









IL-6 in the spotlight: From cardiovascular pathophysiology to therapy

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Abstract

Background: Interleukin-6 (IL-6), a pro-inflammatory cytokine and anti-inflammatory myokine, is involved in immune regulation and metabolic control. Persistent IL-6 activation, particularly via trans-signalling, is implicated in endothelial dysfunction, atherosclerosis, myocardial fibrosis, and adverse remodelling.

Methods: This review summarizes experimental, genetic, and clinical evidence on the role of IL-6 in cardiovascular disease (CVD), focusing on the molecular pathways, prognostic value, and its potential as a therapeutic target. Experimental and genome-wide association studies, as well as clinical trials—including IL-6-targeted interventions—are comprehensively reviewed.

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Results: While acute activation of IL-6/STAT signalling, that is, before or during myocardial infarction, is protective, chronic activation leads to maladaptive changes in the myocardium. Mechanistically, IL-6 promotes vascular inflammation, monocyte recruitment, coagulation, and fibrosis. Mendelian randomization implies a causal role of IL-6 signalling in coronary artery disease, stroke, and atrial fibrillation. Simultaneously, elevated IL-6 levels predict adverse outcomes in acute coronary syndromes and heart failure. Pharmacological IL-6 inhibition reduces systemic inflammation, with pilot data implying acceptable safety profiles. Large phase III outcome trials are ongoing, shedding light on the impact of IL-6 blockade in high-risk populations, such as those with chronic kidney disease and heart failure with preserved or mildly reduced ejection fraction.

Conclusions: IL-6 plays a dual role-protective versus pathogenic and acute versus chronic. Chronic IL-6 links inflammation and metabolic dysregulation to structural cardiovascular damage. Chronic IL-6 activation requires therapeutic approaches tailored to disease context, timing and signalling mode. The results of ongoing trials will clarify whether IL-6-targeted interventions can be integrated into cardiovascular prevention and management strategies, complementing established therapies and addressing residual inflammatory risk.

KEYWORDS

cardiovascular disease, chronic kidney disease, heart failure, IL-6, inflammation, monoclonal antibodies

1 | INTRODUCTION

Cardiovascular and cerebrovascular diseases (CVD) cause one of three deaths globally, with a tremendous effect on quality of life and overall morbidity, particularly so in patients with cardiometabolic disease, including renal disease. Therefore, CVD imposes a substantial societal and economic burden. This observation calls for a better understanding of their mechanisms to improve and refine strategies for their prevention and therapy. The intricate pathogenesis of CVD is multifactorial, with chronic inflammatory states accelerating their development, contributing to their high fatality rate.¹⁻⁴

Whereas acute low-grade upregulation of the pro-inflammatory cytokines is described to have cardioprotective roles during acute myocardial infarction, they are potentially harmful if sustained or present at high concentrations.^{5,6} Hallmarks of low-grade inflammatory states involved in CVD pathogenesis include the activation of the innate immune system and increased circulating levels of pro-inflammatory mediators, such as interleukins (ILs), tumour necrosis factors (TNFs), as well as the biomarker C-reactive protein (CRP). Among these pro-inflammatory molecules, IL-6 is an immune-mediated cytokine that is elevated in systemic inflammatory states. In

the MESA (Multi-Ethnic Study of Atherosclerosis) study, patients with higher IL-6 levels tended to be older, more often female and more frequently had a smoking history.⁷ They also had higher rates of hypertension, diabetes, and a family history of coronary heart disease. Additionally, on median, these patients exhibited higher systolic blood pressure, heart rate, and waist circumference, and included a larger proportion of African American and Hispanic individuals compared to those with lower IL-6 levels,⁷ highlighting that this molecule may sit at the crossroads of cardiovascular, renal, and metabolic risk pathways.

Anti-inflammatory therapies have been studied for their effectiveness in reducing CV events in patients with established CVD.⁸⁻¹⁰ Among these, colchicine gained the Food and Drug Administration (FDA) approval for reducing secondary CV risk, despite a recent trial challenging its effectiveness in patients with a recent acute myocardial infarction (MI) receiving intensive treatment.^{10,11} More specific anti-inflammatory drugs including inhibitory antibodies against ILs are being tested. The Canakinumab Anti-inflammatory Thrombosis Outcomes Study (CANTOS) demonstrated that targeted use of IL-1 β inhibition substantially reduces CV events.¹² Notably, beneficial effects were most pronounced in those with the largest reduction of IL-6 following anti-IL1 β injections.¹³

Consequently, ongoing trials are investigating novel therapies aimed at inhibiting circulating IL-6 to reduce CVD events.^{14,15}

This review provides a comprehensive and up-to-date overview of the role of IL-6 in cardio-cerebrovascular diseases (Figure 1), focusing on the molecular pathways through which it contributes to cardiovascular dysfunction and the onset of CVD. Furthermore, it explores the translational relevance of these mechanisms in clinical practice and highlights ongoing trials targeting this pathway.

2 | A COMPLEX MEDIATOR

The journey of IL-6 dates back to 1988, previously known as B cell stimulatory factor 2, hybridoma plasmacytoma growth factor, hybridoma growth factor, or hepatocyte-stimulating factor.¹⁶ Since then, studies have established IL-6 as a pleiotropic cytokine with a central role in inflammation and immune response, haematopoiesis, bone

metabolism, the endocrine system, and the development of the central nervous and CV systems.^{17,18} Similarly to IL-1 β and TNF- α , IL-6 stimulates the synthesis of acute phase proteins; thus, acting as a primary pro-inflammatory cytokine.¹⁹ In addition, it modulates the transition from the acute to the chronic phase of inflammation, enhancing the immune response by promoting end-stage B-cell differentiation, immunoglobulin secretion, and T-cell activation,²⁰ with potential anti-inflammatory roles.

IL-6 is composed of a single chain phosphorylated glycoprotein, comprising four helix-long α -chain bundles, which are structured in three dimensions to achieve the tertiary shape.^{21,22} As a pleiotropic mediator, it is produced by a wide range of cell types across different tissues and physiological contexts. Among immune cells, IL-6 is primarily secreted by monocytes/macrophages, dendritic cells, B cells, and T lymphocytes—particularly Th17 cells—in response to infections and inflammatory stimuli. Within non-immune tissues, adipocytes release IL-6, classifying it as an adipocytokine, while skeletal muscle fibres produce it during physical activity, where it acts as

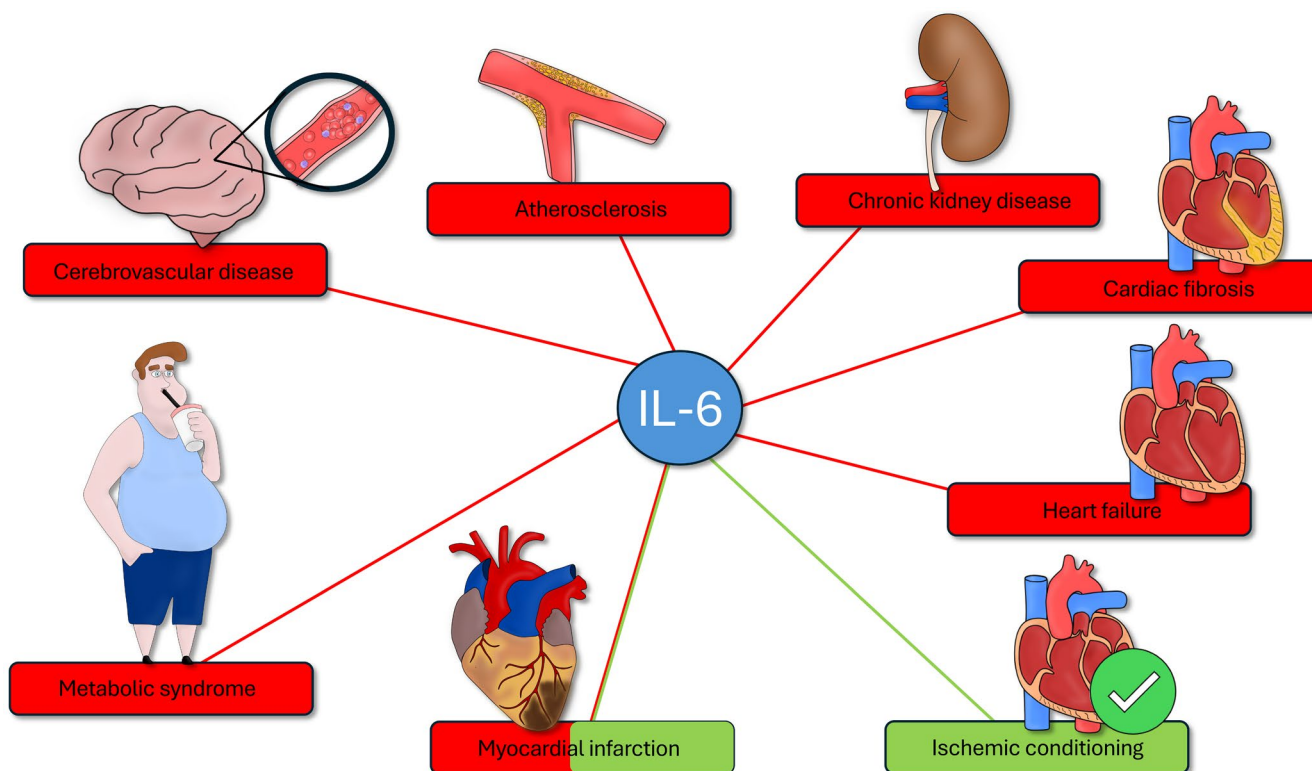


FIGURE 1 Schematic representation of the dual role of interleukin-6 (IL-6). IL-6 promotes endothelial activation, macrophage recruitment, and accelerated atherosclerosis. It alters cardiomyocyte function, increases neurohormonal tone, and contributes to interstitial fibrosis, coronary microvascular dysfunction, and systemic inflammation. IL-6 is also associated with metabolic dysregulation, further amplifying cardiovascular risk. Conversely, transient and localized IL-6 activation can exert protective effects, supporting ischemic preconditioning, tissue repair, and adaptive immune responses during the early phase of myocardial infarction healing. The overall impact of IL-6 thus depends on the balance between acute, reparative activation and chronic, maladaptive inflammation, reflecting its context-dependent role in cardiovascular pathophysiology. IL-6, Interleukin 6.

a myokine. Endothelial cells and vascular smooth muscle cells also generate IL-6 in response to pro-inflammatory signals, contributing to vascular inflammation and remodelling. In the bone microenvironment, osteoblasts are a significant source of IL-6, implicating it in bone metabolism and inflammation. Additionally, fibroblasts, keratinocytes, mesangial cells, and certain epithelial cells—including bronchial and intestinal epithelial cells—can produce IL-6 under specific pathological conditions, further underscoring its multifaceted role in both local and systemic immune responses. IL-6's signalling varies throughout the body and occurs via two primary pathways, defined as classic signalling and trans-signalling, with potential opposite effects (Figure 2). The cytokine binds to cells via a membrane-bound type I cytokine receptor complex composed of a transmembrane glycoprotein called IL-6R α (or CD126), and a gp130/IL6R β (gp130 or CD130) signalling subunit.²³ The complex formation begins with the four alpha-chains of IL-6 capturing the alpha-chain of IL6R α , creating a five alpha-chain complex, followed by gp130 binding. No signalling can occur until the trimer IL-6/IL-6R α /gp130 undergoes homodimerization to

form a hexameric complex.²⁴ The effects of IL-6 depend on the localization and expression of its specific receptor, IL-6R α . While the signal-transducing subunit gp130 is ubiquitously expressed across virtually all cell types, the membrane-bound IL-6 receptor (IL-6R α) is expressed only by a limited subset of cells, such as hepatocytes and certain leukocyte populations.

