

Gut microbiota-derived imidazole propionate predicts cardiometabolic risk in patients with coronary artery disease

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Abstract

1 **Background and Aims:** The gut microbiota is a modulator of cardiometabolic disease.
2
3 Circulating imidazole propionate (ImP) is a microbiota-derived proatherogenic amino acid
4 metabolite modulating the inflammatory response of myeloid cells, endothelial function and
5 glucose metabolism. This study examined the prognostic value of ImP in patients with coronary
6 artery disease (CAD).

7 **Methods:** Circulating ImP levels were measured in independent prospective cohorts of patients
8 with acute coronary syndrome (ACS, Switzerland $n=4937$, Germany $n=1497$) and chronic
9 coronary syndrome (CCS, Germany $n=701$). Major adverse cardiovascular events (MACE),
10 defined as the first occurrence of a composite of death, nonfatal myocardial infarction, or
11 nonfatal stroke after admission, were the primary endpoint. Cox models, accounting for
12 established risk factors including the gut-derived cardiovascular risk factor trimethylamine N-
13 oxide (TMAO), were used to evaluate the predictive value of ImP.

14 **Results:** Circulating ImP was associated with more advanced CAD and with cardiometabolic
15 characteristics including diabetes and elevated high-sensitivity C-reactive protein. High ImP was
16 an independent predictor of MACE (Swiss ACS cohort: hazard ratio [HR] per log₂ increase 1.22,
17 95% confidence interval [CI] 1.10–1.35, $P<0.001$; German ACS cohort: HR 2.34, 95% CI 1.46–
18 3.76, $P<0.001$; German CCS cohort: HR 1.32, 95% CI 1.13–1.53, $P<0.001$) and of mortality
19 (Swiss ACS cohort: HR 1.34, 95% CI 1.17–1.54, $P<0.001$; German ACS cohort: HR 2.38, 95%
20 CI 1.48–3.82, $P<0.001$; German CCS cohort: HR 1.50, 95% CI 1.14–1.98, $P=0.004$) after
21 adjustment for established risk factors. ImP provided predictive value beyond TMAO (Swiss
22 ACS cohort: HR 1.30, 95% CI 1.05–1.61, $P=0.014$; German CCS cohort: HR 1.31, 95% CI 1.11–
23 1.53, $P=0.001$).

1 **Conclusions:** Gut microbiota-derived ImP predicted MACE in patients with CAD independently
2 of traditional risk factors and holds promise as a therapeutic target. ImP may refine risk
3 stratification for personalised secondary prevention strategies.

4

5 **Keywords:** Imidazole Propionate – Gut Microbiota – Cardiometabolic Risk – Coronary Artery
6 Disease

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Introduction

Despite substantial advances in the control of traditional drivers of atherosclerosis, coronary artery disease (CAD) remains the leading cause of death in adults worldwide.^{1,2} With the evolution of risk factor profiles, attention has shifted increasingly to non-traditional cardiovascular risk factors.^{3,4}

Imidazole propionate (ImP) is a gut microbiota-derived proatherogenic histidine metabolite⁵⁻⁷ that impairs endothelial function⁸ and glucose metabolism.^{5,9} At a molecular level, ImP induces inflammatory activation of myeloid and endothelial cells, hampers endothelial repair, and promotes atherogenesis. Circulating levels of ImP are primarily determined by the composition of the gut microbiota.^{5,6} Indeed, ImP is microbially produced by gut bacteria that are abundant in patients with type 2 diabetes^{9,10} and in individuals with subclinical coronary atherosclerosis.¹¹

ImP may thus constitute a novel therapeutic target for modulating cardiometabolic risk¹² and holds potential to refine cardiovascular risk stratification. Clinical studies suggest that circulating ImP is associated with systemic inflammation¹⁰, diabetes¹⁰, heart failure,¹³ and subclinical atherosclerosis in asymptomatic volunteers.⁷ A recent study⁸ reported elevated ImP levels in subjects with CAD yet extensive clinical investigations are lacking.

Here, we aim to assess the relationship between ImP and clinical outcomes in patients with CAD including acute coronary syndromes (ACS) and chronic coronary syndromes (CCS).

Methods

Study design and participants

We aimed to encompass the full spectrum of CAD including patients presenting with acute and stable clinical conditions. We used prospective ACS and CCS cohorts from Switzerland and Germany (*Supplementary data online, Figures S1 and S2*).

In the investigator-initiated prospective multicentre Swiss ACS cohort (SPUM-ACS Biomarker Study, ClinicalTrials.gov Identifier: NCT01000701), a total of 4787 patients with ACS were recruited from December 8, 2009 until December 31, 2017.¹⁴⁻²³ Diagnoses of ACS were independently confirmed by expert cardiologists at local study facilities.^{14,17} Participants received guideline-directed therapy regimens, as reported previously.¹⁷ To investigate the coronary microcirculation, we analysed patients with ACS, enrolled in the Controlled Level EVERolimus in Acute Coronary Syndromes (CLEVER-ACS; ClinicalTrials.gov Identifier: NCT01529554) study in Switzerland from November 26, 2014 to October 29, 2021 undergoing cardiac magnetic resonance (CMR) imaging at 30-day follow-up (Swiss CMR cohort; $n = 60$; 8 of whom were co-enrolled in SPUM-ACS).

The prospective German ACS cohort (Heidelberg II-ACS study) includes 1428 patients with ACS presenting to Heidelberg University Hospital, Heidelberg, Germany between September 14, 2010, and July 21, 2014. Patients in this cohort were analysed using a case-cohort design, permitting unbiased assessment of exposure–outcome relationships, while reducing measurement costs. The design of case-cohort analyses has been reported previously^{24,25} and a detailed description is provided in the *Suppl. material*. In brief, we selected a random sample of patients (i.e., the subcohort) from the pool of eligible participants. We then also selected all individuals

1 with incident events who were not part of the subcohort. The final selection of patients ($n = 250$)
2 consists of the subcohort and participants with incident events outside the subcohort.

3 The prospective German CCS cohort (LipidCardio Study, German Clinical Trial Register
4 Identifier: DRKS00020915) included 701 consecutive patients with CSS undergoing elective
5 diagnostic cardiac catheterisation at the Department of Cardiology, Campus Benjamin Franklin,
6 Charité-Universitätsmedizin, Berlin, Germany between October 27, 2016 and March 8, 2018.
7 Chronic coronary syndrome diagnoses were confirmed by angiographic evidence of obstructive
8 CAD ($\geq 50\%$ stenosis).

9 The cohort profiles and detailed inclusion and exclusion criteria of the involved cohorts have
10 been reported previously²⁶ and are summarised in the *Suppl. material*.

11 **Imidazole propionate measurements**

12 Blood samples were obtained at the time of presentation without a dedicated fasting interval and
13 stored at -80°C . ImP and its precursor molecules, histidine and trans-urocanate, were measured
14 centrally using ultra-high performance liquid chromatography coupled to tandem mass
15 spectrometry. Samples were prepared and analysed at the University of Gothenburg, Gothenburg,
16 Sweden, using an established method with minor modifications.¹⁰ In brief, 25 μL of sample
17 volume were extracted with 6 volumes of acetonitrile containing internal standards (100 nmol/L
18 ImP- $^{13}\text{C}_3$ and urocanate- $^{13}\text{C}_3$, AstraZeneca, Cambridge, United Kingdom and 1 $\mu\text{mol/L}$ L-
19 histidine D₅, $^{15}\text{N}_3$, Cambridge Isotope Laboratories, Inc. Andover, MA, USA) before drying under
20 a nitrogen stream. The samples were then reconstituted in 5% HCl (37%) in 1-butanol for n-butyl
21 ester derivatization at 70°C for 40 min dried and finally reconstituted in 150 μL water:acetonitrile
22 (9:1). ImP, urocanate, and histidine levels were quantified using multiple reaction monitoring of
23 the transitions 197.2/81.2, 195.2/93.0, 213.2/110.1 and internal standards of the transitions

1 200.2/82.0, 198.2/95.0 and 220.3/118.1. Investigators performing the measurements were blinded
2 to sample allocation and patient data other than the deidentified barcode label.

3 **Cardiac magnetic resonance imaging**

4 In Switzerland, patients with ACS included in the CLEVER-ACS study underwent CMR
5 imaging at 30-day follow-up according to a pre-specified study protocol. In brief, within 24 h
6 after onset of chest pain, patients with proximal occlusion of a coronary artery underwent primary
7 percutaneous coronary intervention (PCI) with placement of a drug-eluting stent in the culprit
8 lesion.²⁷ The CMR imaging protocol at 30-day follow-up included T1- and T2-weighted
9 sequences for tissue characterisation and calculation of ventricular volumes, function, and mass.²⁷
10 Fifteen minutes after administration of a gadolinium-chelate (0.20 mmol/kg of body weight), an
11 inversion recovery fast-gradient echo imaging sequence was used to visualise scar tissue.²⁷ The
12 extent of coronary microvascular obstruction was quantified by contouring the dark core areas
13 manually in the core of the necrotic zone.²⁷ Imaging data analysis was performed in blinded
14 fashion by expert physicians in the CMR Core Laboratory at the University Hospital Zurich,
15 Zurich, Switzerland, employing dedicated heart imaging software (GTVolume, Gyrotools Ltd).

16 **Study endpoints, follow-up, and outcome adjudication**

17 Major adverse cardiovascular events (MACE), defined as the first occurrence of a composite of
18 death, nonfatal myocardial infarction, or nonfatal stroke after admission, were the primary
19 endpoint. Mortality was analysed as an additional endpoint. An event horizon of 1 year (365
20 days) for the ACS cohorts and of 3 years (1095 days) for the German CCS cohort was used.

