprehensive effects of biologics on the asthma disease trajectory (s¹⁴¹, t¹², u¹⁴², v¹⁴³). It is important to note that the studies regarding the early and late effects of biologics are essentially based on adults with severe asthma. Further research is needed to evaluate early, late, and potential preventive effects of biologics in pediatric cohorts. ASM, airway smooth muscle; ECM, collagen and extracellular matrix; FEV₁, forced expiratory volume in 1 s; IFN, interferon; IL, interleukin; pDC, plasmacytoid dendritic cell; RBM, reticular basement membrane.

a reduction in ASM mass in severe eosinophilic asthma patients.¹¹⁹ Indices of airway caliber (R5 and R20) measured by oscillometry improved after 4 weeks of treatment with benralizumab despite

unchanged spirometry values, suggesting that oscillometric parameters may be more sensitive to detect changes in the small airways than conventional spirometry.¹⁵⁴

Further evidence for the effects of biologics on lung function and airway structural changes has been provided through functional respiratory imaging. In the VESTIGE study (NCT04400318), dupilumab reduced airway inflammation and mucus plugging, which led to improved airway volume and flow.¹⁵⁵ In addition, dupilumab improved ventilation heterogeneity measured by magnetic resonance imaging (MRI), suggesting its role in modulating different aspects of airway obstruction.¹⁵⁶ Blocking of IL-4R α has also been shown to improve CT-derived measures of pulmonary vascular volume.¹⁵⁷ In a placebo-controlled RCT, it was demonstrated that dupilumab reduces CT biomarkers of airway mucus, gas trapping, and remodeling, thereby improving ventilation as assessed by MRI and measures of small airway function via oscillometry in patients with severe asthma.¹⁵⁸ In a mouse model, blocking IL-4R α was found to inhibit the entry of eosinophils into lung tissue by reducing chemotaxis and endothelial activation. Moreover, IL-4R α blockade prevents house dust mite (HDM)-induced epithelial barrier dysfunction, tissue remodeling, and lung function decline.¹³³

In the phase 2 CASCADE study performed in moderate-to-severe asthma patients, tezepelumab increased CT scan-determined lumen area across airway generations.⁹⁷ In addition, there was a numerical but statistically nonsignificant improvement in RBM thickening, epithelial damage, and denudation. Future studies evaluating the “late effects” of biologics on AiRem should be performed at later time points in the course of treatment and/or in larger cohorts of patients with different asthma phenotypes. Moreover, there is a need to study the late effects of each biologic on AiRem. The results arising from these studies will contribute to our understanding of the underlying mechanisms of different aspects of AiRem. These findings will form the basis for even better targeted, personalized therapies of severe asthma.

5.3 | Potential preventive effects of biologics

Preventive effects of biologics on AiRem represent a promising yet unexplored avenue in asthma management. There is growing evidence suggesting that modulating early inflammatory cascades with biologics could limit the progression of airway fibrosis.¹² Biopsy studies conducted in young children suggest that AiRem might be an early event in asthma development, preceding the onset of symptoms.^{159–162} However, clinical studies on the preventive effects of biologics in asthma and/or AiRem development are lacking, particularly in the pediatric population. Early intervention with biologics may be beneficial in altering the disease trajectory from its inception, not only in pediatric populations but also in adults. Pediatric and adult studies targeting early intervention could provide valuable insights into the long-term outcomes of asthma management. Furthermore, upstream inhibition of inflammatory cascades (e.g., anti-alarmins) but also direct inhibition of specific immunological/inflammatory pathways (IgE, cytokines, and immune cells) can interfere with the progression of airway fibrosis.^{21,22,27,28} Based on these findings, it can be reasonably hypothesized that early intervention with biologics blocking these initial events may be useful in preventing asthma or the progression of milder forms into more severe diseases. Moreover, such early intervention could potentially prevent both functional as well as structural aspects of AiRem (Figure 3).

This would include the prevention of cytotoxic effects of eosinophil-derived mediators leading to fibrosis (e.g., eosinophilic endomyocardial fibrosis),^{163,164} but also the steric effects of eosinophil accumulation in airway walls, including mucus plugging. Similarly, the use of anti-IL-5 mAbs for eosinophilic diseases¹⁶⁵ possibly suggests these biologics as first-line treatment in eosinophilic inflammatory diseases, including asthma, as recently suggested.¹⁶⁶

TABLE 1 Outstanding pathophysiological and therapeutic questions.

Research domain	Outstanding questions	Clinical and research implications
Early intervention with biologics	At which disease stage is intervention most effective in modifying disease progression?	Identifying critical windows and patient profiles for early biologic therapy to prevent or reverse AiRem.
Long-term and preventive effects	Can biologics prevent the progression to chronic asthma and halt secondary structural changes?	Evaluating the potential disease-modifying effects of biologics by preventing structural changes and minimizing long-term treatment needs.
Pediatric asthma and airem	How does early biologic treatment impact the development and progression of AiRem in pediatric asthma?	Assessing safety, efficacy, and long-term outcomes of biologic treatments in children to guide early intervention strategies.
Stratification for airem prevention	Which biomarkers or clinical indicators can predict the development of AiRem and response to biologic therapy?	Developing biomarkers and predictive models for early identification of patients at risk of AiRem who may benefit from targeted biologic interventions.
Biologics vs. conventional therapies	How do biologics compare to conventional treatments in preventing or reversing AiRem?	Direct comparison studies to clarify the superiority of biologics in modifying disease progression and structural changes.
T2 and non-T2 asthma and airem	What is the impact of biologics on AiRem across T2-high and non-T2 asthma phenotypes?	Expanding research to understand the therapeutic potential of biologics on AiRem in various asthma phenotypes, including non-T2 inflammation.
Emerging targets for airem	What novel targets for biologic therapy have the potential to address AiRem more effectively?	Investigating new biologic agents and their mechanisms of action with the potential to specifically target and alter AiRem processes.

TABLE 2 Ongoing clinical trials evaluating the reversing effects of biologics on airway remodeling in asthma.

Biologic	Clinical trial identifier	Phase	Population	Primary outcomes	Duration	Status
Dupilumab	NCT04400318 (VESTIGE)	Phase 4	Adults with severe asthma	Effect of dupilumab on lung inflammation and related changes in airway volumes detectable by functional respiratory imaging	Minimum 29 weeks and up to 41 weeks	Completed
Mepolizumab	NCT04612556 (MESILICO)	Phase 4	Adults with severe asthma	Any improvement in airway remodeling after 156 weeks of treatment	3 years	Active, not recruiting
Mepolizumab	NCT03797404 (REMOMEPO)	Phase 4	Adults with severe asthma	Changes in RBM thickening, in ASM area after 6 months and 12 months of mepolizumab treatment	12 months	Completed
Mepolizumab	NCT05708300 (CALIOP)	Phase 4	Adults with severe asthma and CRSwNPs	Evaluation of the airway remodeling in various anatomic locations of the airways (nose, bronchus) at baseline-before treatment initiation and after 156 weeks of mepolizumab treatment.	3 years	Recruiting
Benralizumab	NCT04365205 (BENRAMOD)	Phase 4	Adults with severe asthma	Number of bronchial smooth muscle (BSM) cells obtained from severe asthmatics measured in vitro using BrdU incorporation.	12 months	Recruiting
Benralizumab	NCT03953300 (CHINOOK)	Phase 4	Adults with severe asthma	The change in eosinophil numbers expressed as number/mm ² in submucosa as measured by major basic protein (MBP) staining in endobronchial biopsies	1 year	Recruiting
Benralizumab	NCT06288516 (BREATH)	Phase 4	Adults with severe asthma	Any improvement in airway remodeling after 52 weeks of treatment	12 months	Recruiting
Tezepelumab	NCT05651841 (REVERT)	Phase 4	Adults with severe asthma	Comparison on CT-scan in the change in mean percentage bronchial wall area (%WA) at the B1 and B8 bronchi, generations 3, 4, and 5	12 months	Recruiting

It is critical to recognize that not all individuals with mild or moderate asthma will develop severe asthma or AiRem. Therefore, studies on cohorts such as The Mild/Moderate Asthma Network in Italy (MANI) are needed to understand individual patients' trajectories and their risk factors.¹⁶⁷ Furthermore, the use of canonical (e.g., blood eosinophil counts and FeNO levels) and the identification of novel biomarkers predictive of severe asthma could play a critical role in optimizing the use of biologics in preventive strategies. These efforts could help stratify patients at risk of severe asthma and/or AiRem, guiding early biologic interventions more efficiently.