A further pathway, known as trans-signalling, has been described. This pathway begins with the cleavage of CD126, mediated by the metalloproteases ADAM10 and ADAM17, producing its soluble form, sIL-6R, which acts as a carrier of IL-6 to the gp130 site in cells lacking IL-6R.²⁵ A third mode of IL-6 signalling, termed cluster signalling, involves preassembled membrane-bound IL-6–IL-6R α complexes on one cell engaging gp130 receptors on adjacent target cells. Classical IL-6R signalling likely promotes a stronger anti-inflammatory response, whereas the trans-signalling plays a pro-inflammatory role.²⁶ Based on this hypothesis, autoimmune disorders and cancer seem to be primarily driven by IL-6 trans-signalling, whereas regenerative and protective activities are mediated by classic IL-6 signalling.²⁷ The distinct modes of IL-6 receptor

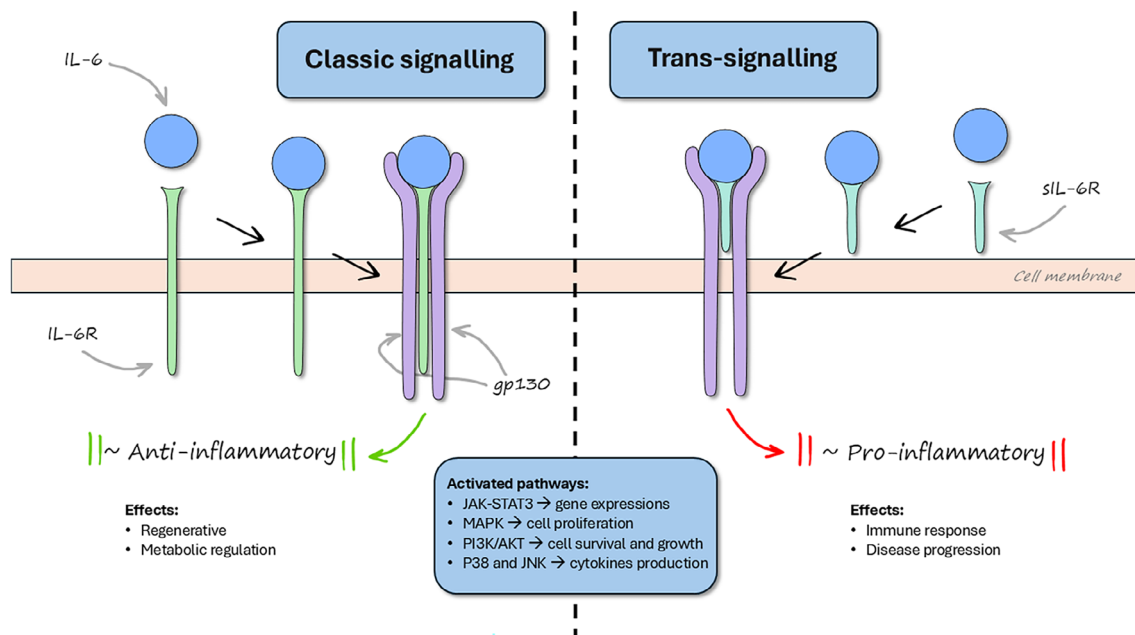


FIGURE 2 IL-6 signalling. IL-6 signalling occurs through two distinct mechanisms: Classic signalling and trans-signalling. Classic signalling involves the binding of IL-6 to its membrane-bound receptor (IL-6R) and is generally associated with anti-inflammatory effects. In contrast, trans-signalling occurs when IL-6 binds to the soluble IL-6 receptor (sIL-6R), which allows IL-6 to act on a broader range of cells, and is primarily linked to pro-inflammatory responses. While both pathways converge on similar downstream cascades, such as JAK-STAT3 and PI3K/AKT, they differ in intensity, duration, and cellular consequences. Classic signalling is limited to cells expressing membrane-bound IL-6R, including hepatocytes and certain leukocytes, whereas trans-signalling can influence a wider array of cell types due to the widespread distribution of sIL-6R. Moreover, the same signalling input can elicit distinct responses depending on the specific transcriptional landscape and co-regulatory factors within different cell types, resulting in varied gene expression profiles and functional outcomes. AKT, Protein kinase B; IL-6, Interleukin 6; IL-6R, Interleukin 6 receptor; JAK, Janus kinase-signal transducer; JNK, C-Jun N-terminal kinase; MAPK, Mitogen-activated protein kinase; PI3K, Phosphoinositide 3-kinase; sIL-6R, Soluble interleukin 6 receptor; STAT3, Activator of transcription 3.

activation lead to the engagement of different intracellular signalling pathways. Depending on the microenvironment and the cell type involved, downstream cascades may include Janus kinases (JAKs), mitogen-activated protein kinases (MAPKs), phosphatidylinositol 3-kinase/protein kinase B (PI3K/Akt), with the activation of different transcription factors such as signal transducer and activator of transcription 3 (STAT3) and nuclear factor kappa B (NF- κ B), ultimately resulting in specific physiological or pathological outcomes.

3 | A KEY DRIVER OF INFLAMM-AGING AND METAINFLAMMATION

From a physiological perspective, IL-6 functions as a key alarm signal, rapidly induced in response to a variety of insults—such as infections, tissue injury, and malignant transformation—and serving as a potent trigger of inflammatory responses.²⁸ By promoting inflammation, IL-6 plays a pivotal role in the pathogenesis of numerous conditions, including cardiovascular²⁹ and musculoskeletal diseases,³⁰ cancer,³¹ neurological disorders,³² anaemia,³³ inflammatory bowel disease,³⁴ asthma, and other chronic inflammatory pulmonary conditions.³⁵ IL-6 is also deeply implicated in the development and maintenance of chronic inflammatory states.³⁶ It has emerged as a central mediator in age-related inflammation, contributing to a process known as *inflammaging*—a state of persistent, low-grade inflammation that develops with aging. This phenomenon is driven by a complex interplay of mechanisms, including immunosenescence, impaired cellular clearance (“garbaging”), chronic infections, dysbiosis, metaflammation, clonal haematopoiesis of indeterminate potential (CHIP), as well as both genetic and epigenetic alterations.³ IL-6 has been proven to mediate most of such mechanisms. Being produced at high levels by senescent cells, IL-6 is part of the senescence-associated secretory phenotype (SASP) where it further amplifies the inflammatory state and contributes to the development of age-related diseases by promoting tissue remodelling and dysfunction linking cellular aging to systemic inflammatory processes.

Among the contributors to inflammaging, the gut plays a particularly prominent role. As the splanchnic organ with the highest exposure to exogenous pathogens, it harbours a dynamic microbiome that can serve as a continuous source of inflammatory stimuli. Age-related shifts in microbial composition—particularly the replacement of beneficial commensals with facultative anaerobes—have been associated with increased systemic levels of pro-inflammatory cytokines, including IL-6.³⁷ These changes

can significantly impact homeostatic regulation, especially through cardiovascular and neurological pathways, further reinforcing IL-6's role as a molecular link between chronic inflammation, aging, and disease.^{38,39}

IL-6 is also a key mediator in *metaflammation*, a chronic, low-grade inflammatory state closely linked to metabolic dysfunction. In metabolic diseases such as obesity, type 2 diabetes, and insulin resistance, IL-6 is produced by adipose tissue and muscle cells, where it plays a dual role. While it contributes to the inflammatory milieu that impairs insulin signalling and promotes insulin resistance, it also has adaptive functions in response to metabolic stress, particularly during exercise, where it acts as a myokine to enhance muscle adaptation and overall energy homeostasis. However, in the context of *metaflammation*, the chronic overproduction of IL-6 exacerbates metabolic disturbances, creating a vicious cycle of inflammation and metabolic dysfunction that accelerates disease progression. In the absence of inflammation, the adipose tissue contributes 10–35% of circulating IL-6⁴⁰ with deep implications in body fat modulation. IL-6 gene knockout in mice leads to obesity later in life.⁴¹ Additionally, IL-6 appears to influence fat mass reduction through mechanisms within the central nervous system (CNS).^{41,42} Specifically, its anti-obesity effects in rodents are believed to occur primarily in the brain, particularly in the hypothalamus and hindbrain.⁴³ Research in animal models suggests that IL-6 within the CNS may mediate the reduction of food intake and body weight following activation of glucagon-like peptide-1 (GLP-1) receptors.⁴⁴ Beyond the CNS, IL-6 seems to stimulate GLP-1 production in both the endocrine pancreas and the gut.⁴⁵ This interaction underscores IL-6's central role in both the regulation of metabolic homeostasis and the pathogenesis of metabolic diseases, linking it to both acute and chronic inflammatory responses in a variety of organ systems.

4 | THE ROLE OF IL-6 DURING THE LIFE CYCLE OF AN ATHEROSCLEROTIC PLAQUE

Atherosclerosis is a chronic inflammatory disease driven by various proinflammatory mediators, which contribute to plaque formation and vulnerability.^{46,47} According to a recent meta-analysis, the increase in risk of coronary artery disease per standard deviation of systemic inflammation, defined by either high-sensitivity C-reactive protein (hsCRP) or IL-6, exceeds that of one standard deviation increase in cholesterol or blood pressure.⁴⁸ Aligning with this notion, recent murine studies have shown that the alternative pathway of IL-6 by trans-signalling through pg130 substantially contributes to atherosclerosis.^{49,50} These data

highlight the significance of low, chronic inflammation in increasing the risk of atherosclerotic diseases, possibly surpassing traditional cardiovascular risk factors.

IL-6 is produced by various cell types in response to inflammation, angiotensin II, oxidative stress, and vascular injury.⁵¹ By promoting smooth muscle cell proliferation and migration, endothelial dysfunction, and the recruitment and activation of inflammatory mediators, IL-6 plays a key role in regulating the downstream inflammatory responses that contribute to the development and destabilization of atherosclerotic plaques.^{52,53} IL-6 facilitates macrophage-monocyte activation and migration into the intima, lipid uptake, and low-density lipoprotein (LDL) oxidation.⁵⁴ Furthermore, IL-6 plays a crucial role in promoting a procoagulant state. IL-6 induces thrombocytosis⁵⁵ and facilitates monocyte procoagulant activity by enhancing their mRNA, protein content, and surface expression of tissue factor.^{56,57} Of interest, we recently demonstrated that direct IL-6 inhibition blunts arterial thrombosis by specifically reducing collagen-mediated platelet activation in a mouse model of inflammaging.⁵⁸ Also, IL-6 trans-signalling induces plasminogen activator inhibitor type 1 (PAI-1) in endothelial cells,⁵⁹ thereby playing a crucial role in stabilizing blood clots. At the vessel wall, IL-6 colocalizes with angiotensin II in aortic atherosclerotic plaques,⁶⁰ upregulating cell adhesion molecules,⁶¹ potentiating vascular permeability, and leading to sustained loss of endothelial barrier function.⁶² Anti-IL-6 therapy partially reverses the attenuated endothelial-dependent arteriolar dilatation that develops in diabetic mice knockout for TNF- α (dbTNF⁻/dbTNF⁻).⁶³ Similarly, treatment with the anti-IL-6 receptor antibody MR16-1 has been shown to downregulate inflammation, and reduce atherosclerotic lesion size in LDLr^{-/-} mice.⁶⁴ However, lifetime IL-6 deficiency has been found to be associated with more severe atherosclerosis rather than inhibition of plaque formation, likely due to the disruption of the balance between IL-6 and IL10.⁶⁵

Genome-wide association studies (GWAS) support a potential causal role for the IL-6 pathway in inflammation, atherosclerosis and coronary artery disease. Elevated CRP levels were associated in GWAS with both loci implicated in the immune system (CRP, IL-1 receptor antagonist, IL-6 receptor, IL-1 Family Member 10, Interferon Regulatory Factor 1, and NLRP3) as well as with loci associated with metabolic syndrome (Apolipoprotein C-1, Hepatocyte Nuclear Factor 1, Leptin receptor, Glucokinase Regulatory Protein, Hepatocyte Nuclear Factor 4 Alpha, and Protein Tyrosine Phosphatase, Non-Receptor Type 2).⁶⁶⁻⁶⁸ The IL-6R locus itself associates with coronary artery disease,⁶⁹ while a genetically mediated reduction in IL-6 signalling is associated with longevity and lower risk of atrial fibrillation, stroke and CHD.⁷⁰ Furthermore, Mendelian

randomization analyses show that genetic variation in IL-6 signalling is positively associated with plasma IL-6 receptor (IL-6R) levels and is correlated with lifelong CRP levels and CV risk.^{71,72}

5 | LOOKING AT THE SEQUELAE OF ATHEROSCLEROSIS

5.1 | Acute myocardial infarction

IL-6 plays a highly complex role in the pathophysiology of myocardial infarction (MI), mediating a broad range of inflammatory and cardioprotective/repairative processes that influence cardiac injury, remodelling, and long-term outcomes.