21 In Switzerland, patients with ACS were followed up at hospital discharge, at 30 days (via
22 phone call) and at 1 year (clinical visit). Baseline and event data were collected by study
23 personnel at the respective study centres via a centralised web-based data entry system

1 (CARDIOBASE, Clinical Trial Unit and Department of Cardiology, University Hospital Bern,
2 Bern, Switzerland, and Webspirit Systems GmbH, Ulm, Germany).¹⁴ Events were assessed based
3 on pre-specified adjudication forms by an independent clinical endpoint committee composed of
4 three certified experienced cardiologists, who were blinded to patients' baseline characteristics .¹⁴
5 At each study site, a committee of expert cardiologists supervised data collection. In the German
6 ACS cohort, patients were followed up via telephone call and/or a questionnaire forwarded by
7 post or e-mail, as previously described.²⁸ All deaths (retrieved via local resident registry records)
8 and nonfatal adverse cardiovascular events were recorded.²⁸ In the German CCS cohort, patients
9 were followed up via phone call at 3 years after study enrolment²⁶ to assess nonfatal adverse
10 cardiovascular events.²⁶ Mortality was retrieved from the obligatory registry office.²⁶

11 **Statistical analyses**

12 Continuous variables are presented as median and interquartile range (IQR).¹⁴ Categorical data
13 are reported as counts and valid percentages.¹⁴ Continuous variables were compared with the
14 Mann-Whitney U test for comparisons across two groups and the Kruskal-Wallis test for
15 comparisons across three or more groups. Trends across independent groups were tested using
16 the Jonckheere-Terpstra test. We used partial Spearman rank-correlation to examine the
17 association between pairs of continuous variables while adjusting for additional variables.²⁹

18 The association of ImP with cardiometabolic characteristics was examined in the pooled CAD
19 patient population (data from the Swiss ACS cohort and the German CCS cohort) to increase the
20 number of patients in the analyses, as well as stratified by ACS and CCS status. Given the
21 different clinical implications, risk modelling was stratified according to presentation with ACS
22 and presentation with CCS. The association of circulating ImP with clinical outcomes was
23 studied in complete case Cox proportional hazards regression models,^{17,28,30} using log 2-

1 transformed ImP levels^{14,31,32}, i.e., relative effect estimates correspond to a doubling in ImP
2 levels. In addition, patients were divided according to ImP quartile (Q) and biomarker levels
3 analysed on a categorical scale (***Supplementary data online, Table S20***).^{10,31} Quartile cut-offs
4 were derived from the largest cohort and applied to all cohorts (Q1: 9.4 nM; Q2: 9.4–14.3 nM;
5 Q3: 14.3–24.4 nM; Q4: > 24.4 nM; ***Supplementary data online***). In a sensitivity analysis, cohort-
6 specific ImP cut-offs were explored (***Supplementary data online, Table S19***).

7 Cumulative incidence curves were plotted and differences between curves were examined using
8 log-rank statistics. We constructed smoothed hazard ratio (HR) plots with three knots across the
9 ImP distribution. Multivariable analyses were conducted using a stepwise approach adjusting for
10 predefined covariables, based on sample size and clinical considerations (***Supplementary data***
11 ***online***). ImP levels were analysed in (i) crude, (ii) sex-, age- and diabetes-adjusted, and (iii) fully
12 adjusted models. In the fully adjusted model, covariables included age, sex, diabetes, high-
13 sensitivity troponin T (hs-TnT), high-sensitivity C-reactive protein (hs-CRP), revascularisation,
14 ACS type (if applicable), body mass index and estimated glomerular filtration rate. Analyses on
15 mortality in patients with ACS were additionally adjusted for the GRACE 2.0 score for 1-year
16 mortality.³³ We further evaluated the predictive value of circulating ImP after additionally
17 adjusting for its precursor molecules histidine and urocanate (***Supplementary data online, Tables***
18 ***S15 and S16***). Additionally, we accounted for ethnicity (only available in German CCS cohort),
19 lipoprotein(a) [Lp(a)] levels, history of chronic liver disease, and leukocyte count in the Swiss
20 ACS and in the German CCS cohorts (***Supplementary data online, Tables S15 and S16***). The
21 proportional hazards assumption was checked in the fully adjusted model using Schoenfeld
22 residuals (***Supplementary data online***). In a subgroup of patients from the Swiss ACS cohort and
23 the German CCS cohort with available TMAO levels,^{13,34} the incremental predictive value of ImP
24 beyond TMAO was assessed by additionally adjusting for TMAO. Moreover, we analysed joint

1 groups of high (\geq Q4 cut-off) vs. low ($<$ Q4 cut-off) levels of ImP and TMAO yielding four
2 strata: Low ImP/Low TMAO, Low ImP/High TMAO, High ImP/Low TMAO, and High
3 ImP/High TMAO. Patients with CMR imaging at 30-day follow-up were stratified into two
4 groups according to baseline ImP levels (i.e., Q4 vs. Q1–3) and coronary microvascular
5 obstruction was compared between groups using analysis of covariance adjusted for age, sex,
6 diabetes, hypertension, low-density lipoprotein cholesterol, glucose, and preprocedural coronary
7 Thrombolysis in Myocardial Infarction (TIMI) flow grade. We conducted several subgroup
8 analyses to assess the consistency of the results with respect to different population
9 characteristics including sex, age, diabetes, proton pump inhibitor use, and dietary habits (only
10 available in German CCS cohort). In addition, rehospitalisation rates were examined in an
11 exploratory analysis (*Supplementary data online, Figure S14*). P-values and confidence
12 intervals (CI) are two-sided. The study was conducted in accordance with the principles of the
13 STROBE statement.³⁵ Statistical analyses were performed with R software version 4.3 or later
14 and Stata version 16.1 or later. A detailed description of the statistical analyses is provided in the
15 *Supplementary data online*.

17 Results

18 Circulating imidazole propionate is linked to the extent of coronary artery disease

19 A total of 7066 patients were enrolled in Switzerland (ACS $n = 4937$) and Germany (ACS $n =$
20 1428, CCS $n = 701$) (*Supplementary data online, Figures S1*). Baseline and treatment
21 characteristics are summarised in *Tables 1 and 2* and in the *Supplementary data online (Tables*
22 *S1-S12)*. Circulating ImP levels were significantly elevated in patients with CAD (*Figure 1A*).
23 Moreover, high ImP was associated with more advanced atherosclerotic disease, as determined

1 by a higher number of affected coronary arteries, lower left ventricular ejection fraction, and
2 more severe symptoms of heart failure (**Figure 1A**). These findings were consistent across
3 clinical presentation with ACS and CCS (**Supplementary data online, Figures S7 and S9**) but
4 absent or less pronounced for the precursor molecules histidine and urocanate (**Supplementary**
5 **data online, Figures S6, S8, S10, Table S13**). ImP levels were comparable across different times
6 of the day (**Supplementary data online, Figures S3 and S4**).

7 **High imidazole propionate associates with cardiometabolic characteristics**

8 Given the established effects of ImP on glucose metabolism, we studied its relationship to
9 cardiometabolic characteristics of patients with CAD. High ImP was associated with high body
10 mass index, systemic inflammation, impaired glucose metabolism, and hypertension (**Figure 1B**;
11 **Supplementary data online, Figure S11**). These results were similarly observed in patients with
12 ACS and in patients with CCS (**Supplementary data online, Figures S7 and S9**) but were absent
13 or less pronounced for the precursor molecules histidine and urocanate (**Supplementary data**
14 **online, Figures S8 and S10**).

15 **High imidazole propionate is associated with coronary microvascular obstruction**

16 Considering the intricate involvement of ImP in central processes of myocardial microcirculation
17 in preclinical experiments including endothelial dysfunction, regulation of endothelium-
18 dependent vascular tone, leucocyte function and adhesion, and osmotic regulation, we
19 performed an exploratory analysis to examine a potential association of ImP with CMR-assessed
20 microvascular obstruction. High ImP levels were associated with increased coronary
21 microvascular obstruction at 30 days (**Figure 2, Supplementary data online, Table S14**). These
22 results remained consistent on multivariable adjustment but were not observed for the precursor
23 molecules histidine and urocanate (**Supplementary data online, Figure S12**).