Based on the previous observations, existing biologics as well as those in development, offer a challenging opportunity to also study early treatment (GINA Step 3 or even 2) of asthma with biologics. These strategies could prevent or limit not only the side effects of OCS but also the structural changes resulting from longstanding uncontrolled airway inflammation. Future studies should aim to prevent the onset of AiRem, taking into account the cost-effectiveness of early treatment with biologics, potentially at lower doses or different schedules. The inclusion of predictive modeling and risk stratification, as pursued by MANI,¹⁶⁷ could guide the judicious selection of patients for these preventive interventions.

6 | CONCLUSION

Our proposal concerning the effects of biologics on AiRem aligns with the evolving perspective of considering biologics as disease-modifying anti-asthmatic drugs.¹ Table 1 outlines key unanswered questions about biologics and AiRem in asthma, emphasizing the need for further research to clarify their impact on AiRem. This research is crucial for identifying the potential preventive effects of biologics and involves exploring the impact of early biological treatment on AiRem. We advocate for studies leveraging cutting-edge technologies and imaging techniques to investigate the role of biologics in mitigating AiRem among patients with moderate-to-severe asthma. Currently, several clinical trials are evaluating the REVERSING effect of biologics on AiRem. These include dupilumab (NCT04400318), mepolizumab (NCT04612556, NCT03797404, and NCT05708300), benralizumab (NCT04365205, NCT03953300, and NCT06288516), and tezepelumab (NCT05651841) (Table 2). Future research should include both short- and long-term evaluations of clinical outcomes, biomarkers, treatment burden, and socioeconomic implications.

AUTHOR CONTRIBUTIONS

GV, GWC, and JCV wrote the first draft of the manuscript. GV, RP, ML, GB, FB, GWC, and JCV reviewed and modified the manuscript. All authors contributed to the refinement of the final version and agreed to the decision to submit for publication.

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CONFLICT OF INTEREST STATEMENT

GV reports research support from AstraZeneca. RP has no potential conflicts of interest to declare. ML has received consulting fees or honoraria, or both for lectures from ALK, Allergopharma, AstraZeneca, Berlin-Chemie, Boehringer Ingelheim, Chiesi, GSK, HAL Allergy, Leti, Novartis, MSD, Sanofi, and TEVA; and grants for research or clinical trials, or both from Deutsche Forschungsgemeinschaft, AstraZeneca, and GSK. GB reports personal fees from AstraZeneca, personal fees from Boehringer-Ingelheim, personal fees from Chiesi, personal fees from GSK, personal fees from Novartis, personal fees from Sanofi, grants from MSD, outside the submitted work. G.W.C. has received consulting fees or honoraria, or both for lectures from AstraZeneca, Chiesi, Novartis, Sanofi, Menarini, Stallergenes Greer, GSK, and HAL Allergy. FB has been on the scientific Board for AZ, BI, Chiesi, GSK, Menarini group, Sanofi and P&G; received honoraria from AZ, BI, Chiesi, GSK, Menarini group and Sanofi. JCV is a full time employee of the University of Rostock as a full time professor and chair of the Departments of Pneumology and Intensive Care Medicine has given independent advice, lectured for and received honoraria from AstraZeneca, Avontec, Bayer, Bencard, Bionorica, Boehringer-Ingelheim, Chiesi, Essex/Schering-Plough, GSK, Janssen-Cilag, Leti, MEDA, Merck, MSD, Mundipharma, Novartis, Nycomed/Altana, Pfizer, Revotar, Sandoz-Hexal, Stallergenes, TEVA, UCB/Schwarz-Pharma, Zydus/Cadila, has participated in advisory boards for Avontec, Boehringer-Ingelheim, Chiesi, Essex/Schering-Plough, GSK, Janssen-Cilag, MEDA, MSD, Mundipharma, Novartis, Regeneron, Revotar, Roche, Sanofi-Aventis, Sandoz-Hexal, TEVA, UCB/Schwarz-Pharma and has received research grants from the Deutsche Forschungsgesellschaft, Land Mecklenburg-Vorpommern, GSK, MSD. G.W.C. received honoraria for lectures, presentations, speakers from AstraZeneca, GSK, Novartis, Sanofi, Stallergenes, Greer, Hal Allergy, Menarini, Chiesi, Mylan, Valeas, Faes.

DATA AVAILABILITY STATEMENT

Data sharing not applicable to this article as no datasets were generated or analysed during the current study.

ORCID

G. Varricchi  <https://orcid.org/0000-0002-9285-4657>

R. Poto  <https://orcid.org/0000-0002-4723-0167>

M. Lommatzsch  <https://orcid.org/0000-0002-9305-9348>

J. C. Virchow  <https://orcid.org/0000-0003-4291-1956>

REFERENCES

1. Lommatzsch M, Brusselle GG, Canonica GW, et al. Disease-modifying anti-asthmatic drugs. *Lancet*. 2022;399:1664-1668. doi:10.1016/S0140-6736(22)00331-2
2. Bousquet J, Chanez P, Lacoste JY, et al. Asthma: a disease remodeling the airways. *Allergy*. 1992;47:3-11. doi:10.1111/j.1398-9995.1992.tb02242.x
3. Vignola AM, Chanez P, Bonsignore G, Godard P, Bousquet J. Structural consequences of airway inflammation in asthma.