The protective functions of IL-6 are best characterized in ischemia-reperfusion injury and myocardial infarction, both of which induce IL-6 production. Conceptually, cardioprotective cytosolic signalling pathways are divided into three major cardioprotective pathways and one of those is the so-called survival activating factor enhancement (SAFE) pathway.⁷³ In response to IL-6 and IL-6-like cytokines or tumour necrosis factor alpha (TNF) and through sarcolemmal glycoprotein 130 or TNF receptors, Janus kinase (JAK) is activated and phosphorylates signal transducer and activator of transcription (STAT)3.⁷⁴ The strongest and most robust cardioprotective intervention is ischemic conditioning, which is effective in all species tested so far, including humans.^{75,76} Cardioprotection by late local and remote ischemic conditioning involves the canonical function of STAT3^{77,78} and STAT5.⁷⁹ STAT3 is the key protein of the SAFE pathway^{73,80,81} and the non-canonical function of STAT3 plays a unique role in acute cardioprotection. STAT3 is the only cytosolic protein whose activation is consistently associated with a reduction in infarct size by all ischemic conditioning procedures. Pharmacological blockade of STAT3 activation proves its causal involvement in cardioprotection. STAT3 activation has been observed in all species, including larger mammals such as pigs.^{73,76,80,81} The intracellular targets of the SAFE pathway are mitochondria and preservation of mitochondrial function is central for cardiomyocyte function and viability and thus the reduction of ischemia/reperfusion injury.⁸²

Following acute MI, IL-6 levels rise sharply in the plasma, with early elevations detectable upon hospital admission and peaking within the first 48 hours.⁸³ These levels often remain elevated for several weeks to months, paralleling other inflammatory biomarkers such as hs-CRP.⁸³ Basic and translational research has shown that this IL-6 surge reflects its role in orchestrating the immune response to ischemic injury and modulating both

deleterious and protective processes in the infarcted myocardium.⁸⁴ In murine models, single-cell RNA sequencing has identified cardiac fibroblasts as the principal source of IL-6 in the infarcted heart.⁸⁵ These fibroblasts become activated in response to ischemia and hypoxia, and their IL-6 production is finely regulated by purinergic signalling, particularly through adenosine acting on the A2b receptor. Interestingly, this signalling pathway is influenced by T cell-derived adenosine, underscoring a metabolic interplay between immune cells and stromal cells that governs cytokine secretion in the injured heart.⁸⁵ Cardiac myocytes, especially those in the border zone of reperfused infarctions, also contribute to the local production of IL-6.⁸⁶ In vitro, isolated ventricular myocytes have been shown to upregulate IL-6 mRNA in response to pro-inflammatory cytokines such as TNF- α and IL-1 β , as well as postischemic lymph. This response is often accompanied by an increase in intracellular adhesion molecule-1 (ICAM-1) expression, a key mediator in neutrophil-myocyte interactions and subsequent myocyte injury.⁸⁶ Therefore, both fibroblasts and cardiomyocytes actively shape the IL-6-mediated microenvironment following MI, modulated by the broader inflammatory milieu. The chronic IL-6 trigger also activates the Janus kinase (JAK)/STAT pathway, which here however activates the canonical function of STAT, which is central for the deleterious effects.⁸⁷ Moreover, IL-6 also activates the TGF- β 1/Smad3 signalling axis, promoting myocardial fibrosis through enhanced expression of matrix metalloproteinases (MMP-2/9) and collagen deposition.⁸⁸ These fibrotic processes contribute to adverse ventricular remodelling and progressive deterioration in cardiac function.

Despite its well-established role in promoting inflammation, the chronic IL-6 also exerts important protective and regenerative effects. Animal studies have shown that IL-6 overexpression can enhance cardiomyocyte proliferation and reduce apoptosis, in part through upregulation of cyclin D1 and Bcl-2.⁸⁹ The IL-6/sIL-6R complex has been implicated in mobilizing protective signalling during early reperfusion, possibly attenuating myocyte apoptosis and promoting tissue salvage.⁸⁹ These dual roles illustrate the context-dependent nature of IL-6 biology in the heart: while necessary for initiating repair and regeneration, excessive or prolonged IL-6 expression contributes to maladaptive remodelling and functional decline.⁹⁰

In patients, elevated IL-6 levels are associated with poor outcomes following MI, including increased mortality, recurrent infarction, and the development of heart failure.^{13,91-96} Studies have shown a strong correlation between IL-6 levels and infarct size, as well as markers of myocardial necrosis and left ventricular (LV) dysfunction. Importantly, persistent elevation of IL-6 beyond the acute phase is a predictor of adverse ventricular remodelling

and long-term systolic impairment. These findings have led to considerable interest in IL-6 as a prognostic biomarker and as a potential therapeutic target in acute coronary syndromes. In clinical settings, elevated IL-6 levels are also more frequently observed in patients with traditional cardiovascular risk factors such as smoking and hypertension, suggesting that IL-6 may mediate part of their pathogenic effect on MI risk.⁹⁷

IL-6 was then investigated as a potential therapeutic target. Experimental results displayed mixed results.⁹⁸⁻¹⁰¹ In the ASSAIL-MI trial (Assessing the effect of Anti-IL-6 treatment in Myocardial Infarction), a single dose of the anti-IL-6R antibody tocilizumab, resulted in a modest 6% improvement in myocardial salvage index by cardiac magnetic resonance at 3 to 6 days, without significant benefit at 6 months.¹⁰² Nevertheless, tocilizumab significantly reduced microvascular obstruction and CRP levels during the index hospitalization, without affecting the troponin curve. Furthermore, treatment with tocilizumab led to modification of five proteins associated with the acute phase response, including monocyte chemoattraction, and neutrophil function, as identified through aptamer-based proteomics.¹⁰³ This data began to reinforce the potential importance of the IL-6 blockade in CVDs. Similarly, preprocedural administration of colchicine has been found to attenuate the increase in IL-6 and hsCRP in 400 individuals undergoing elective percutaneous coronary intervention.¹⁰⁴ However, no benefits on the risk of percutaneous coronary intervention-related myocardial injury were reported.

This therapeutic ambiguity highlights the need for a nuanced, temporally tailored approach to IL-6 modulation. Rather than broad or continuous inhibition, selective targeting of specific IL-6 signalling pathways—such as trans-signalling via the sIL-6R—may offer a more refined strategy that mitigates harmful inflammation while preserving reparative processes. Furthermore, understanding the timing, cellular sources, and receptor dynamics of IL-6 signalling will be crucial for optimizing its therapeutic modulation. This may also involve combining IL-6-targeting therapies with agents that modulate fibrosis, immune cell recruitment, or metabolic responses to injury.

5.2 | Cardiac fibrosis and heart failure

IL-6 is also critically involved in the pathophysiology of cardiac fibrosis (CF) and HF, acting as a potent regulator of inflammatory and remodelling processes in the myocardium.¹⁰⁵ In CF, regardless of aetiology, fibroblast activation and migration, matrix metalloproteinase activation, and accumulation of collagen,

glycoproteins, and proteoglycans lead to scar formation and stabilization.^{106,107} IL-6 promotes fibrosis via MAPK- and CAMKII-STAT3-dependent pathways and acts downstream of hypoxia-induced mitogenic factor, with its inhibition preventing fibroblast activation.^{108,109} IL-6 overexpression amplifies TGF- β 1-mediated MMP2/MMP3 signalling, enhancing myofibroblast proliferation, differentiation, and fibrotic matrix deposition.¹¹⁰ Experimental models demonstrate that IL-6 knockout mice develop less CF,¹¹¹ preserve better cardiac function, and exhibit reduced cardiomyocyte apoptosis in dilated cardiomyopathy through STAT3 inhibition.¹¹² Fibroblast-derived IL-6 directly activates STAT3 following myocardial infarction,¹¹³ and its genetic deletion attenuates cardiac hypertrophy and fibrosis after angiotensin II stimulation.¹¹⁴

In HF, inflammatory biomarkers such as hsCRP and IL-6 predict incident HF, adverse remodelling, and worse outcomes despite optimal medical therapy.^{115–119} In the BIOSTAT-CHF cohort, elevated IL-6 levels predicted HF hospitalization, cardiovascular mortality, and all-cause mortality, and correlated with NT-proBNP, procalcitonin, TNF- α , IL-1, and hepcidin,¹²⁰ the latter—together with IL-6—being a major regulator of systemic iron metabolism and iron-deficiency anaemia commonly associated with HF.¹²¹ The CANTOS trial provided proof-of-concept that cytokine inhibition can improve HF outcomes: higher baseline IL-6 predicted a two-fold increased risk of HF hospitalization, and canakinumab reduced incident HF events in a dose-dependent manner, with the greatest benefit in patients achieving the largest reductions in inflammation.¹²² IL-6 blockade emerges as a promising strategy to attenuate CF across HF phenotypes, including HF with preserved ejection fraction.¹²³ Furthermore, standard HF therapies— β -blockers, renin-angiotensin-aldosterone system inhibitors, mineralocorticoid receptor antagonists, and sodium-glucose cotransporter 2 inhibitors—possess anti-inflammatory properties that may contribute to their capacity to counteract CF, promote reverse remodelling, improve endothelial barrier function, and reduce inflammatory cell recruitment by modulating NF- κ B signalling, monocyte chemoattractant protein-1, TNF- α , IL-6, hsCRP, NLRP3 inflammasome, and adhesion molecules such as ICAM-1, VCAM-1, P- and E-selectin.^{124–127}

6 | INTERLEUKIN-6 IN THE CARDIO-RENAL CONTINUUM

The kidney is both a target and a potent amplifier of systemic inflammation, with IL-6 playing a role in the bidirectional cardio-renal interaction. In the setting of chronic

kidney disease (CKD), IL-6 is produced locally by mesangial cells, podocytes, and tubular epithelial cells in response to ischemic, metabolic, or hemodynamic stress.^{128–132} These cells express both membrane-bound and soluble IL-6 receptors, allowing for classical and trans-signalling activation.¹³⁰ The downstream JAK/STAT3, MAPK, and NF- κ B pathways drive the expression of adhesion molecules, monocyte chemoattractant protein-1, and profibrotic mediators such as collagen I, PAI-1, and TGF- β , promoting endothelial dysfunction, mesangial proliferation, and tubulointerstitial fibrosis.^{133–135} Experimental models of hypertensive, diabetic, and ischemia-reperfusion nephropathy demonstrate that IL-6 blockade or genetic deletion attenuates macrophage infiltration, fibrotic remodelling, and albuminuria, underscoring its causal role in progressive renal injury.^{128,136–141}