1 **High imidazole propionate predicts cardiovascular outcomes beyond established risk** 2 **factors and GRACE 2.0**

3 Across all cohorts, there were a total of 595 MACE and 270 deaths. Cumulative incidence curves
4 showed a substantial increase in MACE and mortality in patients with high ImP with HR plots
5 indicating a dose-response relationship (**Figure 3**). After multivariable adjustment, high ImP
6 levels were associated with increased risk of MACE (Swiss ACS cohort: fully adjusted HR per
7 log₂ increase, 1.22, 95% CI 1.10–1.35, $P < 0.001$, fully adjusted HR Q4 vs. Q1–3, 1.33, 95%
8 CI 1.03–1.72, $P = 0.032$; German ACS cohort: fully adjusted HR per log₂ increase, 2.34, 95% CI
9 1.46–3.76 $P < 0.001$, fully adjusted HR Q4 vs. Q1–3, 7.97, 95% CI 2.70–23.52, $P < 0.001$;
10 German CCS cohort: fully adjusted HR per log₂ increase, 1.32, 95% CI 1.13–1.53, $P < 0.001$,
11 fully adjusted HR Q4 vs. Q1–3, 1.83 95% CI 1.22–2.74, $P = 0.004$; **Supplementary data online,**
12 **Table S15**). Likewise, high plasma ImP emerged as a strong independent predictor of all-cause
13 mortality (Swiss ACS cohort: fully adjusted + GRACE 2.0 HR per log₂ increase, 1.24, 95% CI
14 1.06–1.46, $P = 0.007$, fully adjusted HR + GRACE 2.0 Q4 vs. Q1–3, 1.47, 95% CI 0.96–2.26, P
15 = 0.076; German ACS cohort: fully adjusted + GRACE 2.0 HR per log₂ increase, 2.32, 95% CI
16 1.46–3.41, $P < 0.001$, fully adjusted HR + GRACE 2.0 Q4 vs. Q1–3, 4.84, 95% CI 1.42–16.47, P
17 = 0.012; German CCS cohort: fully adjusted HR per log₂ increase, 1.50, 95% CI 1.14–1.98, $P =$
18 0.004, fully adjusted HR Q4 vs. Q1–3, 2.01, 95% CI 0.96–4.19, $P = 0.063$; **Supplementary data**
19 **online, Table S16**). These findings were consistent across patient subgroups (**Supplementary**
20 **data online, Tables S17 and S18, Figure S5**). Similar results were obtained when additionally
21 adjusting for Lp(a), liver disease, leukocyte count and for the precursor molecules of ImP
22 (**Supplementary data online, Tables S15 and S16**).

23 **Imidazole propionate adds to microbiota-related risk beyond TMAO**

1 ImP provided incremental predictive value for MACE beyond TMAO (Swiss ACS cohort: fully
2 adjusted + TMAO HR per log₂ increase: 1.30, 95% CI 1.05–1.61, $P = 0.014$; German CCS
3 cohort: fully adjusted + TMAO HR per log₂ increase: 1.31, 95% CI 1.12 – 1.53, $P = 0.001$;
4 *Supplementary data online, Table S21*). When stratifying patients into joint groups according to
5 both ImP and TMAO level, individuals with high levels of both markers exhibited excessive risk
6 of MACE (*Figure 4A*). After multivariable adjustment, joint elevation of ImP and TMAO
7 conferred a more than 2-fold increased risk of MACE in patients with ACS and CCS (Swiss ACS
8 cohort: fully adjusted HR: 2.28, 95% CI 1.14–4.56, $P = 0.020$; German CCS cohort: fully
9 adjusted HR: 4.30, 95% CI 2.53–7.32, $P < 0.001$; *Figure 4B*). Circulating ImP ranked among the
10 strongest predictors of MACE (*Figure 4C*). These findings were similarly observed for mortality
11 (*Supplementary data online, Figure S13 and Table S22*).

13 Discussion

14 Here, we show for the first time, that the gut microbiota-derived amino-acid metabolite ImP
15 is linked to cardiometabolic characteristics and independently predicts adverse outcomes in
16 patients with ACS and CCS beyond established risk markers (Structured Graphical Abstract).

17 Despite advancements in the management of patients with CAD over the past decades, there
18 is considerable residual cardiovascular risk attributed to non-traditional risk factors such as
19 inflammation, disturbed sleep, air pollution, physical inactivity, environmental stress, and
20 microbial dysbiosis. ImP is produced from urocanate, a metabolite of the essential amino-acid
21 histidine, by the microbial enzyme urocanate reductase.⁶ Previous studies have shown that
22 circulating ImP levels are primarily determined by the composition and metabolism of gut
23 microbiota, rather than histidine intake.¹⁰

1 At the cellular level, ImP induces imidazoline-1 receptor (I1R) signalling⁷ and inhibits
2 insulin-receptor signalling,^{5,9} in turn leading to activation of inflammation-associated pathways in
3 macrophages, fibroblasts and endothelial cells,⁸ impaired endothelial cell regeneration after
4 vascular injury, reduced endothelium-dependent arterial relaxation, impaired glucose
5 metabolism^{5,6} and increased atherogenesis.⁸ Notably, ImP promotes the expression of leucocyte
6 adhesion molecules for leucocyte-endothelial cell interactions leading to an inflammatory milieu
7 at the endothelial interface and increased immune cell infiltration, a hallmark of atherosclerosis.³
8 ⁴ The atherogenic effects of ImP involve different molecular pathways compared to TMAO.³⁶⁻³⁸

9 In line with previous mechanistic studies, our results demonstrate that ImP is associated with
10 signs of cardiometabolic dysregulation and more extensive coronary atherosclerosis in patients
11 with CAD. An exploratory analysis of a small cohort of patients with ACS undergoing primary
12 PCI followed up with CMR, indicated that patients with high ImP also had more extensive
13 coronary microvascular obstruction. These findings align well with preclinical studies showing
14 that ImP can influence key processes related to the microcirculation including endothelial cell
15 regeneration, regulation of the endothelium-dependent vascular tone,⁸ leucocyte adhesion, and
16 osmotic regulation. This suggests the possibility that ImP may contribute to the individual
17 susceptibility to microvascular dysfunction in response to ischaemia-reperfusion injury,³⁹ as
18 encountered in patients with ACS undergoing primary PCI. In contrast, no such associations were
19 observed for the precursor molecules histidine and urocanate.

20 Our study affirms that interindividual heterogeneity in the gut microbiome has important
21 implications for clinical risk stratification. Circulating ImP was strongly associated with both
22 MACE and all-cause mortality. These findings were consistent across patients with ACS and
23 CCS, geographies, subgroups, and follow-up periods. Notably, high ImP conferred incremental

1 gut microbiota-related risk above and beyond TMAO, an established gut-derived cardiovascular
2 risk factor. Patients with high levels of both metabolites had more than double the risk of MACE
3 after accounting for established risk factors. Indeed, elevation of both ImP and TMAO may
4 characterise a novel gut microbiota-related signature associated with increased cardiovascular
5 risk. Compared to established risk factors, ImP ranked among the strongest predictors of MACE
6 holding promise to refine risk stratification for personalised secondary prevention strategies.
7 Moreover, the absence of these findings for precursor molecules of ImP supports a potential
8 causal contribution of ImP to adverse outcomes. Currently, no cut-off value for elevated ImP is
9 established in the general population or in patients with CAD. In this study, we observed
10 particularly high risk of adverse events in patients in the highest ImP quartile (i.e., > 24 nM).

11 The biological actions of ImP and its complex interplay with cardiometabolic disease are the
12 subject of ongoing investigations. While more studies are needed to decipher its precise role in
13 atherogenesis and identify thresholds for optimal risk stratification, future treatment approaches⁸
14 may target microbial ImP production by inhibiting the microbial enzyme urocanate reductase to
15 modulate cardiometabolic risk.¹² The management of traditional cardiovascular risk factors has
16 significantly improved outcomes in the past decades, yet the current drug armamentarium lacks
17 strategies directed at modulating the microbiome and its metabolites. The potential integration of
18 the gut-heart axis⁴⁰ in future treatment approaches holds promise for more personalised and
19 effective secondary prevention.

20 In conclusion, our results identify circulating ImP as a novel biomarker of
21 cardiometabolic risk in patients with CAD and a potential therapeutic target for secondary
22 prevention.

1 **Strengths and limitations**

2 This study has several strengths. First, the rigorous multinational design including external
3 validation of the results in independent, well-characterised cohorts from different countries
4 confirmed the external validity of the findings and accounts for regional differences in treatment
5 practices, patient characteristics, and gut microbiota composition. Second, the SPUM-ACS
6 Biomarker Study comprises one of the largest and best characterised cohorts of patients with
7 ACS worldwide. Its prospective multicentre design together with a contemporary patient
8 population undergoing guideline-based treatment approaches grants for high quality of the
9 collected data in SPUM-ACS. Moreover, the central assessment of established biomarkers
10 including hs-TnT and hs-CRP in the core laboratory at the University Hospital Zurich, Zurich,
11 Switzerland, using a validated assay minimises the impact of analytical variability. Third,
12 assessment bias is prevented by external adjudication of clinical outcomes during follow-up by
13 an independent clinical event adjudication committee. Further, centralised blinded measurement
14 of ImP according to established protocols enhancing the technical validity of the measurements
15 and the comparability of the results. Finally, we also measured the precursor molecules of ImP to
16 inform hypothesis generation regarding potential causal relationships.

17 Our study also has limitations. First, there was a limited number of events at 1 year in the
18 validation cohort. However, the results were consistent across endpoints and at 3 years
19 considering a higher number of events. Second, for external validation of our results in patients
20 with ACS, we did not measure ImP in all patients of the German ACS cohort due to the use of a
21 case-cohort design. Nonetheless, the results of the case-cohort analysis are statistically robust and
22 provide valid approximations of full-cohort analyses. Third, a limited number of female patients
23 (less than a third across all cohorts) was included, and ethnicity was only available in the German

1 CCS cohort. Of note, we considered these variables in specific subgroup analyses and adjusted
2 models. Nevertheless, further studies are required to validate our findings in more ethnically
3 diverse cohorts, accounting for differences in dietary patterns across geographies and ethnicities.
4 In addition, due to limited availability of TMAO, analyses including this variable could only be
5 performed in a subcohort of patients. These analyses should therefore be interpreted with great
6 caution yet are consistent with the main results. Further, CMR data were only available in a small
7 subset of patients with ACS, precluding any definitive conclusions and should be interpreted as
8 hypothesis-generating. Next, the pooled analyses in *Figure 1* do not account for cohort-specific
9 data properties. However, similar results were observed in the individual cohorts. Finally, drug
10 dosages were not routinely recorded, and there was limited availability of gastrointestinal, hepatic
11 and host immune system-related parameters in the included cohorts.