- J Allergy Clin Immunol.* 2000;105:S514-S517. doi:[10.1016/s0091-6749\(00\)90053-8](https://doi.org/10.1016/s0091-6749(00)90053-8)
4. Green FH, Williams DJ, James A, et al. Increased myoepithelial cells of bronchial submucosal glands in fatal asthma. *Thorax.* 2010;65:32-38. doi:[10.1136/thx.2008.111435](https://doi.org/10.1136/thx.2008.111435)
 5. James AL, Elliot JG, Jones RL, et al. Airway smooth muscle hypertrophy and hyperplasia in asthma. *Am J Respir Crit Care Med.* 2012;185:1058-1064. doi:[10.1164/rccm.201110-1849OC](https://doi.org/10.1164/rccm.201110-1849OC)
 6. Mauad T, Ferreira DS. Asthma as a cause of death. *Pulmão RJ.* 2012;21(2):18-24.
 7. Mauad T, Ferreira DS, Costa MB, et al. Characterization of autopsy-proven fatal asthma patients in Sao Paulo, Brazil. *Rev Panam Salud Publica.* 2008;23:418-423. doi:[10.1590/s1020-49892008000600007](https://doi.org/10.1590/s1020-49892008000600007)
 8. de Magalhaes Simoes S, dos Santos MA, da Silva Oliveira M, et al. Inflammatory cell mapping of the respiratory tract in fatal asthma. *Clin Exp Allergy.* 2005;35:602-611. doi:[10.1111/j.1365-2222.2005.02235.x](https://doi.org/10.1111/j.1365-2222.2005.02235.x)
 9. Dolhnikoff M, da Silva LF, de Araujo BB, et al. The outer wall of small airways is a major site of remodeling in fatal asthma. *J Allergy Clin Immunol.* 2009;123:1090-1097. doi:[10.1016/j.jaci.2009.02.032](https://doi.org/10.1016/j.jaci.2009.02.032)
 10. James AL, Donovan GM, Green FHY, et al. Heterogeneity of airway smooth muscle remodeling in asthma. *Am J Respir Crit Care Med.* 2023;207:452-460. doi:[10.1164/rccm.202111-2634OC](https://doi.org/10.1164/rccm.202111-2634OC)
 11. Holgate ST. Pathogenesis of asthma. *Clin Exp Allergy.* 2008;38:872-897. doi:[10.1111/j.1365-2222.2008.02971.x](https://doi.org/10.1111/j.1365-2222.2008.02971.x)
 12. Varricchi G, Ferri S, Pepys J, et al. Biologics and airway remodeling in severe asthma. *Allergy.* 2022;77:3538-3552. doi:[10.1111/all.15473](https://doi.org/10.1111/all.15473)
 13. Fehrenbach H, Wagner C, Wegmann M. Airway remodeling in asthma: what really matters. *Cell Tissue Res.* 2017;367:551-569. doi:[10.1007/s00441-016-2566-8](https://doi.org/10.1007/s00441-016-2566-8)
 14. Liu G, Philp AM, Corte T, et al. Therapeutic targets in lung tissue remodelling and fibrosis. *Pharmacol Ther.* 2021;225:107839. doi:[10.1016/j.pharmthera.2021.107839](https://doi.org/10.1016/j.pharmthera.2021.107839)
 15. Hoshino M, Takahashi M, Aoike N. Expression of vascular endothelial growth factor, basic fibroblast growth factor, and angiogenin immunoreactivity in asthmatic airways and its relationship to angiogenesis. *J Allergy Clin Immunol.* 2001;107:295-301. doi:[10.1067/mai.2001.111928](https://doi.org/10.1067/mai.2001.111928)
 16. Chetta A, Zanini A, Foresi A, et al. Vascular component of airway remodeling in asthma is reduced by high dose of fluticasone. *Am J Respir Crit Care Med.* 2003;167:751-757. doi:[10.1164/rccm.200207-710OC](https://doi.org/10.1164/rccm.200207-710OC)
 17. Hauber HP, Foley SC, Hamid Q. Mucin overproduction in chronic inflammatory lung disease. *Can Respir J.* 2006;13:327-335. doi:[10.1155/2006/901417](https://doi.org/10.1155/2006/901417)
 18. Redhu NS, Shan L, Al-Subait D, et al. IgE induces proliferation in human airway smooth muscle cells: role of MAPK and STAT3 pathways. *Allergy Asthma Clin Immunol.* 2013;9:41. doi:[10.1186/1710-1492-9-41](https://doi.org/10.1186/1710-1492-9-41)
 19. Samitas K, Delimpoura V, Zervas E, Gaga M. Anti-IgE treatment, airway inflammation and remodelling in severe allergic asthma: current knowledge and future perspectives. *Eur Respir Rev.* 2015;24:594-601. doi:[10.1183/16000617.00001715](https://doi.org/10.1183/16000617.00001715)
 20. Matsuda K, Piliponsky AM, Iikura M, et al. Monomeric IgE enhances human mast cell chemokine production: IL-4 augments and dexamethasone suppresses the response. *J Allergy Clin Immunol.* 2005;116:1357-1363. doi:[10.1016/j.jaci.2005.08.042](https://doi.org/10.1016/j.jaci.2005.08.042)
 21. Dolgachev V, Berlin AA, Lukacs NW. Eosinophil activation of fibroblasts from chronic allergen-induced disease utilizes stem cell factor for phenotypic changes. *Am J Pathol.* 2008;172:68-76. doi:[10.2353/ajpath.2008.070082](https://doi.org/10.2353/ajpath.2008.070082)
 22. Tanaka H, Komai M, Nagao K, et al. Role of interleukin-5 and eosinophils in allergen-induced airway remodeling in mice. *Am J Respir Cell Mol Biol.* 2004;31:62-68. doi:[10.1165/rcmb.2003-0305OC](https://doi.org/10.1165/rcmb.2003-0305OC)