Clinically, IL-6 levels increase in parallel with the severity of renal dysfunction and independently predict accelerated decline in estimated glomerular filtration rate (eGFR) and progression to end-stage kidney disease.^{142–144} In the Chronic Renal Insufficiency Cohort (CRIC) study and other longitudinal cohorts, individuals in the highest IL-6 quartile had approximately 40–60% higher risk of CKD progression and cardiovascular death compared with those in the lowest quartile, even after adjustment for traditional risk factors and CRP.^{145–148} IL-6 also contributes to the anaemia of CKD via induction of hepatic hepcidin, which suppresses intestinal iron absorption and promotes iron sequestration within macrophages, further enhancing oxidative stress and tissue hypoxia.^{149,150} Elevated IL-6 concentrations in CKD are strongly associated with vascular stiffness, coronary calcification, and left-ventricular hypertrophy, reflecting its capacity to extend renal inflammation to systemic vascular and cardiac compartments.^{144,151–153} This mechanistic link defines a “cardio-renal inflammatory loop”, in which renal dysfunction fuels systemic IL-6 production, and IL-6 in turn aggravates vascular injury and cardiac remodelling. The consequence is a state of chronic low-grade inflammation that accelerates atherosclerosis, thrombosis, and progression of heart failure. Importantly, in patients with CKD or on dialysis, IL-6 and hsCRP outperform LDL-cholesterol as predictors of adverse cardiovascular events, highlighting the predominant role of inflammation over lipid burden in this population.^{154,155} Furthermore, IL-6 levels predict responsiveness to erythropoiesis-stimulating agents (ESA) and long-term mortality, further supporting its pathophysiological centrality. Indeed, in a phase 1/2 randomized, placebo-controlled trial, ziltivekimab administered intravenously every 2 weeks for 12 weeks led to dose-dependent reductions in ESA usage, ESA resistance index, and increases in serum iron, transferrin saturation, and albumin, without dose-limiting toxicity.¹⁵⁶

Given this convergence of renal and cardiovascular inflammation, IL-6 signalling has emerged as a compelling therapeutic target in CKD. By interrupting the JAK/STAT3 axis and reducing hepatic acute-phase protein synthesis, IL-6 inhibition could mitigate systemic inflammation, preserve renal function, and reduce atherothrombotic risk. This rationale underpins ongoing trials testing IL-6 blockade in CKD and dialysis populations (detailed in the upcoming paragraph), where suppression of IL-6-mediated inflammation may help to break the vicious cycle linking kidney injury, vascular dysfunction, and cardiovascular death.

7 | THE OLYMPIAN ERA OF ANTI-IL-6 TRIALS

Over the past decade, increasing mechanistic and clinical evidence has positioned IL-6 as a central driver of residual inflammatory risk in cardiovascular disease, with promising implications for targeted therapy. The proof-of-concept that modulating the IL-1 β -IL-6 signalling axis can reduce CV event rates was established by CANTOS, which demonstrated that lowering downstream IL-6 levels through IL-1 β inhibition significantly reduced major adverse cardiovascular events (MACE).^{12,13} However,

TABLE 1 Ongoing Phase III trial for IL-6 inhibiting agents with cardiovascular outcomes.

	NCT ID	Estimated study completion	Estimated enrollment	IL-6 inhibitor	Population	Endpoints
ZEUS	NCT05021835	June 2026	6200	Ziltivekimab	Patients with established atherosclerotic cardiovascular disease, chronic kidney disease and systemic inflammation	Composite endpoint consisting of cardiovascular death, non-fatal myocardial infarction and non-fatal stroke
HERMES	NCT05636176	July 2027	5600	Ziltivekimab	Patients with heart failure with mildly reduced or preserved ejection fraction and systemic inflammation	Composite heart failure endpoint consisting of cardiovascular death, heart failure hospitalization or urgent HF visit
ATHENA	NCT06200207	January 2027	680	Ziltivekimab	Patients with heart failure with mildly reduced or preserved ejection fraction and systemic inflammation	Change in Kansas City Cardiomyopathy Questionnaire (KCCQ) clinical summary score (KCCQ-CSS)
ARTEMIS	NCT06118281	September 2026	10,000	Ziltivekimab	Patients with acute myocardial infarction	Composite endpoint consisting of cardiovascular death, non-fatal myocardial infarction, non-fatal stroke
POSIBIL6ESKD	NCT05485961	August 2029	2310	Clazakizumab (CSL 300)	Patients with end-stage kidney disease undergoing dialysis	Composite endpoint consisting of cardiovascular death or myocardial infarction

IL-1 β antagonism is associated with high costs and potential adverse effects, and canakinumab is not approved for CV prevention, which has shifted attention toward directly inhibiting IL-6 as a potentially more efficient and specific strategy (Table 1).

Patients with CKD represent an especially compelling target population for IL-6-directed interventions. CKD confers a high absolute CV risk, is characterized by persistent systemic inflammation, and limits the use of other anti-inflammatory therapies such as colchicine due to renal excretion.¹⁵⁷ In this context, the phase IIb RESCUE trial evaluated the IL-6 ligand-blocking monoclonal antibody ziltivekimab in patients with CKD and elevated hsCRP. Treatment led to an approximately 80% reduction in hsCRP to levels below 2 mg/L, a magnitude of inflammatory lowering nearly double that achieved in CANTOS, without adversely affecting lipid profiles or causing significant cytopenias or injection-site reactions over 24 weeks.¹⁵⁸ Although limited by sample size and duration, these findings provided the rationale for advancing ziltivekimab into multiple phase III cardiovascular outcome trials.

The largest of these, ZEUS (Ziltivekimab Cardiovascular Outcomes Study; NCT05021835), is enrolling 6200 patients with stage 3–4 CKD and elevated hsCRP to determine whether chronic IL-6 inhibition reduces the incidence of MI, stroke, and CV death. Secondary objectives include assessing the effect of ziltivekimab on renal disease progression, measured by estimated glomerular filtration rate decline, changes in urinary albumin-to-creatinine ratio, and development of end-stage renal disease. The trial will also stratify outcomes by baseline left ventricular ejection fraction, enabling analysis of effects across HF phenotypes.

Beyond CKD, IL-6 inhibition is being tested in HF populations with elevated inflammatory activity, where it is hypothesized to mitigate adverse remodelling by reducing CF, microvascular dysfunction, and systemic inflammation.¹⁵⁹ The HERMES trial (A Research Study to Look at How Ziltivekimab Works Compared to Placebo in People With Heart Failure and Inflammation; NCT05636176) is a phase III, randomized, double-blind, placebo-controlled study enrolling approximately 5600 patients with HF with preserved ejection fraction (HFpEF) or mildly reduced ejection fraction (HFmrEF), elevated NT-proBNP, echocardiographic evidence of structural or functional abnormalities, and hsCRP \geq 2 mg/L. Participants receive monthly 15 mg ziltivekimab or placebo in addition to standard care, with the primary endpoint being time to first CV death, HF hospitalization, or urgent HF visit.

Two additional phase III studies are expanding the evaluation of ziltivekimab. ATHENA (A Research Study Looking Into How Ziltivekimab Works Compared

to Placebo in Participants With Heart Failure and Inflammation, NCT06200207) enrolls 680 patients with HFpEF and systemic inflammation (hsCRP \geq 2 mg/L) to test whether IL-6 blockade can improve patient-reported quality of life, assessed by the Kansas City Cardiomyopathy Questionnaire clinical summary score over 12 months. Meanwhile, the ARTEMIS trial (A Research Study to Look at How Ziltivekimab Works Compared to Placebo in People With a Heart Attack, NCT06118281) is designed to evaluate the efficacy and safety of ziltivekimab in approximately 10,000 patients with recent ACS. Patients are enrolled within 14 days post-ACS and randomized to receive monthly subcutaneous injections of ziltivekimab at doses ranging from 7.5 to 30 mg, or placebo, in addition to standard of care. The primary endpoint is a composite of MACE, including CV death, nonfatal MI, and stroke. By targeting IL-6-mediated inflammation early in the post-infarction period, ARTEMIS aims to assess whether IL-6 inhibition can reduce residual inflammatory risk and improve long-term CV outcomes in this high-risk population.

While most current phase III programs focus on ziltivekimab, other IL-6-targeting agents are advancing in selected high-risk groups. Clazakizumab, an IL-6 ligand inhibitor, is under investigation in the phase III POSIBIL6ESKD trial (Combined Dose-Finding and CV Outcomes Study With CSL300 (Clazakizumab) in Adult Subjects With ESKD Undergoing Dialysis, NCT05485961), enrolling patients with end-stage kidney disease, diabetes, or established atherosclerotic CVD and elevated hsCRP. The primary endpoint is time to CV death or MI, with long-term follow-up planned to assess safety in this complex and underserved population.

Together, these ongoing phase III studies will address whether direct IL-6 inhibition can safely and effectively reduce CV events across different high-risk groups. If successful, IL-6-targeted therapy could represent the first precision anti-inflammatory approach broadly applicable in CV prevention, offering a new therapeutic pillar alongside lipid-lowering, antithrombotic, and neurohormonal modulation strategies. While most ongoing trials focus on cardiovascular outcomes, IL-6 blockade may have broader implications. Preclinical and translational studies suggest that IL-6 modulation could benefit other organs commonly affected by chronic inflammatory stress, including the kidney and metabolic tissues. Beyond attenuating vascular inflammation, IL-6 inhibition may help preserve renal function and improve metabolic homeostasis through indirect effects on insulin sensitivity, lipid metabolism, and hepatic acute-phase responses. These hypotheses remain to be confirmed in dedicated clinical studies, but collectively they highlight the potential of IL-6-targeted therapy as a strategy to address residual inflammatory risk in a multi-organ context.

8 | CONCLUSION

IL-6 is a prime driver of cardiovascular inflammation, linking systemic inflammation to vascular dysfunction, atherosclerosis, myocardial fibrosis, and adverse cardiac remodelling. Yet, IL-6 signalling exerts context-dependent effects: transient, classic signalling via the membrane-bound IL-6 receptor promotes tissue repair, metabolic adaptation, and cardioprotection in acute injury or infection, whereas persistent or excessive activation (particularly through trans-signalling) triggers endothelial activation, immune cell recruitment, and chronic inflammatory damage. Thus, the net impact of IL-6 depends on timing, receptor engagement, signalling pathway, and cellular context, highlighting its double-edged role in cardiovascular pathophysiology and other organ systems such as the kidney. Epidemiological, genetic, and mechanistic evidence supports a causal role of IL-6 in CV health and disease. Early clinical studies indicate that IL-6 inhibition can substantially lower systemic inflammatory markers with an acceptable short-term safety profile, but the key question remains whether such interventions can translate into improved cardiovascular outcomes, and which patient subgroups are most likely to benefit. Future strategies will need to modulate IL-6 activity with precision, attenuating its pathogenic effects while preserving its homeostatic and reparative functions, to complement existing therapies and address residual inflammatory risk in CVD.

AUTHOR CONTRIBUTIONS

AP, AT, and GL drafted the first version of the manuscript and provided the figures and the table. LL conceptualized the article and revised the first draft. All authors provided critical feedback, helped to shape the manuscript and approved the final version.

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CONFLICT OF INTEREST STATEMENT

LL and GGC are coinventors on the International Patent WO/2020/226993 filed in April 2020; the patent relates to the use of antibodies which specifically bind interleukin-1a to reduce various sequelae of ischemia–reperfusion injury to the central nervous system. LL has received

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DATA AVAILABILITY STATEMENT

No new data is presented in this work.