12 **Conclusions**

13 Circulating levels of ImP are linked to cardiometabolic traits and independently predict adverse
14 outcomes in patients with ACS and CCS. ImP represents a novel biomarker of cardiometabolic
15 risk and a potential therapeutic target in patients with CAD. Our findings highlight the increasing
16 importance of non-traditional risk factors and may open new avenues for targeting the gut-heart
17 axis to address residual cardiovascular risk.

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5 **Author contributions**

6 F.A.W., A.H., U.L., F.B., and T.F.L. lead the conceptualisation of the study. Data curation was
7 carried out by F.A.W., P.W., F.K., K.R.B., B.E.S., and A.H.. F.A.W. and P.W. were involved in
8 the formal analysis of the data. F.A.W., P.W., and M.A.S were involved in the visualisation of
9 the results. F.A.W. wrote the original draft of the manuscript. F.A.W., A.H., U.L., F.B., and
10 T.F.L were primarily involved in funding acquisition. F.A.W. was primarily concerned with the
11 administration and coordination of the project. K.R.B. and F.B. supervised the measurements of
12 gut microbiota-derived metabolites. A.v.E. supervised the biomarker measurements in the
13 SPUM-ACS core laboratory. R.M. directed the analysis of the CMR images. T.F.L. supervised
14 the patient recruitment in SPUM-ACS and CLEVER-ACS in Switzerland. A.H. and U.L.
15 supervised the patient recruitment at Charité University Hospital, Germany. E.G. supervised the
16 patient recruitment at University Hospital Heidelberg, Germany. All the authors vouch for the
17 data and analyses reported. All authors revisited the work critically for important intellectual
18 content, were involved in reviewing and editing of the manuscript, approved the submission for
19 publication and agreed to be accountable for all aspects of the work.

20

References

1. Global burden of 369 diseases and injuries in 204 countries and territories, 1990-2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet* 2020;**396**:1204-1222. doi: 10.1016/s0140-6736(20)30925-9
2. Nabel EG, Braunwald E. A tale of coronary artery disease and myocardial infarction. *N Engl J Med* 2012;**366**:54-63. doi: 10.1056/NEJMra1112570
3. Libby P. The changing landscape of atherosclerosis. *Nature* 2021;**592**:524-533. doi: 10.1038/s41586-021-03392-8
4. Zaman S, Wasfy JH, Kapil V, Ziaeian B, Parsonage WA, Sriswasdi S, *et al.* The Lancet Commission on rethinking coronary artery disease: moving from ischaemia to atheroma. *Lancet* 2025. doi: 10.1016/s0140-6736(25)00055-8
5. Koh A, Molinaro A, Ståhlman M, Khan MT, Schmidt C, Mannerås-Holm L, *et al.* Microbially Produced Imidazole Propionate Impairs Insulin Signaling through mTORC1. *Cell* 2018;**175**:947-961.e917. doi: 10.1016/j.cell.2018.09.055
6. Venskutonytė R, Koh A, Stenström O, Khan MT, Lundqvist A, Akke M, *et al.* Structural characterization of the microbial enzyme urocanate reductase mediating imidazole propionate production. *Nat Commun* 2021;**12**:1347. doi: 10.1038/s41467-021-21548-y
7. Mastrangelo A, Robles-Vera I, Mañanes D, Galán M, Femenía-Muñina M, Redondo-Urzainqui A, *et al.* Imidazole propionate is a driver and therapeutic target in atherosclerosis. *Nature* 2025. doi: 10.1038/s41586-025-09263-w
8. Nageswaran V, Carreras A, Reinshagen L, Beck KR, Steinfeldt J, Henricsson M, *et al.* Gut Microbial Metabolite Imidazole Propionate Impairs Endothelial Cell Function and Promotes the Development of Atherosclerosis. *Arterioscler Thromb Vasc Biol* 2025. doi: 10.1161/atvbaha.124.322346
9. Koh A, Mannerås-Holm L, Yunn NO, Nilsson PM, Ryu SH, Molinaro A, *et al.* Microbial Imidazole Propionate Affects Responses to Metformin through p38 γ -Dependent Inhibitory AMPK Phosphorylation. *Cell Metab* 2020;**32**:643-653.e644. doi: 10.1016/j.cmet.2020.07.012
10. Molinaro A, Bel Lassen P, Henricsson M, Wu H, Adriouch S, Belda E, *et al.* Imidazole propionate is increased in diabetes and associated with dietary patterns and altered microbial ecology. *Nat Commun* 2020;**11**:5881. doi: 10.1038/s41467-020-19589-w
11. Sayols-Baixeras S, Dekkers KF, Baldanzi G, Jönsson D, Hammar U, Lin YT, *et al.* Streptococcus Species Abundance in the Gut Is Linked to Subclinical Coronary Atherosclerosis in 8973 Participants From the SCAPIS Cohort. *Circulation* 2023;**148**:459-472. doi: 10.1161/circulationaha.123.063914
12. Xu Q, Wang W, Li Y, Liu Y, Liu Y. Imidazole propionate in type 2 diabetes mellitus and cardiovascular diseases: a mini review. *Front Immunol* 2024;**15**:1454210. doi: 10.3389/fimmu.2024.1454210
13. Molinaro A, Nemet I, Bel Lassen P, Chakaroun R, Nielsen T, Aron-Wisnewsky J, *et al.* Microbially Produced Imidazole Propionate Is Associated With Heart Failure and Mortality. *JACC Heart Fail* 2023;**11**:810-821. doi: 10.1016/j.jchf.2023.03.008
14. Wenzl FA, Bruno F, Kraler S, Klingenberg R, Akhmedov A, Ministrini S, *et al.* Dipeptidyl peptidase 3 plasma levels predict cardiogenic shock and mortality in acute coronary syndromes. *Eur Heart J* 2023;**44**:3859-3871. doi: 10.1093/eurheartj/ehad545
15. Wenzl FA, Kraler S, Ambler G, Weston C, Herzog SA, Raber L, *et al.* Sex-specific evaluation and redevelopment of the GRACE score in non-ST-segment elevation acute coronary syndromes in populations from the UK and Switzerland: a multinational analysis with external cohort validation. *Lancet* 2022;**400**:744-756. doi: 10.1016/S0140-6736(22)01483-0
16. Davies A, Wenzl FA, Li XS, Winzap P, Obeid S, Klingenberg R, *et al.* Short and medium chain acylcarnitines as markers of outcome in diabetic and non-diabetic subjects with acute coronary syndromes. *Int J Cardiol* 2023;**389**:131261. doi: 10.1016/j.ijcard.2023.131261
17. Kraler S, Wenzl FA, Georgiopoulos G, Obeid S, Liberale L, von Eckardstein A, *et al.* Soluble lectin-like oxidized low-density lipoprotein receptor-1 predicts premature death in acute coronary syndromes. *Eur Heart J* 2022;**43**:1849-1860. doi: 10.1093/eurheartj/ehac143
18. Wenzl FA, Luscher TF. Application of a sex-specific GRACE score in practice - Authors' reply. *Lancet* 2023;**401**:23. doi: 10.1016/S0140-6736(22)02457-6
19. Kraler S, Wenzl FA, Vykoukal J, Fahrman JF, Shen MY, Chen DY, *et al.* Low-density lipoprotein electronegativity and risk of death after acute coronary syndromes: A case-cohort analysis. *Atherosclerosis* 2023;**376**:43-52. doi: 10.1016/j.atherosclerosis.2023.05.014
20. Bruno F, Adjibodou B, Obeid S, Kraler SC, Wenzl FA, Akhtar MM, *et al.* Occlusion of the infarct-related coronary artery presenting as an acute coronary syndrome with and without ST-elevation: impact of inflammation and