 23. Doucet C, Brouty-Boye D, Pottin-Clemenceau C, et al. Interleukin (IL) 4 and IL-13 act on human lung fibroblasts. Implication in asthma. *J Clin Invest.* 1998;101:2129-2139. doi:[10.1172/JCI741](https://doi.org/10.1172/JCI741)
 24. Wen FQ, Liu XD, Terasaki Y, et al. Interferon-gamma reduces interleukin-4- and interleukin-13-augmented transforming growth factor-beta2 production in human bronchial epithelial cells by targeting Smads. *Chest.* 2003;123:372S-373S.
 25. Sugita K, Steer CA, Martinez-Gonzalez I, et al. Type 2 innate lymphoid cells disrupt bronchial epithelial barrier integrity by targeting tight junctions through IL-13 in asthmatic patients. *J Allergy Clin Immunol.* 2018;141:300-310.e11. doi:[10.1016/j.jaci.2017.02.038](https://doi.org/10.1016/j.jaci.2017.02.038)
 26. Cohn L, Elias JA, Chupp GL. Asthma: mechanisms of disease persistence and progression. *Annu Rev Immunol.* 2004;22:789-815. doi:[10.1146/annurev.immunol.22.012703.104716](https://doi.org/10.1146/annurev.immunol.22.012703.104716)
 27. Malavia NK, Mih JD, Raub CB, Dinh BT, George SC. IL-13 induces a bronchial epithelial phenotype that is profibrotic. *Respir Res.* 2008;9:27. doi:[10.1186/1465-9921-9-27](https://doi.org/10.1186/1465-9921-9-27)
 28. Doucet C, Brouty-Boye D, Pottin-Clemenceau C, et al. IL-4 and IL-13 specifically increase adhesion molecule and inflammatory cytokine expression in human lung fibroblasts. *Int Immunol.* 1998;10:1421-1433. doi:[10.1093/intimm/10.10.1421](https://doi.org/10.1093/intimm/10.10.1421)
 29. Booth BW, Sandifer T, Martin EL, Martin LD. IL-13-induced proliferation of airway epithelial cells: mediation by intracellular growth factor mobilization and ADAM17. *Respir Res.* 2007;8:51. doi:[10.1186/1465-9921-8-51](https://doi.org/10.1186/1465-9921-8-51)
 30. Yang G, Volk A, Petley T, et al. Anti-IL-13 monoclonal antibody inhibits airway hyperresponsiveness, inflammation and airway remodeling. *Cytokine.* 2004;28:224-232. doi:[10.1016/j.cyto.2004.08.007](https://doi.org/10.1016/j.cyto.2004.08.007)
 31. Varricchi G, Pecoraro A, Marone G, et al. Thymic stromal Lymphopoietin isoforms, inflammatory disorders, and cancer. *Front Immunol.* 2018;9:1595. doi:[10.3389/fimmu.2018.01595](https://doi.org/10.3389/fimmu.2018.01595)
 32. Nagarkar DR, Poposki JA, Comeau MR, et al. Airway epithelial cells activate TH2 cytokine production in mast cells through IL-1 and thymic stromal lymphopoietin. *J Allergy Clin Immunol.* 2012;130:225-232.e4. doi:[10.1016/j.jaci.2012.04.019](https://doi.org/10.1016/j.jaci.2012.04.019)
 33. Braile M, Fiorelli A, Sorriento D, et al. Human lung-resident macrophages express and are targets of thymic stromal Lymphopoietin in the tumor microenvironment. *Cells.* 2021;10:2012. doi:[10.3390/cells10082012](https://doi.org/10.3390/cells10082012)
 34. Cane L, Poto R, Palestra F, et al. TSLP is localized in and released from human lung macrophages activated by T2-high and T2-low stimuli: relevance in asthma and COPD. *Eur J Intern Med.* 2024;124:89-98. doi:[10.1016/j.ejim.2024.02.020](https://doi.org/10.1016/j.ejim.2024.02.020)
 35. Cao L, Liu F, Liu Y, et al. TSLP promotes asthmatic airway remodeling via p38-STAT3 signaling pathway in human lung fibroblast. *Exp Lung Res.* 2018;44:288-301. doi:[10.1080/01902148.2018.1536175](https://doi.org/10.1080/01902148.2018.1536175)
 36. Cai LM, Zhou YQ, Yang LF, et al. Thymic stromal lymphopoietin induced early stage of epithelial-mesenchymal transition in human bronchial epithelial cells through upregulation of transforming growth factor beta 1. *Exp Lung Res.* 2019;45:221-235. doi:[10.1080/01902148.2019.1646841](https://doi.org/10.1080/01902148.2019.1646841)
 37. Chen ZG, Zhang TT, Li HT, et al. Neutralization of TSLP inhibits airway remodeling in a murine model of allergic asthma induced by chronic exposure to house dust mite. *PLoS One.* 2013;8:e51268. doi:[10.1371/journal.pone.0051268](https://doi.org/10.1371/journal.pone.0051268)
 38. Ye L, Mou Y, Wang J, Jin ML. Effects of microRNA-19b on airway remodeling, airway inflammation and degree of oxidative stress by targeting TSLP through the Stat3 signaling pathway in a mouse model of asthma. *Oncotarget.* 2017;8:47533-47546. doi:[10.18632/oncotarget.17258](https://doi.org/10.18632/oncotarget.17258)
 39. Lin SC, Chou HC, Chen CM, Chiang BL. Anti-thymic stromal lymphopoietin antibody suppresses airway remodeling in asthma through reduction of MMP and CTGF. *Pediatr Res.* 2019;86:181-187. doi:[10.1038/s41390-018-0239-x](https://doi.org/10.1038/s41390-018-0239-x)