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REFERENCES

1. Soysal P, Stubbs B, Lucato P, et al. Inflammation and frailty in the elderly: a systematic review and meta-analysis. *Ageing Res Rev.* 2016;31:1-8. doi:10.1016/j.arr.2016.08.006
2. Liberale L, Tual-Chalot S, Sedej S, et al. Roadmap for alleviating the manifestations of ageing in the cardiovascular system. *Nat Rev Cardiol.* 2025;22:577-605. doi:10.1038/s41569-025-01130-5
3. Liberale L, Badimon L, Montecucco F, Lüscher TF, Libby P, Camici GG. Inflammation, aging, and cardiovascular disease: JACC review topic of the week. *J Am Coll Cardiol.* 2022;79:837-847. doi:10.1016/j.jacc.2021.12.017
4. Liberale L, Montecucco F, Tardif JC, Libby P, Camici GG. Inflamm-aging: the role of inflammation in age-dependent cardiovascular disease. *Eur Heart J.* 2020;41:2974-2982. doi:10.1093/eurheartj/ehz961
5. Puzianowska-Kuźnicka M, Owczarż M, Wieczorowska-Tobis K, et al. Interleukin-6 and C-reactive protein, successful aging,

- and mortality: the PolSenior study. *Immun Ageing*. 2016;13:21. doi:10.1186/s12979-016-0076-x
6. Vilahur G, Badimon L. Biological actions of pentraxins. *Vasc Pharmacol*. 2015;73:38-44. doi:10.1016/j.vph.2015.05.001
 7. Khan MS, Talha KM, Maqsood MH, et al. Interleukin-6 and cardiovascular events in healthy adults: MESA. *JACC Adv*. 2024;3:101063. doi:10.1016/j.jacadv.2024.101063
 8. Liberale L, Montecucco F, Schwarz L, Luscher TF, Camici GG. Inflammation and cardiovascular diseases: lessons from seminal clinical trials. *Cardiovasc Res*. 2021;117:411-422. doi:10.1093/cvr/cvaa211
 9. Kraler S, Wenzl FA, Luscher TF. Repurposing colchicine to combat residual cardiovascular risk: the LoDoCo2 trial. *Eur J Clin Invest*. 2020;50:e13424. doi:10.1111/eci.13424
 10. Bonaventura A, Potere N, Liberale L, Kraler S, Weber BN, Abbate A. Colchicine in coronary artery disease: where do we stand? *J Cardiovasc Pharmacol*. 2025;85:243-247. doi:10.1097/FJC.0000000000001672
 11. Kraler S, Mueller C, Libby P, Bhatt DL. Acute coronary syndromes: mechanisms, challenges, and new opportunities. *Eur Heart J*. 2025;46:2866-2889. doi:10.1093/eurheartj/ehaf289
 12. Ridker PM, Everett BM, Thuren T, et al. Antiinflammatory therapy with Canakinumab for atherosclerotic disease. *N Engl J Med*. 2017;377:1119-1131. doi:10.1056/NEJMoa1707914
 13. Ridker PM, Libby P, MacFadyen JG, et al. Modulation of the interleukin-6 signalling pathway and incidence rates of atherosclerotic events and all-cause mortality: analyses from the Canakinumab anti-inflammatory thrombosis outcomes study (CANTOS). *Eur Heart J*. 2018;39:3499-3507. doi:10.1093/eurheartj/ehy310
 14. Ridker PM. From RESCUE to ZEUS: will interleukin-6 inhibition with ziltivekimab prove effective for cardiovascular event reduction? *Cardiovasc Res*. 2021;117:e138-e140. doi:10.1093/cvr/cvab231
 15. Petrie M, Borlaug B, Buchholtz K, et al. HERMES: effects of Ziltivekimab versus placebo on morbidity and mortality in patients with heart failure with mildly reduced or preserved ejection fraction and systemic inflammation. *J Card Fail*. 2024;30:126. doi:10.1016/j.cardfail.2023.10.024
 16. Regulation of the acute phase and immune responses: interleukin-6. *Ann NY Acad Sci*. 1989;557:1-583.
 17. Su JH, Luo MY, Liang N, et al. Interleukin-6: a novel target for cardio-cerebrovascular diseases. *Front Pharmacol*. 2021;12:745061. doi:10.3389/fphar.2021.745061
 18. Ghanemi A, St-Amand J. Interleukin-6 as a "metabolic hormone". *Cytokine*. 2018;112:132-136. doi:10.1016/j.cyt.2018.06.034
 19. Kushner I. Regulation of the acute phase response by cytokines. *Perspect Biol Med*. 1993;36:611-622. doi:10.1353/pbm.1993.0004
 20. Kaplanski G, Marin V, Montero-Julian F, Mantovani A, Farnarier C. IL-6: a regulator of the transition from neutrophil to monocyte recruitment during inflammation. *Trends Immunol*. 2003;24:25-29. doi:10.1016/s1471-4906(02)00013-3
 21. Simpson RJ, Hammacher A, Smith DK, Matthews JM, Ward LD. Interleukin-6: structure-function relationships. *Protein Sci*. 1997;6:929-955. doi:10.1002/pro.5560060501
 22. Bazan JF. Structural design and molecular evolution of a cytokine receptor superfamily. *Proc Natl Acad Sci USA*. 1990;87:6934-6938. doi:10.1073/pnas.87.18.6934
 23. Hibi M, Murakami M, Saito M, Hirano T, Taga T, Kishimoto T. Molecular cloning and expression of an IL-6 signal transducer, gp130. *Cell*. 1990;63:1149-1157. doi:10.1016/0092-8674(90)90411-7
 24. Lacroix M, Rousseau F, Guilhot F, et al. Novel insights into interleukin 6 (IL-6) cis- and trans-signaling pathways by differentially manipulating the assembly of the IL-6 signaling complex. *J Biol Chem*. 2015;290:26943-26953. doi:10.1074/jbc.M115.682138
 25. Kaur S, Bansal Y, Kumar R, Bansal G. A panoramic review of IL-6: structure, pathophysiological roles and inhibitors. *Bioorg Med Chem*. 2020;28:115327. doi:10.1016/j.bmc.2020.115327
 26. Masjedi A, Hashemi V, Hojjat-Farsangi M, et al. The significant role of interleukin-6 and its signaling pathway in the immunopathogenesis and treatment of breast cancer. *Biomed Pharmacother*. 2018;108:1415-1424. doi:10.1016/j.biopha.2018.09.177
 27. Garbers C, Heink S, Korn T, Rose-John S. Interleukin-6: designing specific therapeutics for a complex cytokine. *Nat Rev Drug Discov*. 2018;17:395-412. doi:10.1038/nrd.2018.45
 28. Rose-John S. Interleukin-6 signalling in health and disease. *F1000Res*. 2020;9:1013. doi:10.12688/f1000research.26058.1
 29. Christoffersen M, Tybjaerg-Hansen A. Targeting IL-6 in patients at high cardiovascular risk. *Lancet*. 2021;397:2025-2027. doi:10.1016/S0140-6736(21)00985-5
 30. Fonseca JE, Santos MJ, Canhão H, Choy E. Interleukin-6 as a key player in systemic inflammation and joint destruction. *Autoimmun Rev*. 2009;8:538-542. doi:10.1016/j.autrev.2009.01.012
 31. Kumari N, Dwarakanath BS, Das A, Bhatt AN. Role of interleukin-6 in cancer progression and therapeutic resistance. *Tumour Biol*. 2016;37:11553-11572. doi:10.1007/s13277-016-5098-7
 32. Rothaug M, Becker-Pauly C, Rose-John S. The role of interleukin-6 signaling in nervous tissue. *Biochim Biophys Acta*. 2016;1863:1218-1227. doi:10.1016/j.bbamcr.2016.03.018
 33. Raj DS. Role of interleukin-6 in the anemia of chronic disease. *Semin Arthritis Rheum*. 2009;38:382-388. doi:10.1016/j.semarthrit.2008.01.006
 34. Shahini A, Shahini A. Role of interleukin-6-mediated inflammation in the pathogenesis of inflammatory bowel disease: focus on the available therapeutic approaches and gut microbiome. *J Cell Commun Signal*. 2023;17:55-74. doi:10.1007/s12079-022-00695-x
 35. Rincon M, Irvin CG. Role of IL-6 in asthma and other inflammatory pulmonary diseases. *Int J Biol Sci*. 2012;8:1281-1290. doi:10.7150/ijbs.4874
 36. Franceschi C, Capri M, Monti D, et al. Inflammaging and anti-inflammaging: a systemic perspective on aging and longevity emerged from studies in humans. *Mech Ageing Dev*. 2007;128:92-105. doi:10.1016/j.mad.2006.11.016
 37. Wang N, Li R, Lin H, et al. Enriched taxa were found among the gut microbiota of centenarians in East China. *PLoS One*. 2019;14:e0222763. doi:10.1371/journal.pone.0222763
 38. Iannone LF, Preda A, Blottière HM, et al. Microbiota-gut brain axis involvement in neuropsychiatric disorders. *Expert Rev Neurother*. 2019;19:1037-1050. doi:10.1080/14737175.2019.1638763
 39. Tang WH, Kitai T, Hazen SL. Gut microbiota in cardiovascular health and disease. *Circ Res*. 2017;120:1183-1196. doi:10.1161/circresaha.117.309715