- 1 outcomes in a real-world prospective cohort. *Eur Heart J Qual Care Clin Outcomes* 2023;**9**:564-574. doi:
2 10.1093/ehjqcco/qcad027
- 3 21. Winzap PA, Kraler S, Obeid S, Wenzl FA, Templin C, Klingenberg R, *et al.* Initial systolic blood pressure
4 associates with systemic inflammation, myocardial injury, and outcomes in patients with acute coronary syndromes.
5 *Eur Heart J Acute Cardiovasc Care* 2023;**12**:437-450. doi: 10.1093/ehjacc/zuad047
- 6 22. Kraler S, Balbi C, Vdovenko D, Lapikova-Bryhinska T, Camici GG, Liberale L, *et al.* Circulating GDF11
7 exacerbates myocardial injury in mice and associates with increased infarct size in humans. *Cardiovasc Res* 2023.
8 doi: 10.1093/cvr/cvad153
- 9 23. Bruno F, Wenzl FA, De Filippo O, Kraler S, Giacobbe F, Roffi M, *et al.* Safety and effectiveness of
10 glycoprotein IIb/IIIa inhibitors in acute coronary syndromes: insights from the SPUM-ACS study. *Eur Heart J*
11 *Cardiovasc Pharmacother* 2024. doi: 10.1093/ehjcvp/pvae024
- 12 24. Narula S, Yusuf S, Chong M, Ramasundarahettige C, Rangarajan S, Bangdiwala SI, *et al.* Plasma ACE2
13 and risk of death or cardiometabolic diseases: a case-cohort analysis. *Lancet* 2020;**396**:968-976. doi: 10.1016/s0140-
14 6736(20)31964-4
- 15 25. O'Brien KM, Lawrence KG, Keil AP. The Case for Case-Cohort: An Applied Epidemiologist's Guide to
16 Reframing Case-Cohort Studies to Improve Usability and Flexibility. *Epidemiology* 2022;**33**:354-361. doi:
17 10.1097/ede.0000000000001469
- 18 26. König M, Joshi S, Leistner DM, Landmesser U, Sinning D, Steinhagen-Thiessen E, *et al.* Cohort profile:
19 role of lipoproteins in cardiovascular disease-the LipidCardio study. *BMJ Open* 2019;**9**:e030097. doi:
20 10.1136/bmjopen-2019-030097
- 21 27. Stähli BE, Klingenberg R, Heg D, Branca M, Manka R, Kapos I, *et al.* Mammalian Target of Rapamycin
22 Inhibition in Patients With ST-Segment Elevation Myocardial Infarction. *J Am Coll Cardiol* 2022;**80**:1802-1814.
23 doi: 10.1016/j.jacc.2022.08.747
- 24 28. Stamatelopoulos K, Mueller-Hennessen M, Georgiopoulos G, Lopez-Ayala P, Sachse M, Vlachogiannis NI,
25 *et al.* Cathepsin S Levels and Survival Among Patients With Non-ST-Segment Elevation Acute Coronary
26 Syndromes. *J Am Coll Cardiol* 2022;**80**:998-1010. doi: 10.1016/j.jacc.2022.05.055
- 27 29. Qiu X, Yang J, Hu X, Li J, Zhao M, Ren F, *et al.* Association between hearing ability and cortical
28 morphology in the elderly: multiparametric mapping, cognitive relevance, and neurobiological underpinnings.
29 *EBioMedicine* 2024;**104**:105160. doi: 10.1016/j.ebiom.2024.105160
- 30 30. Stamatelopoulos K, Mueller-Hennessen M, Georgiopoulos G, Sachse M, Boeddinghaus J, Sopova K, *et al.*
31 Amyloid-beta (1-40) and Mortality in Patients With Non-ST-Segment Elevation Acute Coronary Syndrome: A
32 Cohort Study. *Ann Intern Med* 2018;**168**:855-865. doi: 10.7326/M17-1540
- 33 31. Wenzl FA, Wang P, Arrigo M, Parencia J, Jones DJL, Bruno F, *et al.* Proenkephalin Improves Cardio-Renal
34 Risk Prediction in Acute Coronary Syndromes: The KID-ACS Score. *Eur Heart J* 2024. doi:
35 10.1093/eurheartj/ehae602
- 36 32. Grund B, Sabin C. Analysis of biomarker data: logs, odds ratios, and receiver operating characteristic
37 curves. *Curr Opin HIV AIDS* 2010;**5**:473-479. doi: 10.1097/COH.0b013e32833ed742
- 38 33. Fox KA, Fitzgerald G, Puymirat E, Huang W, Carruthers K, Simon T, *et al.* Should patients with acute
39 coronary disease be stratified for management according to their risk? Derivation, external validation and outcomes
40 using the updated GRACE risk score. *BMJ Open* 2014;**4**:e004425. doi: 10.1136/bmjopen-2013-004425
- 41 34. Li XS, Obeid S, Klingenberg R, Gencer B, Mach F, Raber L, *et al.* Gut microbiota-dependent
42 trimethylamine N-oxide in acute coronary syndromes: a prognostic marker for incident cardiovascular events beyond
43 traditional risk factors. *Eur Heart J* 2017;**38**:814-824. doi: 10.1093/eurheartj/ehw582
- 44 35. von Elm E, Altman DG, Egger M, Pocock SJ, Gøtzsche PC, Vandenbroucke JP, *et al.* The Strengthening
45 the Reporting of Observational Studies in Epidemiology (STROBE) statement: guidelines for reporting observational
46 studies. *Lancet* 2007;**370**:1453-1457. doi: 10.1016/S0140-6736(07)61602-X
- 47 36. Seldin MM, Meng Y, Qi H, Zhu W, Wang Z, Hazen SL, *et al.* Trimethylamine N-Oxide Promotes Vascular
48 Inflammation Through Signaling of Mitogen-Activated Protein Kinase and Nuclear Factor- κ B. *J Am Heart Assoc*
49 2016;**5**. doi: 10.1161/jaha.115.002767
- 50 37. Witkowski M, Witkowski M, Friebel J, Buffa JA, Li XS, Wang Z, *et al.* Vascular endothelial tissue factor
51 contributes to trimethylamine N-oxide-enhanced arterial thrombosis. *Cardiovasc Res* 2022;**118**:2367-2384. doi:
52 10.1093/cvr/cvab263
- 53 38. Zhang X, Li Y, Yang P, Liu X, Lu L, Chen Y, *et al.* Trimethylamine-N-Oxide Promotes Vascular
54 Calcification Through Activation of NLRP3 (Nucleotide-Binding Domain, Leucine-Rich-Containing Family, Pyrin
55 Domain-Containing-3) Inflammasome and NF- κ B (Nuclear Factor κ B) Signals. *Arterioscler Thromb Vasc Biol*
56 2020;**40**:751-765. doi: 10.1161/atvbaha.119.313414

- 1 39. Galli M, Niccoli G, De Maria G, Brugaletta S, Montone RA, Vergallo R, *et al.* Coronary microvascular
2 obstruction and dysfunction in patients with acute myocardial infarction. *Nat Rev Cardiol* 2024;**21**:283-298. doi:
3 10.1038/s41569-023-00953-4
4 40. Tang WH, Hazen SL. The Gut Microbiome and Its Role in Cardiovascular Diseases. *Circulation*
5 2017;**135**:1008-1010. doi: 10.1161/circulationaha.116.024251
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Legends

Figure 1: Circulating imidazole propionate is associated with extent of coronary artery disease and metabolic traits. Circulating imidazole propionate (ImP) levels according to extent of coronary artery disease (A) and metabolic traits (B). Data are represented as boxplots and correlation plots. Medians with interquartile ranges are displayed on top of each box. Spearman partial correlation coefficients and P-values were calculated using partial correlation adjusted for Model 1: age and sex, Model 2: Model 1 plus diabetes status, Model 3: Model 2 plus hypercholesterolaemia. False discovery rate (FDR)-adjusted using Benjamini–Hochberg correction $*P < 0.05$, $**P < 0.01$. Values in the group with 0 affected vessels refer to the reference population without coronary artery disease. Results are based on pooled data from the Swiss ACS cohort and the German CCS cohort. See also *Supplementary data online, Table S13*.

Figure 2: Imidazole propionate associates with coronary microvascular obstruction. (A) Coronary microvascular obstruction (MVO) in patients undergoing primary percutaneous coronary intervention (PCI) according to ImP (Q1: 0–9.4 nM; Q2: 9.4–14.3 nM; Q3: 14.3–24.4 nM; Q4: > 24.4 nM). Bars are mean values. (B) Correlation of ImP and coronary MVO. Spearman partial correlation coefficients and P-values were calculated using partial correlations adjusted for Model 1: age, sex, and diabetes status, Model 2: Model 1 plus hypertension, low density lipoprotein cholesterol, and glucose, Model 3: Model 2 plus preprocedural coronary flow based on Thrombolysis in Myocardial Infarction (TIMI) classification and culprit vessel. False discovery rate (FDR)-adjusted using Benjamini–Hochberg correction $*P < 0.05$, $**P < 0.01$. Results in all figure panels are based on data from the Swiss CMR cohort. See also *Supplementary data online, Table S14 and Figure S12*.

Figure 3: Imidazole propionate predicts cardiovascular outcomes in patients with acute and chronic coronary syndrome. (A) Cumulative event curves for major adverse cardiovascular events (MACE), defined as a composite of death, nonfatal myocardial infarction, and nonfatal stroke, of patients presenting with acute (left) and chronic (right) coronary syndrome stratified by circulating imidazole propionate (ImP) level (Q1: 0–9.4 nM; Q2: 9.4–14.3 nM; Q3: 14.3–24.4 nM; Q4: > 24.4 nM). (B) Restricted cubic spline plot showing the adjusted hazard ratio (HR) for MACE at 1 year and at 3 years according to ImP level in patients with acute (left; n=3966) and

1 chronic (right; n=385) coronary syndromes, respectively. The solid line indicates the predicted
2 HR with the colour band signifying the corresponding 95% CI. Three knots were fixed at the
3 10th, 50th, and 90th percentile of the ImP distribution. Note that HRs are shown in log scale (left
4 Y axis) and as untransformed values (right Y axis). The dashed horizontal line indicates the
5 reference. Dashed vertical lines mark quartile cut-offs (Q1: 0–9.4 nM; Q2: 9.4–14.3 nM; Q3:
6 14.3–24.4 nM; Q4: > 24.4 nM). (C) Crude and adjusted HRs for MACE at 1 year and at 3 years
7 in patients with acute (left) and chronic (right) coronary syndromes, respectively. Cox
8 proportional hazards regression models were controlled for established risk factors and precursor
9 molecules of ImP in a stepwise fashion. Squares indicate point estimates of the HR per log₂
10 increase and line lengths equal corresponding 95% CIs. Plots in A) and B) are based on data from
11 the Swiss ACS cohort and the German CCS cohort. Patient numbers and event counts for C) are
12 summarised in the *Supplementary data online, Table S15*.