40. Guo Z, Wu J, Zhao J, et al. IL-33 promotes airway remodeling and is a marker of asthma disease severity. *J Asthma*. 2014;51:863-869. doi:10.3109/02770903.2014.921196
41. Saglani S, Lui S, Ullmann N, et al. IL-33 promotes airway remodeling in pediatric patients with severe steroid-resistant asthma. *J Allergy Clin Immunol*. 2013;132:676-685.e13. doi:10.1016/j.jaci.2013.04.012
42. An G, Wang W, Zhang X, et al. Combined blockade of IL-25, IL-33 and TSLP mediates amplified inhibition of airway inflammation and remodelling in a murine model of asthma. *Respirology*. 2020;25:603-612. doi:10.1111/resp.13711
43. Xu X, Luo S, Li B, Dai H, Zhang J. IL-25 contributes to lung fibrosis by directly acting on alveolar epithelial cells and fibroblasts. *Exp Biol Med (Maywood)*. 2019;244:770-780. doi:10.1177/1535370219843827
44. Gregory LG, Jones CP, Walker SA, et al. IL-25 drives remodelling in allergic airways disease induced by house dust mite. *Thorax*. 2013;68:82-90. doi:10.1136/thoraxjnl-2012-202003
45. Yao X, Wang W, Li Y, et al. IL-25 induces airways angiogenesis and expression of multiple angiogenic factors in a murine asthma model. *Respir Res*. 2015;16:39. doi:10.1186/s12931-015-0197-3
46. Kardas G, Kuna P, Panek M. Biological therapies of severe asthma and their possible effects on airway remodeling. *Front Immunol*. 2020;11:1134. doi:10.3389/fimmu.2020.01134
47. Alagha K, Jarjour B, Bommart S, et al. Persistent severe hypereosinophilic asthma is not associated with airway remodeling. *Respir Med*. 2015;109:180-187. doi:10.1016/j.rmed.2014.12.008
48. Possa SS, Leick EA, Prado CM, Martins MA, Tibério IF. Eosinophilic inflammation in allergic asthma. *Front Pharmacol*. 2013;4:46. doi:10.3389/fphar.2013.00046
49. Halwani R, Vazquez-Tello A, Sumi Y, et al. Eosinophils induce airway smooth muscle cell proliferation. *J Clin Immunol*. 2013;33:595-604. doi:10.1007/s10875-012-9836-3
50. Hancox RJ, Pavord ID, Sears MR. Associations between blood eosinophils and decline in lung function among adults with and without asthma. *Eur Respir J*. 2018;51:1702536. doi:10.1183/13993003.02536-2017
51. Park HY, Chang Y, Kang D, et al. Blood eosinophil counts and the development of obstructive lung disease: the Kangbuk Samsung Health Study. *Eur Respir J*. 2021;58:2003823. doi:10.1183/13993003.03823-2020
52. Bajbouj K, AbuJabal R, Sahnoon L, Olivenstein R, Mahboub B, Hamid Q. IL-5 receptor expression in lung fibroblasts: potential role in airway remodeling in asthma. *Allergy*. 2023;78:882-885. doi:10.1111/all.15627
53. Coumou H, Westerhof GA, de Nijs SB, Zwinderman AH, Bel EH. Predictors of accelerated decline in lung function in adult-onset asthma. *Eur Respir J*. 2018;51:1701785. doi:10.1183/13993003.01785-2017
54. De Prado Gomez L, Pavord I, Busse W, et al. Long-term effect of dupilumab on prevention of lung function decline in patients with uncontrolled moderate-to-severe asthma: ATLAS trial design. *ERJ Open Res*. 2023;9:00417-2022. doi:10.1183/23120541.00417-2022
55. Al Heialy S, Ramakrishnan RK, Hamid Q. Recent advances in the immunopathogenesis of severe asthma. *J Allergy Clin Immunol*. 2022;149:455-465. doi:10.1016/j.jaci.2021.12.765
56. Banafea GH, Bakhashab S, Alshaihi HF, Natesan Pushparaj P, Rasool M. The role of human mast cells in allergy and asthma. *Bioengineered*. 2022;13:7049-7064. doi:10.1080/21655979.2022.2044278
57. Schiffer M, Peters K, Peters M. Comparison of airway remodeling in two different Endotypes of allergic asthma. *Int Arch Allergy Immunol*. 2022;183:1-12. doi:10.1159/000522189
58. Xie Y, Abel PW, Casale TB, Tu Y. TH17 cells and corticosteroid insensitivity in severe asthma. *J Allergy Clin Immunol*. 2022;149:467-479. doi:10.1016/j.jaci.2021.12.769
59. Tanaka J, Watanabe N, Kido M, et al. Human TSLP and TLR3 ligands promote differentiation of Th17 cells with a central memory phenotype under Th2-polarizing conditions. *Clin Exp Allergy*. 2009;39:89-100. doi:10.1111/j.1365-2222.2008.03151.x
60. Peters M, Kohler-Bachmann S, Lenz-Habijan T, et al. Influence of an allergen-specific Th17 response on remodeling of the airways. *Am J Respir Cell Mol Biol*. 2016;54:350-358. doi:10.1165/rcmb.2014-0429OC
61. Gao H, Ying S, Dai Y. Pathological roles of neutrophil-mediated inflammation in asthma and its potential for therapy as a target. *J Immunol Res*. 2017;2017:3743048. doi:10.1155/2017/3743048
62. Nair P, Surette MG, Virchow JC. Neutrophilic asthma: misconception or misnomer? *Lancet Respir Med*. 2021;9:441-443. doi:10.1016/S2213-2600(21)00023-0
63. Bourdin A, Neveu D, Vachier I, Paganin F, Godard P, Chanez P. Specificity of basement membrane thickening in severe asthma. *J Allergy Clin Immunol*. 2007;119:1367-1374. doi:10.1016/j.jaci.2007.01.055
64. Bourdin A, Vachier I, Godard P, Chanez P. Airway wall structural remodeling from endobronchial biopsies: what does it really mean? *J Allergy Clin Immunol*. 2009;123:711-712. doi:10.1016/j.jaci.2008.10.060
65. Cazes E, Giron-Michel J, Baouz S, et al. Novel anti-inflammatory effects of the inhaled corticosteroid fluticasone propionate during lung myofibroblastic differentiation. *J Immunol*. 2001;167:5329-5337. doi:10.4049/jimmunol.167.9.5329
66. Laitinen LA, Altraja A, Karjalainen EM, Laitinen A. Early interventions in asthma with inhaled corticosteroids. *J Allergy Clin Immunol*. 2000;105:S582-S585. doi:10.1016/s0091-6749(00)90062-9
67. Phipatanakul W, Mauger DT, Sorkness RL, et al. Effects of age and disease severity on systemic corticosteroid responses in asthma. *Am J Respir Crit Care Med*. 2017;195:1439-1448. doi:10.1164/rccm.201607-1453OC
68. Bateman ED, Boushey HA, Bousquet J, et al. Can guideline-defined asthma control be achieved? The gaining optimal asthma Control study. *Am J Respir Crit Care Med*. 2004;170:836-844. doi:10.1164/rccm.200401-033OC
69. Lommatzsch M, Lindner Y, Edner A, Bratke K, Kuepper M, Virchow JC. Adverse effects of salmeterol in asthma: a neuronal perspective. *Thorax*. 2009;64:763-769. doi:10.1136/thx.2008.110916
70. Virchow JC, Julius P, Lommatzsch M, et al. Neurotrophins are increased in bronchoalveolar lavage fluid after segmental allergen provocation. *Am J Respir Crit Care Med*. 1998;158:2002-2005. doi:10.1164/ajrccm.158.6.9803023
71. Castro M, Corren J, Pavord ID, et al. Dupilumab efficacy and safety in moderate-to-severe uncontrolled asthma. *N Engl J Med*. 2018;378:2486-2496. doi:10.1056/NEJMoa1804092
72. Corren J, Parnes JR, Wang L, et al. Tezepelumab in adults with uncontrolled asthma. *N Engl J Med*. 2017;377:936-946. doi:10.1056/NEJMoa1704064
73. Corren J, Castro M, O'Riordan T, et al. Dupilumab efficacy in patients with uncontrolled, moderate-to-severe allergic asthma. *J Allergy Clin Immunol Pract*. 2020;8:516-526. doi:10.1016/j.jaip.2019.08.050
74. Paganin F, Mangiapan G, Proust A, et al. Lung function parameters in omalizumab responder patients: An interesting tool? *Allergy*. 2017;72:1953-1961. doi:10.1111/all.13202
75. Busse WW, Bleecker ER, FitzGerald JM, et al. Long-term safety and efficacy of benralizumab in patients with severe, uncontrolled asthma: 1-year results from the BORA phase 3 extension trial. *Lancet Respir Med*. 2019;7:46-59. doi:10.1016/S2213-2600(18)30406-5
76. Castro M, Papi A, Corren J, et al. Effect of dupilumab on lung function parameters in oral corticosteroid-dependent patients with asthma enrolled in liberty asthma traverse. *Chest*. 2021;160:A1877-A1881. doi:10.1016/j.chest.2021.07.1675