40. Faldt J, Wernstedt I, Fitzgerald SM, et al. Reduced exercise endurance in interleukin-6-deficient mice. *Endocrinology*. 2004;145:2680-2686. doi:10.1210/en.2003-1319
41. Wallenius V, Wallenius K, Ahrén B, et al. Interleukin-6-deficient mice develop mature-onset obesity. *Nat Med*. 2002;8:75-79. doi:10.1038/nm0102-75
42. Di Gregorio GB, Hensley L, Lu T, Ranganathan G, Kern PA. Lipid and carbohydrate metabolism in mice with a targeted mutation in the IL-6 gene: absence of development of age-related obesity. *Am J Physiol Endocrinol Metab*. 2004;287:E182-E187. doi:10.1152/ajpendo.00189.2003
43. Campbell IL, Erta M, Lim SL, et al. Trans-signaling is a dominant mechanism for the pathogenic actions of interleukin-6 in the brain. *J Neurosci*. 2014;34:2503-2513. doi:10.1523/JNEUROSCI.2830-13.2014
44. Shirazi R, Palsdottir V, Collander J, et al. Glucagon-like peptide 1 receptor induced suppression of food intake, and body weight is mediated by central IL-1 and IL-6. *Proc Natl Acad Sci USA*. 2013;110:16199-16204. doi:10.1073/pnas.1306799110
45. Ellingsgaard H, Hauselmann I, Schuler B, et al. Interleukin-6 enhances insulin secretion by increasing glucagon-like peptide-1 secretion from L cells and alpha cells. *Nat Med*. 2011;17:1481-1489. doi:10.1038/nm.2513
46. Ross R. Atherosclerosis--an inflammatory disease. *N Engl J Med*. 1999;340:115-126. doi:10.1056/nejm199901143400207
47. Badimon L, Vilahur G. Thrombosis formation on atherosclerotic lesions and plaque rupture. *J Intern Med*. 2014;276:618-632. doi:10.1111/joim.12296
48. Kaptoge S, Di Angelantonio E, Lowe G, et al. C-reactive protein concentration and risk of coronary heart disease, stroke, and mortality: an individual participant meta-analysis. *Lancet*. 2010;375:132-140. doi:10.1016/s0140-6736(09)61717-7
49. Luchtefeld M, Schunkert H, Stoll M, et al. Signal transducer of inflammation gp130 modulates atherosclerosis in mice and man. *J Exp Med*. 2007;204:1935-1944. doi:10.1084/jem.20070120
50. Schuett H, Oestreich R, Waetzig GH, et al. Transsignaling of interleukin-6 crucially contributes to atherosclerosis in mice. *Arterioscler Thromb Vasc Biol*. 2012;32:281-290. doi:10.1161/atvbaha.111.229435
51. Hartman J, Frishman WH. Inflammation and atherosclerosis: a review of the role of interleukin-6 in the development of atherosclerosis and the potential for targeted drug therapy. *Cardiol Rev*. 2014;22:147-151. doi:10.1097/crd.0000000000000021
52. Schuett H, Luchtefeld M, Grothusen C, Grote K, Schieffer B. How much is too much? Interleukin-6 and its signalling in atherosclerosis. *Thromb Haemost*. 2009;102:215-222. doi:10.1160/th09-05-0297
53. Wassmann S, Stumpf M, Strehlow K, et al. Interleukin-6 induces oxidative stress and endothelial dysfunction by overexpression of the angiotensin II type 1 receptor. *Circ Res*. 2004;94:534-541. doi:10.1161/01.Res.0000115557.25127.8d
54. Huber SA, Sakkinen P, Conze D, Hardin N, Tracy R. Interleukin-6 exacerbates early atherosclerosis in mice. *Arterioscler Thromb Vasc Biol*. 1999;19:2364-2367. doi:10.1161/01.atv.19.10.2364
55. Ishibashi T, Kimura H, Shikama Y, et al. Interleukin-6 is a potent thrombopoietic factor in vivo in mice. *Blood*. 1989;74:1241-1244.
56. Neumann FJ, Ott I, Marx N, et al. Effect of human recombinant interleukin-6 and interleukin-8 on monocyte procoagulant activity. *Arterioscler Thromb Vasc Biol*. 1997;17:3399-3405. doi:10.1161/01.atv.17.12.3399
57. Mitani H, Katayama N, Araki H, et al. Activity of interleukin 6 in the differentiation of monocytes to macrophages and dendritic cells. *Br J Haematol*. 2000;109:288-295. doi:10.1046/j.1365-2141.2000.02020.x
58. Ministrini S, Liberale L, Puspitasari YM, et al. Direct Interleukin-6 inhibition blunts arterial thrombosis by reducing collagen-mediated platelet activation. *Arterioscler Thromb Vasc Biol*. 2025;45:1432-1439. doi:10.1161/ATVBAHA.125.322533
59. Kang S, Tanaka T, Inoue H, et al. IL-6 trans-signaling induces plasminogen activator inhibitor-1 from vascular endothelial cells in cytokine release syndrome. *Proc Natl Acad Sci USA*. 2020;117:22351-22356. doi:10.1073/pnas.2010229117
60. Recinos A, LeJeune WS, Sun H, et al. Angiotensin II induces IL-6 expression and the Jak-STAT3 pathway in aortic adventitia of LDL receptor-deficient mice. *Atherosclerosis*. 2007;194:125-133. doi:10.1016/j.atherosclerosis.2006.10.013
61. Romano M, Sironi M, Toniatti C, et al. Role of IL-6 and its soluble receptor in induction of chemokines and leukocyte recruitment. *Immunity*. 1997;6:315-325. doi:10.1016/s1074-7613(00)80334-9
62. Alsaffar H, Martino N, Garrett JP, Adam AP. Interleukin-6 promotes a sustained loss of endothelial barrier function via Janus kinase-mediated STAT3 phosphorylation and de novo protein synthesis. *Am J Physiol Cell Physiol*. 2018;314:C589-c602. doi:10.1152/ajpcell.00235.2017
63. Lee J, Lee S, Zhang H, Hill MA, Zhang C, Park Y. Interaction of IL-6 and TNF- α contributes to endothelial dysfunction in type 2 diabetic mouse hearts. *PLoS One*. 2017;12:e0187189. doi:10.1371/journal.pone.0187189
64. Akita K, Isoda K, Sato-Okabayashi Y, et al. An Interleukin-6 receptor antibody suppresses atherosclerosis in Atherogenic mice. *Front Cardiovasc Med*. 2017;4:84. doi:10.3389/fcvm.2017.00084
65. Schieffer B, Selle T, Hilfiker A, et al. Impact of interleukin-6 on plaque development and morphology in experimental atherosclerosis. *Circulation*. 2004;110:3493-3500. doi:10.1161/01.CIR.0000148135.08582.97
66. Ridker PM, Pare G, Parker A, et al. Loci related to metabolic-syndrome pathways including LEPR, HNF1A, IL6R, and GCKR associate with plasma C-reactive protein: the Women's genome health study. *Am J Hum Genet*. 2008;82:1185-1192. doi:10.1016/j.ajhg.2008.03.015
67. Reiner AP, Beleza S, Franceschini N, et al. Genome-wide association and population genetic analysis of C-reactive protein in African American and Hispanic American women. *Am J Hum Genet*. 2012;91:502-512. doi:10.1016/j.ajhg.2012.07.023
68. Dehghan A, Dupuis J, Barbalic M, et al. Meta-analysis of genome-wide association studies in >80,000 subjects identifies multiple loci for C-reactive protein levels. *Circulation*. 2011;123:731-738. doi:10.1161/circulationaha.110.948570
69. van der Harst P, Verweij N. Identification of 64 novel genetic loci provides an expanded view on the genetic architecture of coronary artery disease. *Circ Res*. 2018;122:433-443. doi:10.1161/circresaha.117.312086
70. Rosa M, Chignon A, Li Z, et al. A Mendelian randomization study of IL6 signaling in cardiovascular diseases, immune-related disorders and longevity. *NPJ Genom Med*. 2019;4:23. doi:10.1038/s41525-019-0097-4
71. Sarwar N, Butterworth AS, Freitag DF, et al. Interleukin-6 receptor pathways in coronary heart disease: a collaborative

- meta-analysis of 82 studies. *Lancet*. 2012;379:1205-1213. doi:[10.1016/s0140-6736\(11\)61931-4](https://doi.org/10.1016/s0140-6736(11)61931-4)
72. Swerdlow DI, Holmes MV, Kuchenbaecker KB, et al. The interleukin-6 receptor as a target for prevention of coronary heart disease: a mendelian randomisation analysis. *Lancet*. 2012;379:1214-1224. doi:[10.1016/s0140-6736\(12\)60110-x](https://doi.org/10.1016/s0140-6736(12)60110-x)
 73. Lecour S. Activation of the protective survivor activating factor enhancement (SAFE) pathway against reperfusion injury: does it go beyond the RISK path? *J Mol Cell Cardiol*. 2009;47:32-40. doi:[10.1016/j.yjmcc.2009.03.019](https://doi.org/10.1016/j.yjmcc.2009.03.019)
 74. Harhous Z, Booz GW, Ovize M, Bidaux G, Kurdi M. An update on the multifaceted roles of STAT3 in the heart. *Front Cardiovasc Med*. 2019;6:150. doi:[10.3389/fcvm.2019.00150](https://doi.org/10.3389/fcvm.2019.00150)
 75. Heusch G, Rassaf T. Time to give up on cardioprotection? A critical appraisal of clinical studies on ischemic pre-, post-, and remote conditioning. *Circ Res*. 2016;119:676-695. doi:[10.1161/CIRCRESAHA.116.308736](https://doi.org/10.1161/CIRCRESAHA.116.308736)
 76. Heusch G. Myocardial ischaemia-reperfusion injury and cardioprotection in perspective. *Nat Rev Cardiol*. 2020;17:773-789. doi:[10.1038/s41569-020-0403-y](https://doi.org/10.1038/s41569-020-0403-y)
 77. Xuan Y-T, Guo Y, Zhu Y, et al. Role of the protein kinase C-e-Raf-1-MEK-1/2-p44/42 MAPK signaling cascade in the activation of signal transducers and activators of transcription 1 and 3 and induction of cyclooxygenase-2 after ischemic preconditioning. *Circulation*. 2005;112:1971-1978. doi:[10.1161/CIRCULATIONAHA.105.561522](https://doi.org/10.1161/CIRCULATIONAHA.105.561522)
 78. Bolli R, Stein AB, Guo Y, et al. A murine model of inducible, cardiac-specific deletion of STAT3: its use to determine the role of STAT3 in the upregulation of cardioprotective proteins by ischemic preconditioning. *J Mol Cell Cardiol*. 2011;50:589-597. doi:[10.1016/j.yjmcc.2011.01.002](https://doi.org/10.1016/j.yjmcc.2011.01.002)
 79. Chen H, Jing XY, Shen YJ, et al. Stat5-dependent cardioprotection in late remote ischaemia preconditioning. *Cardiovasc Res*. 2018;114:679-689. doi:[10.1093/cvr/cvy014](https://doi.org/10.1093/cvr/cvy014)
 80. Boengler K, Hilfiker-Kleiner D, Drexler H, Heusch G, Schulz R. The myocardial JAK/STAT pathway: from protection to failure. *Pharmacol Ther*. 2008;120:172-185. doi:[10.1016/j.pharmthera.2008.08.002](https://doi.org/10.1016/j.pharmthera.2008.08.002)
 81. Kleinbongard P, Skyschally A, Heusch G. Cardioprotection by remote ischemic conditioning and its signal transduction. *Pflugers Arch - Eur J Physiol*. 2017;469:159-181. doi:[10.1007/s00424-016-1922-6](https://doi.org/10.1007/s00424-016-1922-6)
 82. Boengler K, Eickelmann C, Kleinbongard P. Mitochondrial kinase signaling for cardioprotection. *Int J Mol Sci*. 2024;25:4491. doi:[10.3390/ijms25084491](https://doi.org/10.3390/ijms25084491)
 83. Gabriel AS, Martinsson A, Wretling B, Ahnve S. IL-6 levels in acute and post myocardial infarction: their relation to CRP levels, infarction size, left ventricular systolic function, and heart failure. *Eur J Intern Med*. 2004;15:523-528. doi:[10.1016/j.ejim.2004.07.013](https://doi.org/10.1016/j.ejim.2004.07.013)
 84. Vilahur G, Hernández-Vera R, Molins B, et al. Short-term myocardial ischemia induces cardiac modified C-reactive protein expression and proinflammatory gene (cyclo-oxygenase-2, monocyte chemoattractant protein-1, and tissue factor) up-regulation in peripheral blood mononuclear cells. *J Thromb Haemost*. 2009;7:485-493. doi:[10.1111/j.1538-7836.2008.03244.x](https://doi.org/10.1111/j.1538-7836.2008.03244.x)
 85. Alter C, Henseler AS, Owenier C, et al. IL-6 in the infarcted heart is preferentially formed by fibroblasts and modulated by purinergic signaling. *J Clin Invest*. 2023;133:e163799. doi:[10.1172/JCI163799](https://doi.org/10.1172/JCI163799)
 86. Gwechenberger M, Mendoza LH, Youker KA, et al. Cardiac myocytes produce interleukin-6 in culture and in viable border zone of reperfused infarctions. *Circulation*. 1999;99:546-551. doi:[10.1161/01.cir.99.4.546](https://doi.org/10.1161/01.cir.99.4.546)
 87. Dawn B, Xuan YT, Guo Y, et al. IL-6 plays an obligatory role in late preconditioning via JAK-STAT signaling and upregulation of iNOS and COX-2. *Cardiovasc Res*. 2004;64:61-71. doi:[10.1016/j.cardiores.2004.05.011](https://doi.org/10.1016/j.cardiores.2004.05.011)
 88. Wang J, Wang M, Lu X, et al. IL-6 inhibitors effectively reverse post-infarction cardiac injury and ischemic myocardial remodeling via the TGF- β 1/Smad3 signaling pathway. *Exp Ther Med*. 2022;24(3):576. doi:[10.3892/etm.2022.11513](https://doi.org/10.3892/etm.2022.11513)
 89. Matsushita K, Iwanaga S, Oda T, et al. Interleukin-6/soluble interleukin-6 receptor complex reduces infarct size via inhibiting myocardial apoptosis. *Lab Invest*. 2005;85:1210-1223. doi:[10.1038/labinvest.3700322](https://doi.org/10.1038/labinvest.3700322)
 90. Fontes JA, Rose NR, Cihakova D. The varying faces of IL-6: from cardiac protection to cardiac failure. *Cytokine*. 2015;74:62-68. doi:[10.1016/j.cyto.2014.12.024](https://doi.org/10.1016/j.cyto.2014.12.024)
 91. Ridker PM, Everett BM, Pradhan A, et al. Low-dose methotrexate for the prevention of atherosclerotic events. *N Engl J Med*. 2019;380:752-762. doi:[10.1056/NEJMoa1809798](https://doi.org/10.1056/NEJMoa1809798)
 92. Fisman EZ, Benderly M, Esper RJ, et al. Interleukin-6 and the risk of future cardiovascular events in patients with angina pectoris and/or healed myocardial infarction. *Am J Cardiol*. 2006;98:14-18. doi:[10.1016/j.amjcard.2006.01.045](https://doi.org/10.1016/j.amjcard.2006.01.045)
 93. Ritschel VN, Seljeflot I, Arnesen H, et al. Circulating levels of IL-6 receptor and gp130 and long-term clinical outcomes in ST-elevation myocardial infarction. *J Am Heart Assoc*. 2016;5:e003014. doi:[10.1161/jaha.115.003014](https://doi.org/10.1161/jaha.115.003014)
 94. Groot HE, al Ali L, van der Horst ICC, et al. Plasma interleukin 6 levels are associated with cardiac function after ST-elevation myocardial infarction. *Clin Res Cardiol*. 2019;108:612-621. doi:[10.1007/s00392-018-1387-z](https://doi.org/10.1007/s00392-018-1387-z)
 95. Lindmark E, Diderholm E, Wallentin L, Siegbahn A. Relationship between interleukin 6 and mortality in patients with unstable coronary artery disease: effects of an early invasive or noninvasive strategy. *JAMA*. 2001;286:2107-2113. doi:[10.1001/jama.286.17.2107](https://doi.org/10.1001/jama.286.17.2107)
 96. Ridker PM, MacFadyen JG, Glynn RJ, Bradwin G, Hasan AA, Rifai N. Comparison of interleukin-6, C-reactive protein, and low-density lipoprotein cholesterol as biomarkers of residual risk in contemporary practice: secondary analyses from the cardiovascular inflammation reduction trial. *Eur Heart J*. 2020;41:2952-2961. doi:[10.1093/eurheartj/ehaa160](https://doi.org/10.1093/eurheartj/ehaa160)
 97. Zegeye MM, Andersson JSO, Wennberg P, Reipsilber D, Sirsjö A, Ljungberg LU. IL-6 as a mediator of the association between traditional risk factors and future myocardial infarction: a nested case-control study. *Arterioscler Thromb Vasc Biol*. 2021;41:1570-1579. doi:[10.1161/ATVBAHA.120.315793](https://doi.org/10.1161/ATVBAHA.120.315793)
 98. Hartman MH, Hartman MHT, Vreeswijk-Baudoin I, et al. Inhibition of Interleukin-6 receptor in a murine model of myocardial ischemia-reperfusion. *PLoS One*. 2016;11:e0167195. doi:[10.1371/journal.pone.0167195](https://doi.org/10.1371/journal.pone.0167195)
 99. Kobara M, Noda K, Kitamura M, et al. Antibody against interleukin-6 receptor attenuates left ventricular remodeling after myocardial infarction in mice. *Cardiovasc Res*. 2010;87:424-430. doi:[10.1093/cvr/cvq078](https://doi.org/10.1093/cvr/cvq078)
 100. George MJ, Jasmin NH, Cummings VT, et al. Selective Interleukin-6 trans-signaling blockade is more effective than