13
14

15 **Figure 4: Imidazole propionate predicts adverse cardiovascular outcomes beyond**
16 **trimethylamine N-oxide.**

17 (A) Cumulative event curves for major adverse cardiovascular events (MACE), defined as a
18 composite of death, nonfatal myocardial infarction, and nonfatal stroke in patients presenting
19 with acute coronary syndrome (ACS) and chronic coronary syndromes (CCS) according to joint
20 groups of high (\geq 75th percentile; Q4: > 24.4 nM) vs. low (< 75th percentile; Q1-Q3: 0–24.4 nM)
21 levels of imidazole propionate (ImP) and trimethylamine N-oxide (TMAO). (B) Multivariable-
22 adjusted hazard ratio (HR) and 95% confidence interval (CI) for MACE at 1 year in patients with
23 ACS and at 3 years in patients with CCS, according to ImP quartile (Q1: 0–9.4 nM; Q2: 9.4–14.3
24 nM; Q3: 14.3–24.4 nM; Q4: > 24.4 nM). Squares indicate point estimates and line lengths equal
25 corresponding 95% CIs. (C) Feature importance for the prediction of MACE measured by partial
26 Wald χ^2 minus the degrees of freedom. Results are based on 1281 patients from the Swiss ACS
27 cohort and 385 patients from the German CCS cohort in the fully adjusted model.

28

	Swiss ACS cohort (n = 4318)	German ACS cohort (n = 1428)	German CCS cohort (n = 577)
Age (years)	63 (54–73)	68 (57–76)	76 (67–81)
Female	881/4318 (20.4%)	440/1428 (30.8%)	124/577 (21.5%)
BMI (kg/m ²)	26.6 (24.2–29.4)	27.1 (25.0–30.5)*	26.9 (24.5–30.0)
Body surface area (m ²)	1.9 (1.8–2.1)	..	2.0 (1.8–2.1)
Heart rate (bpm)	75 (66–86)	74 (65–85)	..
Systolic blood pressure (mm Hg)	128 (112–143)	150 (137–166)	134 (121–149)
Signs and symptoms of heart failure (ACS: Killip class, CCS: NYHA class)			
I	3682/4204 (87.6%)	1365/1428 (95.6%)	93/555 (16.8%)
II	343/4204 (8.2%)	50/1428 (3.5%)	294/555 (53.0%)
III	85/4204 (2.0%)	11/1428 (0.8%)	166/555 (29.9%)
IV	94/4204 (2.2%)	2/1428 (0.1%)	2/555 (0.4%)
Smoking	1649/4254 (38.8%)	449/1371 (32.8%)	98/577 (17.0%)
Medical history			
Diabetes	753/4318 (17.4%)	319/1414 (22.6%)	178/577 (30.8%)
Prediabetes [#]	630/2153 (29.3%)	194/670 (29.0%)	168/577 (29.1%)
Hypertension	2413/4316 (55.9%)	1099/1417 (77.6%)	477/577 (82.7%)
Hyperlipoproteinemia	2757/4317 (63.9%)	838/1369 (61.2%)	378/577 (65.5%)
Myocardial infarction	524/4310 (12.2%)	344/1402 (24.5%)	405/576 (70.3%)
Family history of CAD	1056/4259 (24.8%)	491/1352 (36.3%)	161/478 (33.7%)
Prior PCI	644/4317 (14.9%)	498/1400 (35.6%)	..
Prior CABG	169/4318 (3.9%)	157/1399 (11.2%)	62/577 (10.7%)
Cerebrovascular disease	160/4318 (3.7%)	..	74/577 (12.8%)
Peripheral artery disease	251/4318 (5.8%)	..	30/577 (5.2%)
Heart failure	54/4316 (1.3%)	294/1348 (21.8%)	..
Chronic liver disease	25/4318 (0.6%)	3/139 (2.2%)*	12/577 (2.1%)
Dialysis	21/4318 (0.5%)
Clinical chemistry and haematology			
Leukocyte count (G/L)	9.6 (7.4–12.2)	7.6 (5.6–13.3)*	7.4 (6.0–8.9)
hs-CRP (mg/L)	3 (1–7)	4 (2–12)	2 (1–7)
Total cholesterol (mg/dL)	190 (159–220)	189 (172–206)*	153 (129–187)
LDL-C (mg/dL)	120 (92–149)	91 (79–106)*	85 (65–115)
HDL-C (mg/dL)	44 (36–53)	47 (43–50)*	46 (38–57)
Lipoprotein (a) (mg/dL)	6.4 (2.5–21.8)	..	8.3 (2.0–34.9)
Triglycerides (mg/dL)	88 (60–136)	148 (109–185)*	121 (91–176)

HbA1c (%)	5.8 (5.5–6.3)	5.8 (5.5–6.3)	5.8 (5.5–6.3)
Glucose (mmol/L)	6.4 (5.6–7.9)	6.6 (5.7–8.2)	5.9 (5.2–7.7)
NT-proBNP (ng/L)	334 (108–1158)	586 (164–2440)	427 (134–1238)
hs-TnT (ng/L)	196 (59–642)	46 (9–220)	15 (9–30)
Creatinine (mg/dL)	0.9 (0.7–1.0)	0.9 (0.8–1.1)*	1.0 (0.8–1.2)
eGFR (mL/min/1.73m ²)	93 (77–103)	86 (76–96)	73 (59–90)
ImP metabolism			
ImP (nM)	14 (9–24)	13 (9–20)*	13 (8–26)
Urocanate (nM)	44 (30–66)	128 (81–203)*	47 (33–84)
Histidine (μM)	66 (57–76)	88 (68–103)*	62 (53–70)
Baseline medication			
Aspirin	1230/2779 (44.3%)	33/139 (23.7%)*	381/577 (66.0%)
P2Y12 inhibitor	321/1915 (16.8%)	..	192/577 (33.3%)
Beta-blocker	978/2779 (35.2%)	22/139 (15.8%)*	371/577 (64.3%)
ACE inhibitor/ARB	1475/2759 (53.5%)	100/139 (71.9%)*	447/577 (77.5%)
Vitamin K antagonist/DOAC	171/2777 (6.2%)	19/139 (13.7%)*	148/577 (25.6%)
Diuretic	674/2773 (24.3%)	50/139 (36.0%)*	247/577 (42.8%)
Insulin	214/2777 (7.7%)	35/139 (25.2%)*	57/577 (9.9%)
Oral antidiabetics	503/2777 (18.1%)	4/139 (2.9%)*	108/577 (18.7%)
Immunosuppressive drugs	119/2777 (4.3%)
PPI	643/2777 (23.2%)	32/139 (23.0%)*	247/577 (42.8%)

1

2 **Table 1. Baseline characteristics of patients with acute or chronic coronary syndromes**

3 Categorical data are shown as numbers and percentages (%). Continuous data are presented as
4 median and interquartile range (IQR). ACE, angiotensin-converting enzyme; ACS, acute coronary
5 syndrome; ARB, angiotensin II receptor blocker; BMI, body mass index; CABG, coronary artery
6 bypass grafting; CAD, coronary artery disease; CCS, chronic coronary syndrome; DOAC, direct
7 oral anticoagulant; eGFR, estimated glomerular filtration rate; HbA1c, hemoglobin A1c; HDL-C,
8 high-density lipoprotein cholesterol; hs-CRP, high-sensitivity C-reactive protein; hs-TnT, high-
9 sensitivity troponin T; ImP, imidazole propionate; LDL-C, low-density lipoprotein cholesterol; NT-
10 proBNP, N-terminal pro B-type natriuretic peptide; NYHA, New York Heart Association; PCI,
11 percutaneous coronary intervention; PPI, proton pump inhibitor; STEMI, ST-segment elevation
12 myocardial infarction. *Values refer to the subcohort #defined as HbA1c of 5.7–6.4% in individuals
13 without a history of diabetes.

14

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	Swiss ACS cohort (n = 4318)	German ACS cohort (n = 1428)	German CCS cohort (n = 577)
Revascularization strategy during baseline visit			
PCI	3949/4318 (91.5%)	843/1283 (65.7%)	350/577 (60.7%)
CABG	127/4318 (2.9%)
Symptom-to-door time (minutes)	180 (105–420)	480 (120–2160)	..
No. of affected vessels†			
One	3371/4095 (82.3%)	209/1283 (16.3%)	135/577 (23.4%)
Two	563/4095 (13.7%)	237/1283 (18.5%)	194/577 (33.6%)
Three	161/4095 (3.9%)	724/1283 (56.4%)	248/577 (43.0%)
Left ventricular ejection fraction (%)			
> 50	1380/2725 (50.6%)	420/1361 (30.9%)	437/577 (75.7%)
50–40	858/2725 (31.5%)	418/1361 (30.7%)	81/577 (14.0%)
40–30	314/2725 (11.5%)	349/1361 (25.6%)	38/577 (6.6%)
< 30	173/2725 (6.3%)	174/1361 (12.8%)	21/577 (3.6%)
Discharge medication			
Aspirin	4233/4273 (99.1%)	1259/1384 (91.0%)	..
P2Y12 inhibitor	4070/4163 (97.8%)	989/1393 (71.0%)	..
Beta-blocker	3339/4269 (78.2%)	1227/1393 (88.1%)	..
ACE inhibitor/ARB	3749/4271 (87.8%)	1218/1393 (87.4%)	..
Vitamin K antagonist/DOAC	303/4318 (7.0%)	147/1391 (10.6%)	..
Diuretic	997/4272 (23.3%)
Insulin	277/4270 (6.5%)	108/1392 (7.8%)	..
Oral antidiabetics	503/4270 (11.8%)	175/1392 (12.6%)	..
Immunosuppressive drugs	112/4272 (2.6%)
PPI	1258/4272 (29.4%)	40/139 (15.8%)*	..
Outcomes§			
MACE	314/4318 (7.3%)	117/1428 (8.2%)	164/577 (28.42%)
Mortality	148/4318 (3.4%)	75/1428 (5.3%)	47/577 (8.15%)

2

3 **Table 2. Treatment characteristics of patients with acute or chronic coronary syndromes**

4 Categorical data are shown as numbers and percentages (%). Continuous data are presented as
5 median and interquartile range (IQR). ACE, angiotensin-converting enzyme; ACS, acute coronary
6 syndrome; ARB, angiotensin II receptor blocker; CABG, coronary artery bypass grafting; CCS,
7 chronic coronary syndrome; DOAC, direct oral anticoagulant; MACE, major adverse
8 cardiovascular events; PCI, percutaneous coronary intervention; PPI, proton pump inhibitor.