77. Chipps BE, Hirsch I, Trudo F, Alacqua M, Zangrilli JG. Benralizumab efficacy for patients with fixed airflow obstruction and severe, uncontrolled eosinophilic asthma. *Ann Allergy Asthma Immunol*. 2020;124:79-86. doi:10.1016/j.anaai.2019.10.006
78. Chupp GL, Bradford ES, Albers FC, et al. Efficacy of mepolizumab add-on therapy on health-related quality of life and markers of asthma control in severe eosinophilic asthma (MUSCA): a randomised, double-blind, placebo-controlled, parallel-group, multicentre, phase 3b trial. *Lancet Respir Med*. 2017;5:390-400. doi:10.1016/S2213-2600(17)30125-X
79. Bel EH, Wenzel SE, Thompson PJ, et al. Oral glucocorticoid-sparing effect of mepolizumab in eosinophilic asthma. *N Engl J Med*. 2014;371:1189-1197. doi:10.1056/NEJMoa1403291
80. Wenzel S, Castro M, Corren J, et al. Dupilumab efficacy and safety in adults with uncontrolled persistent asthma despite use of medium-to-high-dose inhaled corticosteroids plus a long-acting beta2 agonist: a randomised double-blind placebo-controlled pivotal phase 2b dose-ranging trial. *Lancet*. 2016;388:31-44. doi:10.1016/S0140-6736(16)30307-5
81. Shimizu K, Tanabe N, Oguma A, et al. Parenchymal destruction in asthma: fixed airflow obstruction and lung function trajectory. *J Allergy Clin Immunol*. 2022;149:934-942.e8. doi:10.1016/j.jaci.2021.07.042
82. Rutting S, Tonga KO, King GG. Toward explaining fixed airflow obstruction in asthma. *J Allergy Clin Immunol*. 2022;149:890-892. doi:10.1016/j.jaci.2021.12.784
83. Humbert M, Beasley R, Ayres J, et al. Benefits of omalizumab as add-on therapy in patients with severe persistent asthma who are inadequately controlled despite best available therapy (GINA 2002 step 4 treatment): INNOVATE. *Allergy*. 2005;60:309-316. doi:10.1111/j.1398-9995.2004.00772.x
84. Hamada K, Oishi K, Murata Y, Hirano T, Matsunaga K. Feasibility of discontinuing biologics in severe asthma: An algorithmic approach. *J Asthma Allergy*. 2021;14:1463-1471. doi:10.2147/JAA.S340684
85. Menzella F, Fontana M, Galeone C, et al. A real-world evaluation of clinical outcomes of biologics and bronchial Thermoplasty for severe refractory asthma (BIOTERM). *J Asthma Allergy*. 2021;14:1019-1031. doi:10.2147/JAA.S324099
86. Ledford D, Busse W, Trzaskoma B, et al. A randomized multicenter study evaluating Xolair persistence of response after long-term therapy. *J Allergy Clin Immunol*. 2017;140:162-169.e2. doi:10.1016/j.jaci.2016.08.054
87. Moore WC, Kornmann O, Humbert M, et al. Stopping versus continuing long-term mepolizumab treatment in severe eosinophilic asthma (COMET study). *Eur Respir J*. 2022;59:2100396. doi:10.1183/13993003.00396-2021
88. Kavanagh JE, d'Ancona G, Elstad M, et al. Real-world effectiveness and the characteristics of a "super-responder" to Mepolizumab in severe eosinophilic asthma. *Chest*. 2020;158:491-500. doi:10.1016/j.chest.2020.03.042
89. Nair P, Wenzel S, Rabe KF, et al. Oral glucocorticoid-sparing effect of Benralizumab in severe asthma. *N Engl J Med*. 2017;376:2448-2458. doi:10.1056/NEJMoa1703501
90. Harvey ES, Langton D, Katelaris C, et al. Mepolizumab effectiveness and identification of super-responders in severe asthma. *Eur Respir J*. 2020;55:1902420. doi:10.1183/13993003.02420-2019
91. Eger K, Kroes JA, Ten Brinke A, et al. Long-term therapy response to anti-IL-5 biologics in severe asthma—a real-life evaluation. *J Allergy Clin Immunol Pract*. 2021;9:1194-1200. doi:10.1016/j.jaip.2020.10.010
92. Virchow JC, Hickey L, Du E, et al. In patients with severe asthma with eosinophilia in reslizumab clinical trials, high peripheral blood eosinophil levels are associated with low FEV1 reversibility. *Allergy Asthma Clin Immunol*. 2020;16:26. doi:10.1186/s13223-020-00424-2
93. Rabe KF, Nair P, Brusselle G, et al. Efficacy and safety of Dupilumab in glucocorticoid-dependent severe asthma. *N Engl J Med*. 2018;378:2475-2485. doi:10.1056/NEJMoa1804093
94. Castro M, Rabe KF, Corren J, et al. Dupilumab improves lung function in patients with uncontrolled, moderate-to-severe asthma. *ERJ Open Res*. 2020;6:00204-2019. doi:10.1183/23120541.00204-2019
95. Marone G, Spadaro G, Braile M, et al. Tezepelumab: a novel biological therapy for the treatment of severe uncontrolled asthma. *Expert Opin Investig Drugs*. 2019;28:931-940. doi:10.1080/13543784.2019.1672657
96. Pham T, Cook B, Parnes JR, et al. Tezepelumab reduces biomarkers of airway remodeling, MMP-10 and MMP-3: exploratory results from the phase 3 NAVIGATOR study. *Am J Respir Crit Care Med*. 2022;205:A2359.
97. Diver S, Khalfaoui L, Emson C, et al. Effect of tezepelumab on airway inflammatory cells, remodelling, and hyperresponsiveness in patients with moderate-to-severe uncontrolled asthma (CASCADE): a double-blind, randomised, placebo-controlled, phase 2 trial. *Lancet Respir Med*. 2021;9:1299-1312. doi:10.1016/S2213-2600(21)00226-5
98. Tang M, Elicker BM, Henry T, et al. Mucus plugs persist in asthma, and changes in mucus plugs associate with changes in airflow over time. *Am J Respir Crit Care Med*. 2022;205:1036-1045. doi:10.1164/rccm.202110-2265OC
99. Anai M, Yoshida C, Izumi H, et al. Successful treatment with dupilumab for mucus plugs in severe asthma. *Respirol Case Rep*. 2023;11:e01074. doi:10.1002/rcr2.1074
100. Chan R, Duraikannu C, Lipworth B. Clinical associations of mucus plugging in moderate to severe asthma. *J Allergy Clin Immunol Pract*. 2023;11:195-199.e2. doi:10.1016/j.jaip.2022.09.008
101. Liaqat A, Mason M, Foster B, et al. Evidence-based approach of biologic therapy in bronchial asthma. *J Clin Med*. 2023;12:4321. doi:10.3390/jcm12134321
102. Fildan AP, Rajnoveanu RM, Cirjaliu R, et al. Biological therapies targeting the type 2 inflammatory pathway in severe asthma (review). *Exp Ther Med*. 2021;22:1263. doi:10.3892/etm.2021.10698
103. Sakai N, Koya T, Murai Y, et al. Effect of Benralizumab on mucus plugs in severe eosinophilic asthma. *Int Arch Allergy Immunol*. 2023;184:783-791. doi:10.1159/000530392
104. Nordenmark LH, Hellqvist A, Emson C, et al. Tezepelumab and mucus plugs in patients with moderate-to-severe asthma. *NEJM Evid*. 2023;2:EVIDoa2300135. doi:10.1056/EVIDoa2300135
105. Hamakawa M, Ishida T. Usefulness of Mepolizumab for mucus plugs. *Intern Med*. 2024. doi:10.2169/internalmedicine.3531-24
106. Tajiri T, Suzuki M, Nishiyama H, et al. Efficacy of dupilumab for airway hypersecretion and airway wall thickening in patients with moderate-to-severe asthma: a prospective, observational study. *Allergol Int*. 2024;73:406-415. doi:10.1016/j.alit.2024.02.002 Epub ahead of print.
107. Svenningsen S, Eddy RL, Kjarsgaard M, Parraga G, Nair P. Effects of anti-T2 biologic treatment on lung ventilation evaluated by MRI in adults with prednisone-dependent asthma. *Chest*. 2020;158:1350-1360. doi:10.1016/j.chest.2020.04.056
108. Hoshino M, Ohtawa J. Effects of adding omalizumab, an anti-immunoglobulin E antibody, on airway wall thickening in asthma. *Respiration*. 2012;83:520-528. doi:10.1159/000334701
109. Tajiri T, Niimi A, Matsumoto H, et al. Comprehensive efficacy of omalizumab for severe refractory asthma: a time-series observational study. *Ann Allergy Asthma Immunol*. 2014;113:470-475.e2. doi:10.1016/j.anaai.2014.06.004
110. Haldar P, Brightling CE, Hargadon B, et al. Mepolizumab and exacerbations of refractory eosinophilic asthma. *N Engl J Med*. 2009;360:973-984. doi:10.1056/NEJMoa0808991
111. Haldar P, Brightling CE, Singapuri A, et al. Outcomes after cessation of mepolizumab therapy in severe eosinophilic asthma: a