- Panantagonism in Reperfused myocardial infarction. *JACC Basic Transl Sci.* 2021;6:431-443. doi:[10.1016/j.jacbts.2021.01.013](https://doi.org/10.1016/j.jacbts.2021.01.013)
101. Muller J, Gorressen S, Grandoch M, et al. Interleukin-6-dependent phenotypic modulation of cardiac fibroblasts after acute myocardial infarction. *Basic Res Cardiol.* 2014;109:440. doi:[10.1007/s00395-014-0440-y](https://doi.org/10.1007/s00395-014-0440-y)
 102. Broch K, Anstensrud AK, Woxholt S, et al. Randomized trial of Interleukin-6 receptor inhibition in patients with acute ST-segment elevation myocardial infarction. *J Am Coll Cardiol.* 2021;77:1845-1855. doi:[10.1016/j.jacc.2021.02.049](https://doi.org/10.1016/j.jacc.2021.02.049)
 103. George MJ, Kleveland O, Garcia-Hernandez J, et al. Novel insights into the effects of interleukin 6 antagonism in non-ST-segment-elevation myocardial infarction employing the SOMAscan proteomics platform. *J Am Heart Assoc.* 2020;9:e015628. doi:[10.1161/JAHA.119.015628](https://doi.org/10.1161/JAHA.119.015628)
 104. Shah B, Pillinger M, Zhong H, et al. Effects of acute colchicine administration prior to percutaneous coronary intervention: COLCHICINE-PCI randomized trial. *Circ Cardiovasc Interv.* 2020;13:e008717. doi:[10.1161/circinterventions.119.008717](https://doi.org/10.1161/circinterventions.119.008717)
 105. Vilahur G, Radike M, Badimon L. Novel cardioprotective approaches in ischaemic heart disease. *Eur Heart J.* 2024;45:2109-2111. doi:[10.1093/eurheartj/ehae058](https://doi.org/10.1093/eurheartj/ehae058)
 106. Frangogiannis NG. Cardiac fibrosis. *Cardiovasc Res.* 2020;117:1450-1488. doi:[10.1093/cvr/cvaa324](https://doi.org/10.1093/cvr/cvaa324)
 107. Bacmeister L, Schwarzl M, Warnke S, et al. Inflammation and fibrosis in murine models of heart failure. *Basic Res Cardiol.* 2019;114:19. doi:[10.1007/s00395-019-0722-5](https://doi.org/10.1007/s00395-019-0722-5)
 108. Kumar S, Wang G, Zheng N, et al. HIMF (hypoxia-induced Mitogenic factor)-IL (interleukin)-6 signaling mediates Cardiomyocyte-fibroblast crosstalk to promote cardiac hypertrophy and fibrosis. *Hypertension.* 2019;73:1058-1070. doi:[10.1161/hypertensionaha.118.12267](https://doi.org/10.1161/hypertensionaha.118.12267)
 109. O'Reilly S, Ciecchomska M, Cant R, van Laar JM. Interleukin-6 (IL-6) trans signaling drives a STAT3-dependent pathway that leads to hyperactive transforming growth factor-beta (TGF-beta) signaling promoting SMAD3 activation and fibrosis via gremlin protein. *J Biol Chem.* 2014;289:9952-9960. doi:[10.1074/jbc.M113.545822](https://doi.org/10.1074/jbc.M113.545822)
 110. Wang JH, Zhao L, Pan X, et al. Hypoxia-stimulated cardiac fibroblast production of IL-6 promotes myocardial fibrosis via the TGF- β 1 signaling pathway. *Lab Invest.* 2016;96:839-852. doi:[10.1038/labinvest.2016.65](https://doi.org/10.1038/labinvest.2016.65)
 111. Jing R, Long TY, Pan W, Li F, Xie QY. IL-6 knockout ameliorates myocardial remodeling after myocardial infarction by regulating activation of M2 macrophages and fibroblast cells. *Eur Rev Med Pharmacol Sci.* 2019;23:6283-6291. doi:[10.26355/eurrev_201907_18450](https://doi.org/10.26355/eurrev_201907_18450)
 112. Li Q, Ye WX, Huang ZJ, Zhang Q, He YF. Effect of IL-6-mediated STAT3 signaling pathway on myocardial apoptosis in mice with dilated cardiomyopathy. *Eur Rev Med Pharmacol Sci.* 2019;23:3042-3050. doi:[10.26355/eurrev_201904_17586](https://doi.org/10.26355/eurrev_201904_17586)
 113. Li H, Bian Y. Fibroblast-derived interleukin-6 exacerbates adverse cardiac remodeling after myocardial infarction. *Korean J Physiol Pharmacol.* 2024;28:285-294. doi:[10.4196/kjpp.2024.28.3.285](https://doi.org/10.4196/kjpp.2024.28.3.285)
 114. Smart N, Mojet M, Latchman D, Marber M, Duchon M, Heads R. IL-6 induces PI 3-kinase and nitric oxide-dependent protection and preserves mitochondrial function in cardiomyocytes. *Cardiovasc Res.* 2006;69:164-177. doi:[10.1016/j.cardiores.2005.08.017](https://doi.org/10.1016/j.cardiores.2005.08.017)
 115. Tsutamoto T, Hisanaga T, Wada A, et al. Interleukin-6 spillover in the peripheral circulation increases with the severity of heart failure, and the high plasma level of interleukin-6 is an important prognostic predictor in patients with congestive heart failure. *J Am Coll Cardiol.* 1998;31:391-398. doi:[10.1016/s0735-1097\(97\)00494-4](https://doi.org/10.1016/s0735-1097(97)00494-4)
 116. Maeda K, Tsutamoto T, Wada A, et al. High levels of plasma brain natriuretic peptide and interleukin-6 after optimized treatment for heart failure are independent risk factors for morbidity and mortality in patients with congestive heart failure. *J Am Coll Cardiol.* 2000;36:1587-1593. doi:[10.1016/s0735-1097\(00\)00912-8](https://doi.org/10.1016/s0735-1097(00)00912-8)
 117. Vasan RS, Sullivan LM, Roubenoff R, et al. Inflammatory markers and risk of heart failure in elderly subjects without prior myocardial infarction: the Framingham heart study. *Circulation.* 2003;107:1486-1491. doi:[10.1161/01.cir.0000057810.48709.f6](https://doi.org/10.1161/01.cir.0000057810.48709.f6)
 118. Plenz G, Song ZF, Tjan TD, et al. Activation of the cardiac interleukin-6 system in advanced heart failure. *Eur J Heart Fail.* 2001;3:415-421. doi:[10.1016/s1388-9842\(01\)00137-4](https://doi.org/10.1016/s1388-9842(01)00137-4)
 119. Kalogeropoulos A, Georgiopoulou V, Psaty BM, et al. Inflammatory markers and incident heart failure risk in older adults: the health ABC (health, aging, and body composition) study. *J Am Coll Cardiol.* 2010;55:2129-2137. doi:[10.1016/j.jacc.2009.12.045](https://doi.org/10.1016/j.jacc.2009.12.045)
 120. Markousis-Mavrogenis G, Tromp J, Ouwerkerk W, et al. The clinical significance of interleukin-6 in heart failure: results from the BIOSTAT-CHF study. *Eur J Heart Fail.* 2019;21:965-973. doi:[10.1002/ehf.1482](https://doi.org/10.1002/ehf.1482)
 121. Anand IS, Gupta P. Anemia and iron deficiency in heart failure. *Circulation.* 2018;138:80-98. doi:[10.1161/CIRCULATIONAHA.118.030099](https://doi.org/10.1161/CIRCULATIONAHA.118.030099)
 122. Everett BM, Cornel JH, Lainscak M, et al. Anti-inflammatory therapy with Canakinumab for the prevention of hospitalization for heart failure. *Circulation.* 2019;139:1289-1299. doi:[10.1161/circulationaha.118.038010](https://doi.org/10.1161/circulationaha.118.038010)
 123. Berger M, März W, Niessner A, et al. IL-6 and hsCRP predict cardiovascular mortality in patients with heart failure with preserved ejection fraction. *ESC Heart Fail.* 2024;11:3607-3615. doi:[10.1002/ehf2.14959](https://doi.org/10.1002/ehf2.14959)
 124. Ohtsuka T, Hamada M, Hiasa G, et al. Effect of beta-blockers on circulating levels of inflammatory and anti-inflammatory cytokines in patients with dilated cardiomyopathy. *J Am Coll Cardiol.* 2001;37:412-417. doi:[10.1016/S0735-1097\(00\)01121-9](https://doi.org/10.1016/S0735-1097(00)01121-9)
 125. Awad K, Zaki MM, Mohammed M, et al. Effect of the renin-angiotensin system inhibitors on inflammatory markers: a systematic review and meta-analysis of randomized controlled trials. *Mayo Clin Proc.* 2022;97:1808-1823. doi:[10.1016/j.mayocp.2022.06.036](https://doi.org/10.1016/j.mayocp.2022.06.036)
 126. Muñoz-Durango N, Vecchiola A, Gonzalez-Gomez LM, et al. Modulation of immunity and inflammation by the mineralocorticoid receptor and aldosterone. *Biomed Res Int.* 2015;2015:652738. doi:[10.1155/2015/652738](https://doi.org/10.1155/2015/652738)
 127. Preda A, Montecucco F, Carbone F, et al. SGLT2 inhibitors: from glucose-lowering to cardiovascular benefits. *Cardiovasc Res.* 2024;120:443-460. doi:[10.1093/cvr/cvae047](https://doi.org/10.1093/cvr/cvae047)
 128. Patel NS, Patel NSA, Chatterjee PK, et al. Endogenous interleukin-6 enhances the renal injury, dysfunction, and inflammation caused by ischemia/reperfusion. *J Pharmacol Exp Ther.* 2005;312:1170-1178. doi:[10.1124/jpet.104.078659](https://doi.org/10.1124/jpet.104.078659)
 129. de Haij S, Bakker AC, van der Geest RN, et al. NF-kappaB mediated IL-6 production by renal epithelial cells is regulated by c-jun NH2-terminal kinase. *J Am Soc Nephrol.* 2005;16:1603-1611. doi:[10.1681/ASN.2004090781](https://doi.org/10.1681/ASN.2004090781)