9 †Refers to vessels with significant coronary stenosis in Heidelberg II-ACS and LipidCardio.

10 §Refers to 1-year outcomes for ACS and to 3-year outcomes for CCS included in the analyses.

11 *Values refer to the subcohort

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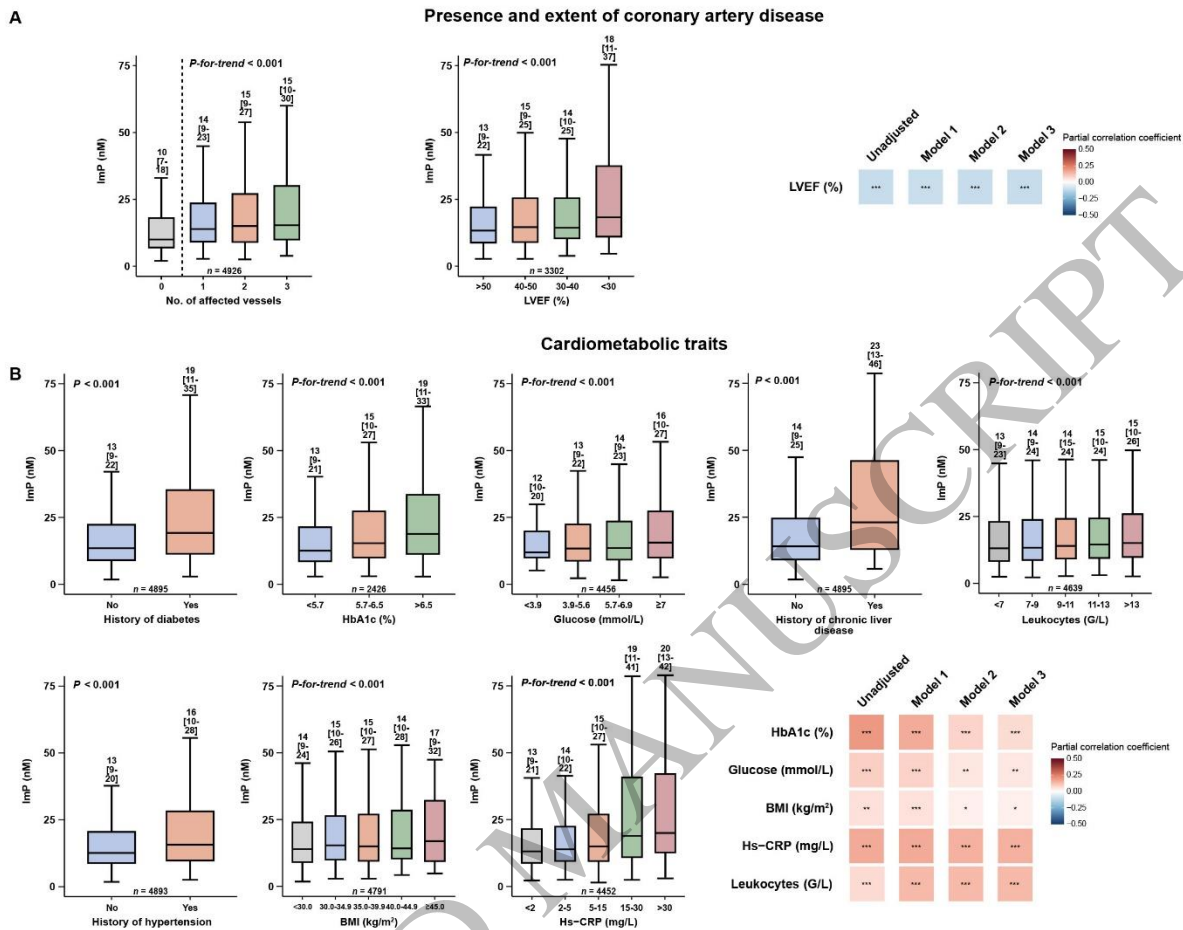


Figure 1
160x123 mm (x DPI)

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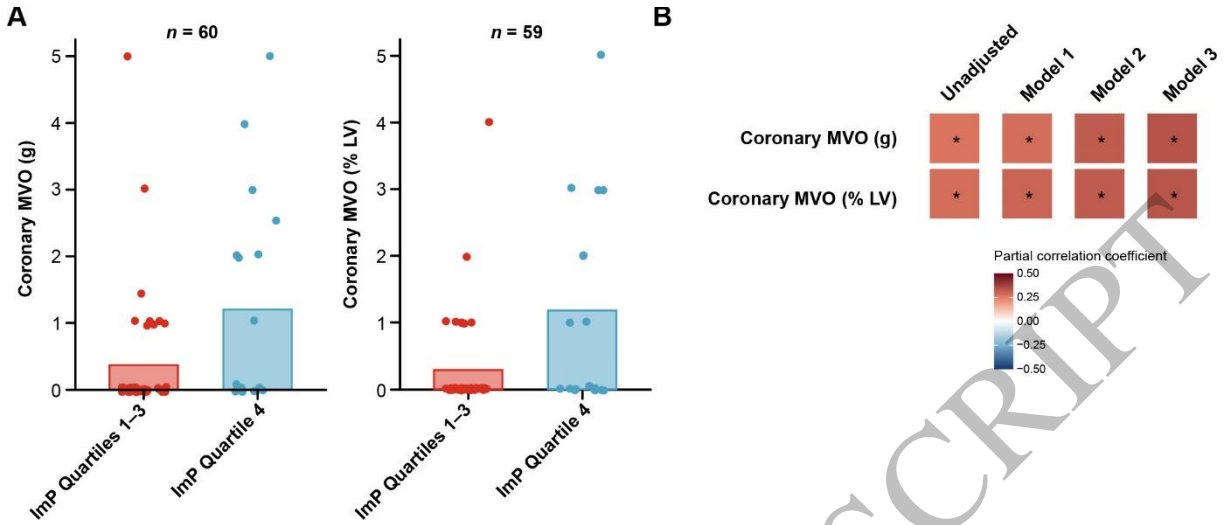


Figure 2
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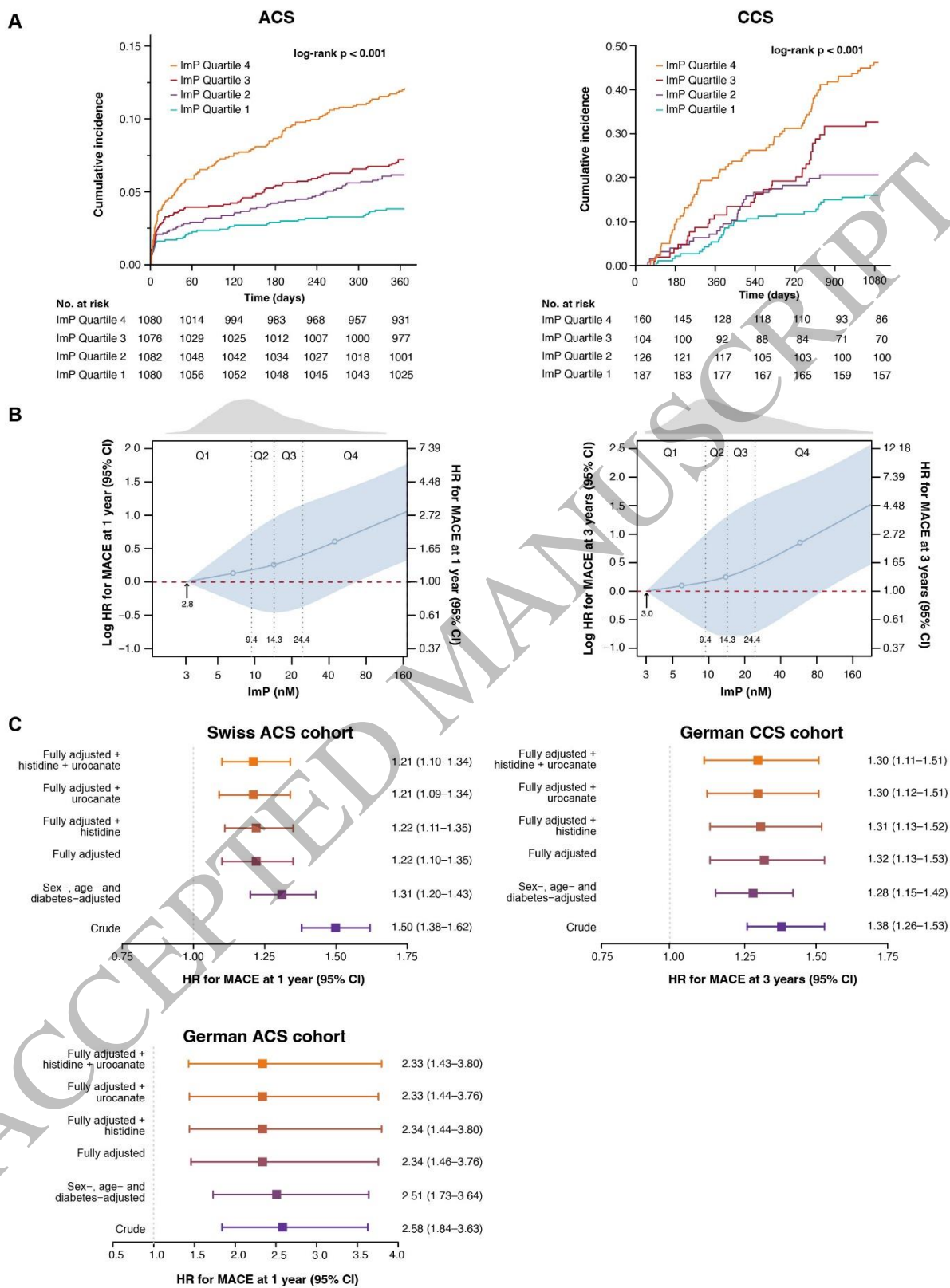