- 12-month follow-up analysis. *J Allergy Clin Immunol.* 2014;133:921-923. doi:10.1016/j.jaci.2013.11.026
112. Przybyszowski M, Gross-Sondej I, Zarychta J, et al. Late breaking abstract - the impact of treatment with mepolizumab on airway remodeling in patients with severe eosinophilic asthma. *Eur Respir J.* 2021;58:PA894. doi:10.1183/13993003.congress-2021.PA894
 113. McIntosh MJ, Koener HK, Eddy RL, et al. CT mucus score and (129)Xe MRI ventilation defects after 2.5 Years' anti-IL-5Ralpha in eosinophilic asthma. *Chest.* 2023;164:27-38. doi:10.1016/j.chest.2023.02.009
 114. Genofre E, Carstens D, DeBacker W, et al. The effects of benralizumab on airway geometry and dynamics in severe eosinophilic asthma: a single-arm study design exploring a functional respiratory imaging approach. *Respir Res.* 2023;24:121. doi:10.1186/s12931-023-02415-4
 115. Varricchi G, Poto R. Towards precision medicine in COPD: targeting type 2 cytokines and alarmins. *Eur J Intern Med.* 2024;125:28-31. doi:10.1016/j.ejim.2024.05.011
 116. Calzetta L, Aiello M, Frizzelli A, et al. The impact of monoclonal antibodies on airway smooth muscle contractility in asthma: a systematic review. *Biomedicine.* 2021;9:1281. doi:10.3390/biomedicines9091281
 117. Flood-Page P, Menzies-Gow A, Phipps S, et al. Anti-IL-5 treatment reduces deposition of ECM proteins in the bronchial subepithelial basement membrane of mild atopic asthmatics. *J Clin Invest.* 2003;112:1029-1036. doi:10.1172/JCI17974
 118. Domvri K, Tsiouprou I, Bakakos P, et al. Effect of Mepolizumab on airways remodeling in patients with late-onset severe eosinophilic asthma and fixed obstruction (preliminary data of the MESILICO study) European Respiratory Society. 2023 pp. OA3152.
 119. Chachi L, Diver S, Kaul H, et al. Computational modelling prediction and clinical validation of impact of benralizumab on airway smooth muscle mass in asthma. *Eur Respir J.* 2019;54:1900930. doi:10.1183/13993003.00930-2019
 120. Zastrzezyńska W, Przybyszowski M, Bazan-Socha S, et al. Omalizumab may decrease the thickness of the reticular basement membrane and fibronectin deposit in the bronchial mucosa of severe allergic asthmatics. *J Asthma.* 2020;57:468-477. doi:10.1080/02770903.2019.1585872
 121. Canonica GW, Bourdin A, Peters AT, et al. Dupilumab demonstrates rapid onset of response across three type 2 inflammatory diseases. *J Allergy Clin Immunol Pract.* 2022;10:1515-1526. doi:10.1016/j.jaip.2022.02.026
 122. Crimi C, Campisi R, Cacopardo G, et al. Real-life effectiveness of mepolizumab in patients with severe refractory eosinophilic asthma and multiple comorbidities. *World Allergy Organ J.* 2020;13:100462. doi:10.1016/j.waojou.2020.100462
 123. Hanania NA, Lanier B, Iqbal A, et al. Significant improvement in lung function following 16 weeks of omalizumab treatment: pooled analyses in patients with moderate-to-severe allergic asthma. *Chest.* 2018;154:2A-3A. doi:10.1016/j.chest.2018.08.003
 124. Papi A, Swanson BN, Staudinger H, et al. Dupilumab rapidly and significantly improves lung function and decreases inflammation by 2 weeks after treatment initiation in patients with uncontrolled persistent asthma. *Am J Respir Crit Care Med.* 2017;195:A6444.
 125. Pelaia C, Busceti MT, Vatrella A, et al. Real-life rapidity of benralizumab effects in patients with severe allergic eosinophilic asthma: assessment of blood eosinophils, symptom control, lung function and oral corticosteroid intake after the first drug dose. *Pulm Pharmacol Ther.* 2019;58:101830. doi:10.1016/j.pupt.2019.101830
 126. Wechsler ME, Ruddy MK, Pavord ID, et al. Efficacy and safety of Itepekimab in patients with moderate-to-severe asthma. *N Engl J Med.* 2021;385:1656-1668. doi:10.1056/NEJMoa2024257
 127. Spahn JD, Brightling CE, O'Byrne PM, et al. Effect of biologic therapies on airway hyperresponsiveness and allergic response: a systematic literature review. *J Asthma Allerg.* 2023;16:755-774. doi:10.2147/JAA.S410592
 128. Ledda AG, Costanzo G, Sambugaro G, et al. Eosinophil cationic protein variation in patients with asthma and CRSwNP treated with Dupilumab. *Life (Basel).* 2023;13:1884. doi:10.3390/life13091884
 129. Chan R, Lipworth BJ. Impact of biologic therapy on the small airways asthma phenotype. *Lung.* 2022;200:691-696. doi:10.1007/s00408-022-00579-2
 130. Krings JG, Gierada DS. Do biologic therapies decrease mucus plugging in asthma? *NEJM Evid.* 2023;2:e2300179. doi:10.1056/EVIDe2300179
 131. Nassenstein C, Braun A, Erpenbeck VJ, et al. The neurotrophins nerve growth factor, brain-derived neurotrophic factor, neurotrophin-3, and neurotrophin-4 are survival and activation factors for eosinophils in patients with allergic bronchial asthma. *J Exp Med.* 2003;198:455-467. doi:10.1084/jem.20010897
 132. Bagnasco D, Ferrando M, Varricchi G, Passalacqua G, Canonica GW. A critical evaluation of anti-IL-13 and anti-IL-4 strategies in severe asthma. *Int Arch Allergy Immunol.* 2016;170:122-131. doi:10.1159/000447692
 133. Scott G, Asrat S, Allinne J, et al. IL-4 and IL-13, not eosinophils, drive type 2 airway inflammation, remodeling and lung function decline. *Cytokine.* 2023;162:156091. doi:10.1016/j.cyto.2022.156091
 134. Chan R, Stewart K, Misirovs R, Lipworth BJ. Targeting downstream type 2 cytokines or upstream epithelial alarmins for severe asthma. *J Allergy Clin Immunol Pract.* 2022;10:1497-1505. doi:10.1016/j.jaip.2022.01.040
 135. Bencze D, Fekete T, Pazmandi K. Type I interferon production of plasmacytoid dendritic cells under control. *Int J Mol Sci.* 2021;22:4190. doi:10.3390/ijms22084190
 136. Hamilton JD, Suarez-Farinas M, Dhingra N, et al. Dupilumab improves the molecular signature in skin of patients with moderate-to-severe atopic dermatitis. *J Allergy Clin Immunol.* 2014;134:1293-1300. doi:10.1016/j.jaci.2014.10.013
 137. Guttman-Yassky E, Bissonnette R, Ungar B, et al. Dupilumab progressively improves systemic and cutaneous abnormalities in patients with atopic dermatitis. *J Allergy Clin Immunol.* 2019;143:155-172. doi:10.1016/j.jaci.2018.08.022
 138. Dellon ES, Rothenberg ME, Collins MH, et al. Dupilumab in adults and adolescents with eosinophilic esophagitis. *N Engl J Med.* 2022;387:2317-2330. doi:10.1056/NEJMoa2205982
 139. Roth M, Zhao F, Zhong J, Lardinois D, Tamm M. Serum IgE induced airway smooth muscle cell remodeling is independent of allergens and is prevented by Omalizumab. *PLoS One.* 2015;10:e0136549. doi:10.1371/journal.pone.0136549
 140. Zhang N, Xu J, Jiang C, Lu S. Neuro-immune regulation in inflammation and airway remodeling of allergic asthma. *Front Immunol.* 2022;13:894047. doi:10.3389/fimmu.2022.894047
 141. Phipatanakul W, Mauger DT, Guilbert TW, et al. Preventing asthma in high risk kids (PARK) with omalizumab: design, rationale, methods, lessons learned and adaptation. *Contemp Clin Trials.* 2021;100:106228. doi:10.1016/j.cct.2020.106228
 142. Tsuge M, Ikeda M, Tsukahara H. Novel lung growth strategy with biological therapy targeting airway remodeling in childhood bronchial asthma. *Children (Basel).* 2022;9:1253. doi:10.3390/children9081253
 143. Jackson DJ, Heaney LG, Humbert M, et al. Reduction of daily maintenance inhaled corticosteroids in patients with severe eosinophilic asthma treated with benralizumab (SHAMAL): a randomised, multicentre, open-label, phase 4 study. *Lancet.* 2024;403:271-281. doi:10.1016/S0140-6736(23)02284-5
 144. Buchheit KM, Sohail A, Hacker J, et al. Rapid and sustained effect of dupilumab on clinical and mechanistic outcomes in aspirin-exacerbated respiratory disease. *J Allergy Clin Immunol.* 2022;150:415-424. doi:10.1016/j.jaci.2022.04.007