130. Lei CT, Su H, Ye C, et al. The classic signalling and trans-signalling of interleukin-6 are both injurious in podocyte under high glucose exposure. *J Cell Mol Med*. 2018;22:251-260. doi:[10.1111/jcmm.13314](https://doi.org/10.1111/jcmm.13314)
131. Kayama F, Yoshida T, Kodama Y, Matsui T, Matheson JM, Luster MI. Pro-inflammatory cytokines and interleukin 6 in the renal response to bacterial endotoxin. *Cytokine*. 1997;9:688-695. doi:[10.1006/cyto.1997.0214](https://doi.org/10.1006/cyto.1997.0214)
132. Grassl C, Luckow B, Schlondorff D, Dendorfer U. Transcriptional regulation of the interleukin-6 gene in mesangial cells. *J Am Soc Nephrol*. 1999;10:1466-1477. doi:[10.1681/ASN.V1071466](https://doi.org/10.1681/ASN.V1071466)
133. Ansari Z, Chaurasia A, Sharma N, Bachheti RK, Gupta PC. Exploring inflammatory and fibrotic mechanisms driving diabetic nephropathy progression. *Cytokine Growth Factor Rev*. 2025;84:120-134. doi:[10.1016/j.cytogfr.2025.05.007](https://doi.org/10.1016/j.cytogfr.2025.05.007)
134. Gai L, Zhu Y, Zhang C, Meng X. Targeting canonical and non-canonical STAT signaling pathways in renal diseases. *Cells*. 2021;10:1610. doi:[10.3390/cells10071610](https://doi.org/10.3390/cells10071610)
135. Bienaime F, Muorah M, Yammine L, et al. Stat3 controls Tubulointerstitial communication during CKD. *J Am Soc Nephrol*. 2016;27:3690-3705. doi:[10.1681/ASN.2015091014](https://doi.org/10.1681/ASN.2015091014)
136. Guo X, Zhu Y, Sun Y, Li X. IL-6 accelerates renal fibrosis after acute kidney injury via DNMT1-dependent FOXO3a methylation and activation of Wnt/beta-catenin pathway. *Int Immunopharmacol*. 2022;109:108746. doi:[10.1016/j.intimp.2022.108746](https://doi.org/10.1016/j.intimp.2022.108746)
137. Kielar ML, John R, Bennett M, et al. Maladaptive role of IL-6 in ischemic acute renal failure. *J Am Soc Nephrol*. 2005;16:3315-3325. doi:[10.1681/ASN.2003090757](https://doi.org/10.1681/ASN.2003090757)
138. Zhang L, Xu F, Hou L. IL-6 and diabetic kidney disease. *Front Immunol*. 2024;15:1465625. doi:[10.3389/fimmu.2024.1465625](https://doi.org/10.3389/fimmu.2024.1465625)
139. Wu R, Liu X, Yin J, et al. IL-6 receptor blockade ameliorates diabetic nephropathy via inhibiting inflammasome in mice. *Metabolism*. 2018;83:18-24. doi:[10.1016/j.metabol.2018.01.002](https://doi.org/10.1016/j.metabol.2018.01.002)
140. Hashmat S, Rudemiller N, Lund H, Abais-Battad JM, van Why S, Mattson DL. Interleukin-6 inhibition attenuates hypertension and associated renal damage in Dahl salt-sensitive rats. *Am J Physiol Renal Physiol*. 2016;311:F555-F561. doi:[10.1152/ajprenal.00594.2015](https://doi.org/10.1152/ajprenal.00594.2015)
141. Zhang W, Wang W, Yu H, et al. Interleukin 6 underlies angiotensin II-induced hypertension and chronic renal damage. *Hypertension*. 2012;59:136-144. doi:[10.1161/HYPERTENSIONAHA.111.173328](https://doi.org/10.1161/HYPERTENSIONAHA.111.173328)
142. Sanchez-Alamo B, Shabaka A, Cachofeiro V, Cases-Corona C, Fernandez-Juarez G, PRONEDI study investigators. Serum interleukin-6 levels predict kidney disease progression in diabetic nephropathy. *Clin Nephrol*. 2022;97:1-9. doi:[10.5414/CN110223](https://doi.org/10.5414/CN110223)
143. Barreto DV, Barreto FC, Liabeuf S, et al. Plasma interleukin-6 is independently associated with mortality in both hemodialysis and pre-dialysis patients with chronic kidney disease. *Kidney Int*. 2010;77:550-556. doi:[10.1038/ki.2009.503](https://doi.org/10.1038/ki.2009.503)
144. Batra G, Ghukasyan L, Lindbäck J, et al. Interleukin 6 and cardiovascular outcomes in patients with chronic kidney disease and chronic coronary syndrome. *JAMA Cardiol*. 2021;6:1440-1445. doi:[10.1001/jamacardio.2021.3079](https://doi.org/10.1001/jamacardio.2021.3079)
145. Gupta J, Mitra N, Kanetsky PA, et al. Association between albuminuria, kidney function, and inflammatory biomarker profile in CKD in CRIC. *Clin J Am Soc Nephrol*. 2012;7:1938-1946. doi:[10.2215/cjn.03500412](https://doi.org/10.2215/cjn.03500412)
146. Amdur RL, Feldman HI, Gupta J, et al. Inflammation and progression of CKD: the CRIC study. *Clin J Am Soc Nephrol*. 2016;11:1546-1556. doi:[10.2215/cjn.13121215](https://doi.org/10.2215/cjn.13121215)
147. Amdur RL, Feldman HI, Dominic EA, et al. Use of measures of inflammation and kidney function for prediction of atherosclerotic vascular disease events and death in patients with CKD: findings from the CRIC study. *Am J Kidney Dis*. 2019;73:344-353. doi:[10.1053/j.ajkd.2018.09.012](https://doi.org/10.1053/j.ajkd.2018.09.012)
148. Dieter BP, McPherson SM, Afkarian M, et al. Serum amyloid A and risk of death and end-stage renal disease in diabetic kidney disease. *J Diabetes Complicat*. 2016;30:1467-1472. doi:[10.1016/j.jdiacomp.2016.07.018](https://doi.org/10.1016/j.jdiacomp.2016.07.018)
149. Garrido P, Ribeiro S, Fernandes J, et al. Iron-hepcidin dysmetabolism, anemia and renal hypoxia, inflammation and fibrosis in the remnant kidney rat model. *PLoS One*. 2015;10:e0124048. doi:[10.1371/journal.pone.0124048](https://doi.org/10.1371/journal.pone.0124048)
150. Weiss G, Goodnough LT. Anemia of chronic disease. *N Engl J Med*. 2005;352:1011-1023. doi:[10.1056/NEJMra041809](https://doi.org/10.1056/NEJMra041809)
151. Gupta J, Dominic EA, Fink JC, et al. Association between inflammation and cardiac geometry in chronic kidney disease: findings from the CRIC study. *PLoS One*. 2015;10:e0124772. doi:[10.1371/journal.pone.0124772](https://doi.org/10.1371/journal.pone.0124772)
152. Roy N, Rosas SE. IL-6 is associated with progression of coronary artery calcification and mortality in incident dialysis patients. *Am J Nephrol*. 2021;52:745-752. doi:[10.1159/000518652](https://doi.org/10.1159/000518652)
153. Kaminska J et al. IL 6 but not TNF is linked to coronary artery calcification in patients with chronic kidney disease. *Cytokine*. 2019;120:9-14. doi:[10.1016/j.cyto.2019.04.002](https://doi.org/10.1016/j.cyto.2019.04.002)
154. Krane V, Winkler K, Drechsler C, et al. Association of LDL cholesterol and inflammation with cardiovascular events and mortality in hemodialysis patients with type 2 diabetes mellitus. *Am J Kidney Dis*. 2009;54:902-911. doi:[10.1053/j.ajkd.2009.06.029](https://doi.org/10.1053/j.ajkd.2009.06.029)
155. Ridker PM, Tuttle KR, Perkovic V, Libby P, MacFadyen JG. Inflammation drives residual risk in chronic kidney disease: a CANTOS substudy. *Eur Heart J*. 2022;43:4832-4844. doi:[10.1093/eurheartj/ehac444](https://doi.org/10.1093/eurheartj/ehac444)
156. Pergola PE, Devalaraja M, Fishbane S, et al. Ziltivekimab for treatment of anemia of inflammation in patients on hemodialysis: results from a phase 1/2 multicenter, randomized, double-blind, placebo-controlled trial. *J Am Soc Nephrol*. 2021;32:211-222. doi:[10.1681/ASN.2020050595](https://doi.org/10.1681/ASN.2020050595)
157. Kreiner FF, Kraaijenhof JM, von Herrath M, Hovingh GKK, von Scholten BJ. Interleukin 6 in diabetes, chronic kidney disease, and cardiovascular disease: mechanisms and therapeutic perspectives. *Expert Rev Clin Immunol*. 2022;18:377-389. doi:[10.1080/1744666X.2022.2045952](https://doi.org/10.1080/1744666X.2022.2045952)
158. Ridker PM, Devalaraja M, Baeres FMM, et al. IL-6 inhibition with ziltivekimab in patients at high atherosclerotic risk (RESCUE): a double-blind, randomised, placebo-controlled, phase 2 trial. *Lancet*. 2021;397:2060-2069. doi:[10.1016/S0140-6736\(21\)00520-1](https://doi.org/10.1016/S0140-6736(21)00520-1)
159. Paulus WJ, Zile MR. From systemic inflammation to myocardial fibrosis: the heart failure with preserved ejection fraction paradigm revisited. *Circ Res*. 2021;128:1451-1467. doi:[10.1161/CIRCRESAHA.121.318159](https://doi.org/10.1161/CIRCRESAHA.121.318159)

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