Figure 3
160x214 mm (x DPI)

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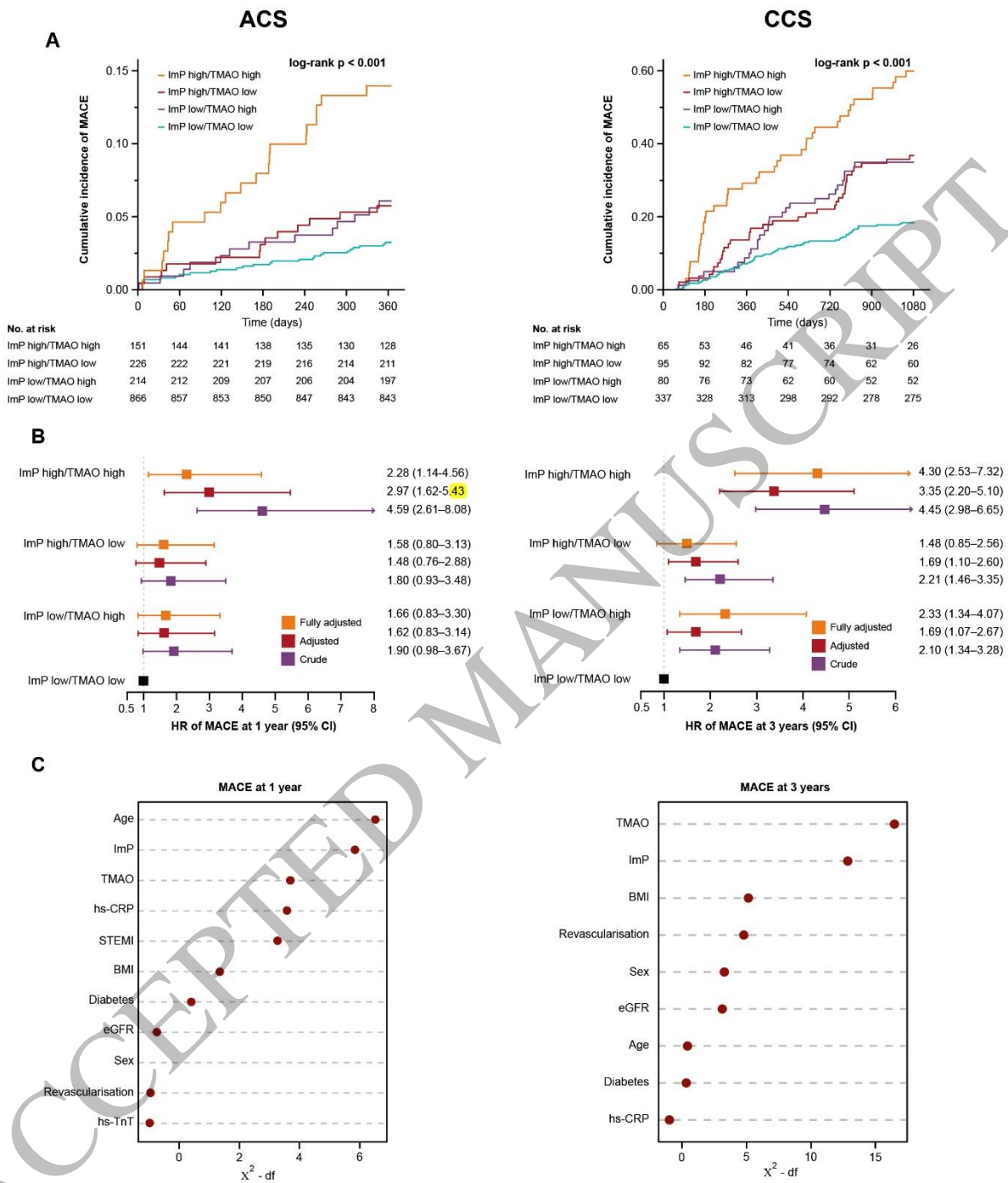


Figure 4
160x185 mm (x DPI)

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Structured Graphical Abstract

Key Question: Experimental evidence shows that gut microbiota-derived imidazole propionate (ImP) is mechanistically linked to atherosclerotic plaque formation and abnormal glucose metabolism. This study investigated the relationship between ImP and adverse outcomes in patients with coronary artery disease (CAD).

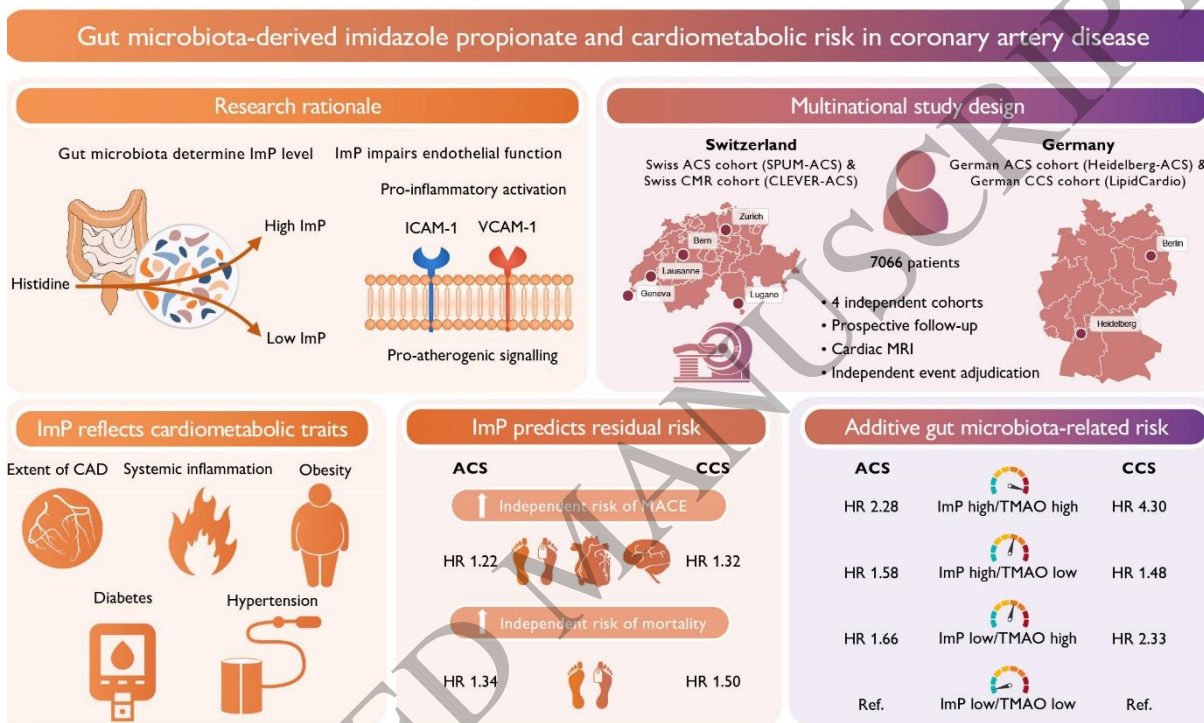
Key Finding: Circulating levels of ImP were linked to cardiometabolic characteristics and independently predicted major adverse cardiovascular events both in patients with acute coronary syndrome and in patients with chronic coronary syndrome.

Take Home Message: Gut microbiota-derived ImP is a novel marker of cardiometabolic risk in patients with CAD and represents a potential therapeutic target.

Legend: Gut microbiota-derived imidazole propionate (ImP) was identified as a novel marker of cardiometabolic risk in patients with coronary artery disease (CAD). Circulating ImP levels were measured in 4 independent cohorts of patients with acute (ACS) and chronic coronary syndrome (CCS) from Switzerland and Germany. ImP results from microbial histidine metabolism and has been mechanistically linked to inflammatory activation of myeloid and endothelial cells driving atherogenesis. Circulating levels of ImP were associated with cardiometabolic traits and independently predicted major adverse cardiovascular events (MACE) in patients with ACS and CCS. ImP added to the gut microbiota-related risk beyond the established microbiota-derived metabolite trimethylamine N-oxide (TMAO). Joint groups of high (\geq 75th percentile; ImP \geq 24.4 nM) vs. low ($<$ 75th percentile; ImP $<$ 24.4 nM) circulating levels of ImP and TMAO were analysed. Values refer to hazard ratios (HR) in the fully adjusted model.

1 CMR, cardiac magnetic resonance; ICAM-1, intercellular adhesion molecule 1; MRI, magnetic
 2 resonance imaging; VCAM-1, vascular cell adhesion molecule 1.

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Graphical Abstract

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 14 not require permission to reuse.