145. Bachert C, Han JK, Desrosiers M, et al. Efficacy and safety of dupilumab in patients with severe chronic rhinosinusitis with nasal polyps (LIBERTY NP SINUS-24 and LIBERTY NP SINUS-52): results from two multicentre, randomised, double-blind, placebo-controlled, parallel-group phase 3 trials. *Lancet*. 2019;394:1638-1650. doi:[10.1016/S0140-6736\(19\)31881-1](https://doi.org/10.1016/S0140-6736(19)31881-1)
146. Nolasco S, Crimi C, Pelaia C, et al. Benralizumab effectiveness in severe eosinophilic asthma with and without chronic rhinosinusitis with nasal polyps: a real-world multicenter study. *J Allergy Clin Immunol Pract*. 2021;9:4371-4380.e4. doi:[10.1016/j.jaip.2021.08.004](https://doi.org/10.1016/j.jaip.2021.08.004)
147. McIntosh MJ, Kooner HK, Eddy RL, et al. Asthma control, airway mucus, and (129)Xe MRI ventilation after a single Benralizumab dose. *Chest*. 2022;162:520-533. doi:[10.1016/j.chest.2022.03.003](https://doi.org/10.1016/j.chest.2022.03.003)
148. Matsuno O, Minamoto S. Rapid effect of benralizumab for severe asthma with chronic rhinosinusitis with nasal polyps. *Pulm Pharmacol Ther*. 2020;64:101965. doi:[10.1016/j.pupt.2020.101965](https://doi.org/10.1016/j.pupt.2020.101965)
149. Farah CS, Badal T, Reed N, et al. Mepolizumab improves small airway function in severe eosinophilic asthma. *Respir Med*. 2019;148:49-53. doi:[10.1016/j.rmed.2019.01.016](https://doi.org/10.1016/j.rmed.2019.01.016)
150. Menzies-Gow A, Corren J, Bourdin A, et al. Tezepelumab in adults and adolescents with severe, uncontrolled asthma. *N Engl J Med*. 2021;384:1800-1809. doi:[10.1056/NEJMoa2034975](https://doi.org/10.1056/NEJMoa2034975)
151. Svenningsen S, Haider E, Boylan C, et al. CT and functional MRI to evaluate airway mucus in severe asthma. *Chest*. 2019;155:1178-1189. doi:[10.1016/j.chest.2019.02.403](https://doi.org/10.1016/j.chest.2019.02.403)
152. Riccio AM, Dal Negro RW, Micheletto C, et al. Omalizumab modulates bronchial reticular basement membrane thickness and eosinophil infiltration in severe persistent allergic asthma patients. *Int J Immunopathol Pharmacol*. 2012;25:475-484. doi:[10.1177/039463201202500217](https://doi.org/10.1177/039463201202500217)
153. Laviolette M, Gossage DL, Gauvreau G, et al. Effects of benralizumab on airway eosinophils in asthmatic patients with sputum eosinophilia. *J Allergy Clin Immunol*. 2013;132:1086-1096.e5. doi:[10.1016/j.jaci.2013.05.020](https://doi.org/10.1016/j.jaci.2013.05.020)
154. Shirai T, Akamatsu T, Hirai K, et al. Oscillometry improves earlier than spirometry after benralizumab initiation in severe asthma. *Allergy*. 2020;75:2678-2680. doi:[10.1111/all.14339](https://doi.org/10.1111/all.14339)
155. Castro M, Papi A, Porsbjerg C, et al. Evaluating the effect of Dupilumab on type 2 airway inflammation and mucus plugging in patients with uncontrolled moderate-to-severe asthma: the VESTIGE trial. *J Allergy Clin Immunol*. 2024;153:AB368.
156. Svenningsen S, Haider EA, Eddy RL, Parraga G, Nair P. Normalisation of MRI ventilation heterogeneity in severe asthma by dupilumab. *Thorax*. 2019;74:1087-1088. doi:[10.1136/thoraxjnl-2019-213415](https://doi.org/10.1136/thoraxjnl-2019-213415)
157. Mappanasingam AA, Friedlander Y, Thakar A, Ragunayagam N, Kjarsgaard M, et al. Abstract: Effect of dupilumab on pulmonary vascular volume in patients with moderate-to-severe asthma. *Am J Respir Crit Care Med*. 2024;209:A1052.
158. Svenningsen S, Kjarsgaard M, Haider E, et al. Effects of Dupilumab on mucus plugging and ventilation defects in patients with moderate-to-severe asthma: a randomized, double-blind, placebo-controlled trial. *Am J Respir Crit Care Med*. 2023;208:995-997. doi:[10.1164/rccm.202306-1102LE](https://doi.org/10.1164/rccm.202306-1102LE)
159. Saglani S, Payne DN, Zhu J, et al. Early detection of airway wall remodeling and eosinophilic inflammation in preschool wheezers. *Am J Respir Crit Care Med*. 2007;176:858-864. doi:[10.1164/rccm.200702-212OC](https://doi.org/10.1164/rccm.200702-212OC)
160. Lezmi G, Gosset P, Deschildre A, et al. Airway remodeling in preschool children with severe recurrent wheeze. *Am J Respir Crit Care Med*. 2015;192:164-171. doi:[10.1164/rccm.201411-1958OC](https://doi.org/10.1164/rccm.201411-1958OC)
161. Pohunek P, Warner JO, Turzikova J, et al. Markers of eosinophilic inflammation and tissue re-modelling in children before clinically diagnosed bronchial asthma. *Pediatr Allergy Immunol*. 2005;16:43-51. doi:[10.1111/j.1399-3038.2005.00239.x](https://doi.org/10.1111/j.1399-3038.2005.00239.x)
162. Varricchi G, Brightling CE, Grainge C, Lambrecht BN, Chanez P. Airway remodelling in asthma and the epithelium: on the edge of a new era. *Eur Respir J*. 2024;63:2301619. doi:[10.1183/13993003.01619-2023](https://doi.org/10.1183/13993003.01619-2023)
163. Kuang FL, Legrand F, Makiya M, et al. Benralizumab for PDGFRA-negative hypereosinophilic syndrome. *N Engl J Med*. 2019;380:1336-1346. doi:[10.1056/NEJMoa1812185](https://doi.org/10.1056/NEJMoa1812185)
164. Alen Coutinho I, Regateiro FS, Loureiro C, Todo-Bom A. Benralizumab in severe and refractory PDGFRA-negative hypereosinophilic syndrome. *J Clin Immunol*. 2021;41:688-690. doi:[10.1007/s10875-020-00946-9](https://doi.org/10.1007/s10875-020-00946-9)
165. Chen MM, Roufosse F, Wang SA, et al. An international, retrospective study of off-label biologic use in the treatment of Hypereosinophilic syndromes. *J Allergy Clin Immunol Pract*. 2022;10:1217-1228.e3. doi:[10.1016/j.jaip.2022.02.006](https://doi.org/10.1016/j.jaip.2022.02.006)
166. Dellon ES, Simon D, Wechsler ME. The potential role of biologics as first-line therapy in eosinophilic disorders. *J Allergy Clin Immunol Pract*. 2022;10:1169-1176. doi:[10.1016/j.jaip.2022.01.043](https://doi.org/10.1016/j.jaip.2022.01.043)
167. Braido F, Blasi F, Canonica GW, et al. Mild/moderate asthma network in Italy (MANI): a long-term observational study. *J Asthma*. 2022;59:1908-1913. doi:[10.1080/02770903.2021.1968895](https://doi.org/10.1080/02770903.2021.1968895)

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